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5 ***Committee 1 Task Group Report***

6
7 **Low-dose Extrapolation of Radiation-Related Cancer Risk**

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PREFACE

Following its meeting in Oxford, UK, in 1997, Committee 1 of ICRP (the International Commission on Radiological Protection) proposed a Task Group to prepare a report on low-dose extrapolation of radiation-related cancer risk estimates based largely on higher-dose epidemiological data, and the possible implications for radiological protection. The Commission accepted this recommendation and established a Task Group, which began its work in April, 1998.

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EXECUTIVE SUMMARY

5 The present report, of a task group of ICRP Committee 1, considers the evidence
6 relating to cancer risk associated with exposure to low doses of low-LET radiation, and
7 particularly doses below current recommended limits for protection of radiation workers
8 and the general public. The focus is on evidence regarding linearity of dose response for
9 all cancers considered as a group, but not necessarily individually, at low doses (the so-
10 called linear, no-threshold (LNT) hypothesis), and the possibility of a universal threshold
11 dose below which there is no risk of radiation-related cancer. According to the LNT
12 hypothesis, the same number of radiation-related cancers would be predicted in a
13 population of a given size exposed to a certain small average radiation dose and in an
14 otherwise similar population many times times larger and exposed to a proportionally
15 smaller average dose. According to the threshold hypothesis, the radiation-related risk in
the larger population would be zero if its average dose were sufficiently small.

16 The present document has been preceded by other, recent reports, notably those of
17 the United Nations Scientific Committee on the Effects of Atomic Radiation and the U.S.
18 National Council of Radiation Protection and Measurements. These reports recommended
19 that radiation protection continue to be guided by the LNT hypothesis. The task group
20 concurs with those recommendations.

21 The report is organized by scientific discipline, beginning with epidemiological
22 studies of exposed human populations (Chapter 2). Epidemiological studies offer the most
23 directly relevant information for risk-based radiation protection. The major scientific
24 issues, as illustrated by the example of cancer incidence from all solid tumors combined
25 in the Life Span Study (LSS) population of atomic bomb survivors, are (1) establishment
26 of the existence of a dose-related risk in this population, (2) modeling radiation-related
27 risk as a statistically uncertain parametric function of dose, modified by other factors such
28 as sex, exposure age, attained age, and time following exposure, (3) extrapolation of
29 estimated risk to other potentially exposed populations, with possibly different baseline
30 cancer rates, (4) projection of the risk in the population to the end of its natural life, and
31 (5) extrapolation of risk estimates from moderate-to-high dose levels of acute exposure,
32 characteristic of the most informative atomic bomb survivor data, to the far more
33 common low-dose and/or protracted exposures that occur in occupational and general
34 settings. Consideration of each of these issues leads to more refined risk estimates but,

1 because information about each is uncertain, the overall uncertainty of the improved
2 estimates is increased. There is some evidence of increased cancer risk associated with
3 exposures on the order of 10 mGy which will be discussed in the report, and other
4 evidence placing an upper limit on the value of any universal threshold that might exist.
5 Also, the risk of mortality and morbidity from all solid cancers combined is proportional
6 to radiation dose down to about 100 mGy, below which statistical variation in baseline
7 risk, as well as small and uncontrollable biases, tend to obscure evidence concerning
8 radiation-related risk. Extrapolation of risk estimates based on observations at moderate
9 to high doses continues to be the primary basis for estimation of radiation-related risk at
10 low doses and dose rates, for example at the present recommended limit for members of
11 the public of 1mGy per year from non-medical man-made sources.

12 The fundamental role of radiation-induced DNA damage in the induction of
13 mutations and chromosome aberrations and the apparent critical involvement of
14 aberrations and mutations in the pathogenesis of cancer provides a framework for the
15 analysis of risks at low radiation doses and low dose rate exposures (Chapter 3). A
16 characteristic type of damage produced by ionizing radiation (IR) involves multiple
17 lesions within close spatial proximity. Such clustered damage can be induced even by a
18 single radiation track through a cell. Although cells have a vast array of damage response
19 mechanisms that facilitate the repair of DNA damage and the removal of damaged cells,
20 these mechanisms are not fool-proof, and emerging evidence suggests that closely spaced
21 lesions can compromise the repair machinery. Also, while many of the cells containing
22 such radiation-induced damage may be eliminated by damage response pathways
23 involving cell cycle checkpoint control and apoptotic pathways, it is clear from analysis
24 of cytogenetics and mutagenesis that damaged or altered cells are capable of escaping
25 these pathways and propagating.

26 Cellular consequences of radiation-induced damage (Chapter 4) include
27 chromosome aberrations and somatic cell mutations. The processing and misrepair of
28 radiation-induced DSBs, particularly complex forms, are responsible for
29 chromosome/gene alterations that manifest as chromosome aberrations and mutations.
30 Current understanding of mechanisms and quantitative data on dose and time-dose
31 relationships support a linear dose response at low doses (i.e., LNT) for total cancer risk.
32 Considered as a whole, the emerging results with regard to radiation-related adaptive
33 response, genomic instability, and bystander effects suggest that the risk of low level

1 exposure to ionizing radiation is uncertain, and a simple extrapolation from high dose
2 effects may not be wholly justified in all instances. However, a better understanding of
3 the mechanisms for these phenomena, the extent to which they are active *in vivo*, and how
4 they are interrelated is needed before they can be evaluated as factors to be included in
5 the estimation of potential risk to the human population of exposure to low levels of
6 ionizing radiation.

7 Experimental approaches using animal models (Chapter 5) are well suited to
8 precise control of radiation dose and dose rate, as well as genetic background and other
9 possible modifiers of dose response, and can facilitate precise determination of biological
10 outcomes. Recent studies using newly developed animal models, cellular, cytogenetic and
11 molecular data for acute myelogenous leukemia (AML), intestinal tumors, and mammary
12 tumors, and cytogenetic and molecular studies on the induction of AML and mammary
13 cancer support the view that the essential radiation-associated events in the tumorigenic
14 process are predominantly early events involving DNA losses targeting specific genomic
15 regions harboring critical genes. As such, the response for early initiating events is likely
16 to correspond to that for the induction of cytogenetic damage. On this basis, mechanistic
17 arguments support a linear response in the low dose region, i.e., the process should be
18 independent of dose rate because interactions between different electron tracks should be
19 rare. Quantitative analyses of dose responses for tumorigenesis and for life shortening in
20 laboratory animals also support this prediction. These studies also support a dose and
21 dose rate effectiveness factor (DDREF), for reduction of estimated risk per unit dose
22 based on acute, high-dose data, in the range of about 2 when data are extrapolated to low
23 doses from effects induced by doses in the range of 2-3 Gy. Extrapolation of results from
24 less than 1 Gy would result in lower DDREF values.

25 Chapter 6 presents a formal exercise in quantitative uncertainty analysis, in which
26 the different uncertain components (as identified in Chapter 2) of estimated cancer risk
27 associated with low-dose, low-LET radiation exposure to a non-Japanese population, in
28 this case that represented by the U.S. National Cancer Institute's SEER registry, are
29 combined. Attention is paid to the resulting uncertainty distribution for excess relative
30 risk per Gy (ERR/Gy), with and without allowing for the uncertain possibility of a
31 universal low-dose threshold, below which there would be no radiation-related risk. In the
32 example, which involves risk from all cancers combined including leukemia, except for
33 non-melanoma skin cancer, the major sources of uncertainty are statistical variation in the

1 estimated ERR at 1 Gy for the atomic bomb survivors population, subjective uncertainty
2 (informed by experimental and epidemiological data) about the DDREF to be applied at
3 low doses and dose rates, and the postulated uncertainty concerning the existence of a
4 universal threshold at some dose above that for which the calculation was being made.
5 Unless the existence of a threshold was assumed to be virtually certain, the effect of
6 introducing the uncertain possibility of a threshold was equivalent to that of an uncertain
7 increase in the value of DDREF, i.e., merely a variation on the result obtained by ignoring
8 the possibility of a threshold.

9 The conclusions of this report are given in Chapter 7. While existence of a low-
10 dose threshold does not seem unlikely for radiation-related cancers of certain tissues, and
11 cannot be ruled out for all cancers as a group, the evidence as a whole does not favor the
12 existence of a universal threshold, and there seems to be no particular reason to factor the
13 possibility of a threshold into risk calculations for purposes of radiation protection. The
14 LNT hypothesis, combined with an uncertain DDREF for extrapolation from high doses,
15 remains a prudent basis for radiation protection at low doses and low dose rates.

1. INTRODUCTION

The purpose of the present report is to summarize scientific evidence relevant to the quantification of cancer risk associated with radiation exposure at (effective) doses of interest for radiation protection, particularly doses below current recommended limits for protection of radiation workers (e.g., 20 mSv per year) and the general public (e.g., 1 mSv per year). (As a rough rule of thumb, effective doses on the order of 1 Sv, 100 mSv, 10 mSv, 1 mSv, and 0.1 mSv will be called “moderately high”, “moderate”, “low”, “very low”, and “extremely low”, respectively, in this report.)

Ionizing radiation exposure is an established cancer risk factor. Compared to other common environmental carcinogens, it is relatively easy to determine organ-specific radiation dose and, as a result, radiation dose-response relationships tend to be highly quantified. Nevertheless, there can be considerable uncertainty about questions of radiation-related cancer risk as they apply to risk protection and public policy, and the interpretations of interested parties can differ radically. A major reason for disagreement is that public and regulatory concern often is focused on exposures at radiation doses far lower than those at which useful information about cancer risk can be obtained directly, that is, than can be obtained by studying populations with such exposures. Thus, risk estimates promulgated by expert committees, for example, are usually based upon epidemiological dose-response data obtained at doses ranging up to 0.2 Gy, 0.5 Gy, 1 Gy, or higher, and the resulting estimates are then extrapolated, with appropriate caveats, to lower doses. The extrapolation rules are based in part upon epidemiological observations, such as the degree of curvature of fitted linear-quadratic dose response models for leukemia and solid cancer morbidity among atomic bomb survivors, and on models derived from experimental systems.

The discussion in the present report is concerned ultimately with biological effects of ionizing radiations of low linear energy transfer (low LET), such as photons (gamma rays and X rays) and electrons (beta particles) of various energies, as contrasted with high-LET radiations such as neutrons and alpha particles. However, some biological effects that have been observed mainly in connection with high-LET exposure are clearly relevant to questions of cancer risk at low levels of low-LET radiation.

Currently, the ICRP radiation protection philosophy is based on the so-called linear, non-threshold (LNT) hypothesis, according to which, at low doses (on the order of 100 mGy or less) and dose rates (less than 6 mGy/hour averaged over the first few hours)

1 (UNSCEAR 1993, EPA 1999) total radiation-related cancer risk is proportional to dose.
2 The hypothesis is not universally accepted as biological truth, but rather, because we do
3 not actually know what level of risk is associated with very low-dose exposure, is
4 considered by many as a prudent rule of thumb for public policy aimed at avoiding
5 unnecessary risk from exposure.

6 A logical consequence of the hypothesis is that, at a sufficiently low dose D ,
7 exposure of N people to average dose D would result in the same number of radiation-
8 related cancers as exposure of $k \cdot N$ people to average dose D / k , for arbitrary $k > 1$. This
9 logical consequence can be used to justify the concept of “collective dose”, that the
10 product of average dose and the number of people exposed is proportional to the number
11 of radiation-related cancers. The concept of collective dose is sometimes used to support a
12 moral argument against widespread use of technologies or practices that would, according
13 to the LNT hypothesis, involve individual exposures at doses so low that any associated
14 risk, from the standpoint of the individual, would be far smaller than other risks that are
15 casually taken in everyday life. A so-called threshold hypothesis, according to which
16 there is no radiation-related risk associated with exposures at doses below some universal
17 threshold dose, would obviate concern about exposures at doses below the threshold and,
18 specifically, arguments based on the concept of collective dose. Aside from collective
19 dose, however, it is worth emphasizing that the practical importance of the LNT vs.
20 threshold question is associated with doses at which the associated risks, if they exist, are
21 high enough to be of “legitimate” concern, as determined by the usual social and political
22 processes.

23 Historically, the LNT vs. threshold controversy has been associated with public
24 policy issues related to exposures that are widespread but (typically) low for individuals,
25 such as local and worldwide exposure to radioactive fallout from above-ground nuclear
26 test explosions carried out by different governments, mainly during the 1950s (Lewis,
27 1957, 1963; Caron, 2004). The threshold hypothesis, as applied to ionizing radiation and
28 to fallout exposure in particular, drew some of its legitimacy from the field of chemical
29 toxicology, where thresholds are the rule (Brues, 1958, 1960), whereas the LNT
30 hypothesis is more consistent with findings from experimental radiation mutagenesis. As
31 described by Caron (2004), the intellectual positions taken by proponents of the opposing
32 sides during the fallout controversy of the 1950s (no compelling evidence of increased
33 cancer risk at low radiation doses, vs. no compelling evidence against a radiation-related
34 increased risk) are very similar to the situation at the present time. Some differences

1 discussed in the present report include the present general acceptance of a mutational
2 basis for carcinogenesis, and evidence that radiation-related mutations tend to be more
3 complex than more common mutations associated with endogenous and other causes.

4 The present report has been preceded by other surveys of the biological and
5 epidemiological information that underlies our understanding of low-dose risk and its
6 estimation by extrapolation from data obtained at higher doses, notably and recently the
7 comprehensive reports of the United Nations Scientific Committee on the Effects of
8 Atomic Radiation (UNSCEAR 2000, Annexes G and I) and of a committee of the U.S.
9 National Council of Radiation Protection and Measurements, entitled “Evaluation of the
10 Linear-Nonthreshold Dose-Response Model for Ionizing Radiation” (NCRP 2001). The
11 existence of these reports has allowed the present ICRP Task Group to be somewhat less
12 comprehensive in its coverage of the field than might otherwise have been necessary, and
13 to concentrate on updated coverage of developments in areas of epidemiology,
14 fundamental biology, experimental radiation mutagenesis and carcinogenesis, and
15 uncertainty analysis.

16 Studies of cancer risk following exposure of human populations are the most
17 obvious sources of information applicable to radiation protection policy. However, as
18 discussed in Chapter 2, generalization of risk information, obtained from one exposed
19 population, to other populations with different characteristics and potentially exposed to
20 radiation from different sources, at different doses and dose rates, requires the use of
21 dose-response models to describe the behavior of risk as a function of radiation dose, as
22 well as possible modification of dose response by individual and environmental factors. It
23 also requires making assumptions that are often based on uncertain information.

24 Chapter 3 deals with events believed to be fundamental to radiation
25 carcinogenesis: radiation-induced DNA damage and its repair. In particular, the chapter
26 discusses the nature of radiation-induced damage and damage response pathways
27 including repair of DNA double-strand breaks (DSB), cell cycle checkpoint control, early
28 sensors of DNA damage, and signal transduction after irradiation. Questions of particular
29 relevance for the current investigation are comparability of molecular damage from
30 radiation exposure and endogenous causes, and comparability between radiation-related
31 damage from ionizing radiation at high cf. low doses and dose rates with respect to
32 mechanisms, pathways, and fidelity of repair.

33 Cellular consequences of radiation-induced damage are discussed in Chapter 4.
34 Rates of radiation-induced chromosome aberrations and somatic cell mutations were

1 among the earliest quantitative measures of the cellular effects of ionizing radiation, and
2 studies of these outcomes have been highly informative about dose response over a wide
3 range of doses, and about effects of dose rate and fractionation. Induction of bystander
4 effects in cells not directly irradiated, genomic instability in the progeny of irradiated
5 cells, and adaptive response are radiation-related phenomena that evoke questions about
6 the generality of inferences based on cellular studies.

7 Considerations of statistical power, and possible bias due to unobservable and
8 uncontrollable confounders, govern the extent to which useful epidemiological
9 information can be obtained at exposure levels of regulatory interest, and some degree of
10 extrapolation is unavoidable. Experimental approaches using animal models, discussed in
11 Chapter 5, offer considerably more control of radiation exposure and dose, genetic
12 background, and modifying factors including other exposures, and can facilitate very
13 precise determination of biological outcomes. On the other hand, analogies between
14 radiation-related risks in human beings on the one hand and inbred strains of
15 experimental animals on the other are necessarily limited. Low statistical power for low-
16 dose studies is problematic for experimental and epidemiological studies alike, but
17 indirect approaches, based on protraction and fractionation of exposure resulting in
18 moderate to high cumulative doses, offer insights into low-dose effects. Experimental
19 studies can of course be replicated, to provide a firmer basis for insights into mechanisms,
20 tissue modifying factors, and quantitative dose response.

21 The material of Chapters 2-5 highlights statistical variation inherent in estimates
22 obtained by fitting parametric models to epidemiological and experimental data, but also
23 more fundamental uncertainties about important factors that cannot be ignored, but about
24 which there may be only limited information. The implications of these uncertainties for
25 conventional estimates of radiation-related cancer risk, especially at low doses and/or low
26 dose rates characteristic of exposures most commonly encountered by radiation workers
27 and the general public, are investigated in Chapter 6. The approach taken is an exercise in
28 quantitative uncertainty analysis similar to approaches used in a number of recent
29 exercises by expert committees concerned with such risks. Central to the approach is
30 recognition of the fact that radiation protection is a political process, responsive to the
31 interests and perceptions of stakeholders with differing points of view, and relying upon a
32 knowledge base that is extensive but also uncertain. Acceptance of this fact implies that it
33 is important, for the benefit and information of participants and stakeholders in the
34 radiation protection process, to identify sources of uncertainty and to quantify the

- 1 implications of such uncertainty for estimated risk. Among the questions addressed is the
- 2 impact on radiation protection policy of treating the existence of a universal low-dose
- 3 threshold for radiation-related cancer risk as an uncertain possibility.

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2. EPIDEMIOLOGICAL CONSIDERATIONS

2.1 Introduction

Like other areas of epidemiological research, the study of radiation-related cancer risk began with clinical observations, the earliest of which may have been the 16th century identification by the physician Georg Bauer (more often known by his Latinized name, Agricola) of a specific condition, which he called “Joachimsthal Mountain Disease”, among miners in the Joachimsthal region of the present-day Czech Republic. The disease, the description of which now appears consistent with radon-related lung cancer but could also include other lung diseases such as silicosis (NAS/NRC, 1999; Toohey, 1987), was thought by Agricola to be caused by “metallic vapors” in mine atmospheres. Roentgen’s discovery of x rays in 1895 and Becquerel’s discovery of natural radioactivity the following year, and the subsequent use of both in science, medicine and industry, led to the recognition, documented by case reports early in its history, that radiation exposure might be harmful (Doll, 1995). The Court Brown and Doll study of mortality among British radiologists (1958; Smith and Doll, 1981; Berrington et al, 2001), which demonstrated a significantly increased risk of cancer mortality among radiologists who had registered with a radiological society before 1921 and who were therefore likely to have received higher doses than radiologists who began their practice later, is an example of an influential study in which the fact of exposure was related to risk but individual dose estimates were not available. However, experimental studies of radiation effects such as cell inactivation, mutation, and carcinogenesis have taken advantage of the experimenters’ ability to regulate, with precision, radiation dose to target cells or tissues. Similarly, epidemiological investigations of exposed populations have benefited enormously from information enabling scientists to reconstruct individual, and even organ-specific, radiation doses. Benefits include the estimation of dose-response relationships and of the modification of such relationships by individual properties such as sex, age, lifestyle, and genetic inheritance. Thus, dose reconstruction is a fundamental component of the epidemiology of radiation carcinogenesis, and tends to be well worth the often considerable effort and expense required.

“Risk” is a concept in common use that is often applied to the past and future experiences of individuals, but a numerical risk value can be estimated and verified only on the basis of population rates, e.g., by comparing cancer rates, in a population exposed

1 to a given radiation dose, with rates in an otherwise comparable population that is either
2 not exposed or exposed to a much lower radiation dose. Thus, when we speak of an
3 individual's risk we are really referring to a property of a population similar to that to
4 which the individual is assumed to belong.

5 The implications of risk for public policy, and for radiation protection in
6 particular, are controversial in large part because risk estimates are uncertain and because
7 there are legitimate interests both in avoiding radiation-related risks on the one hand and
8 in maintaining radiation-related benefits and/or avoiding costs associated with
9 unnecessary exposure reduction on the other. A person who may be at risk of radiation-
10 related cancer will naturally insist on proof that the risk either does not exist or is small
11 enough to be tolerated in view of the presumed benefit. A person whose interest is in
12 maintaining the benefit, or avoiding costs associated with reduction of exposure, will
13 demand proof that there is a risk that is high enough to be of concern. The problem is
14 inherently political, and its fair resolution requires information about risk, including its
15 uncertainty, framed so as to address the concerns of both viewpoints.

16 As epidemiological investigations of radiation-related cancer risk have evolved
17 over time, emphasis has shifted from the discovery that radiation is indeed a cancer risk
18 factor, to demonstration of radiation dose response, to identification of factors that modify
19 dose response, to examination of assumptions inherent in the risk estimation process.
20 Ionizing radiation exposure is a known, and well quantified, human cancer risk factor.
21 Nevertheless, estimation of cancer risk following radiation exposure is a very uncertain
22 process for most cases of regulatory and/or popular concern. One reason is that risk
23 estimates are usually applied to exposed populations different from those on which the
24 estimates are based. Another is that public and regulatory interest is usually with
25 exposures at radiation doses far lower than those at which useful information about risk
26 can be obtained by studying populations with such exposures.

27

28 **2.1.1 Evidence regarding radiation-related transgenerational cancer risk**

29 The current report is concerned mainly with the possibility that cancer risk may be
30 increased following exposure to ionizing radiation. There is a great deal of information
31 about this question. A second possibility, which is also a matter of concern, is that
32 exposure may be associated with increased transgenerational cancer risk. Various
33 epidemiological and laboratory studies have examined whether risks of cancer are raised
34 in offspring, following parental radiation exposure. These studies have been reviewed in

1 detail elsewhere (e.g. COMARE, 2002). Cellular and animal studies indicate that the
2 induction of cancer in the offspring of irradiated parents is possible in principle.
3 However, the findings in mice have not been consistent. In some strains, no effect has
4 been seen (e.g. Cattanaach et al., 1995), whereas in others a raised risk has been observed
5 that is greater than predicted by the conventional induction rate for gene mutations (e.g.
6 Nomura, 1982).

7 Epidemiological studies conducted in several countries do not provide convincing
8 evidence to suggest that occupational radiation exposure alone results in an increased
9 incidence of childhood cancer in the offspring of male workers; data for the offspring of
10 female radiation workers are too sparse to draw conclusions (COMARE, 2002). In the
11 case of a cluster of childhood leukemia cases among children in the village of Sellafield,
12 U.K., possibly associated with paternal employment at the nearby Windscale nuclear
13 reprocessing plant (Gardner, 1990), a better case can perhaps be made in the context of
14 the well-documented phenomenon of increased levels of childhood leukemia in so-called
15 new towns, in which there has been an influx of residents from different areas; the
16 postulated mechanism is an unknown viral etiology affecting previously unexposed
17 residents (Doll et al, 1994; Doll, 1999). In addition, follow-up of about 40,000 offspring
18 of the Japanese atomic bomb survivors has not shown any association between the
19 incidence of cancer in children and young adults and parental dose (Izumi et al., 2003).
20 Thus, the subject of transgenerational risk, while a legitimate subject of scientific
21 investigation, is insufficiently developed to provide much information on risks associated
22 with low-dose radiation. It is briefly discussed in Chapter 5 in connection with radiation-
23 induced genomic instability, but is not pursued further in this report.

24

25 **2.2 Dependence of cancer risk on radiation dose.**

26

27 We have reasonably good epidemiological information on cancer risk following
28 acute exposures in the range 0.2 Gy to 5 Gy and (for partial-body exposures) above.
29 There are numerous epidemiological studies of populations containing “high-dose”
30 subsets with radiation doses in this range. These populations include patients treated with
31 radiation for benign and malignant disease, patients who received extensive diagnostic
32 radiography over a lengthy illness, such as tuberculosis patients treated by lung collapse
33 therapy monitored by frequent fluoroscopy examinations, persons who received
34 substantial exposures because of their occupations, such as uranium miners exposed to

1 radon decay products in mine atmospheres and instrument dial painters who ingested
2 radium contained in luminescent paint, and survivors of the atomic bombings of
3 Hiroshima and Nagasaki, Japan. These studies, and in particular inferences based on the
4 moderate- to high-dose component of the populations under study, form the primary
5 epidemiological basis for estimation of radiation-related risk. Recent, comprehensive
6 reviews of epidemiological information on radiation-related cancer risk are to be found in
7 the UNSCEAR 2000 report (UNSCEAR 2000) and NCRP Report 136 (NCRP 2001a).

8 Some benchmarks of radiation exposure levels are given in Table 2.1. Yearly
9 natural background effective doses in normal background areas are 0.4 mSv from cosmic
10 radiation, depending upon altitude (the dose from a typical round trip between New York
11 and Paris by commercial airline would be 0.03 mSv), 0.5 to 4 mSv from radioactivity in
12 rocks and soil, depending on local geology, 0.25 mSv from naturally occurring
13 radionuclides in the human body, and on the order of 2.5 mSv to the lung from inhaled
14 radionuclides (radon, thoron, and their decay products) (UNSCEAR 2000). Common
15 diagnostic examinations produce effective doses ranging from 0.01 mSv for x rays of a
16 foot or hand to 4 mSv for a barium enema (Mettler and Upton, 1995), to 25 mSv for a
17 pediatric CT scan of the abdomen if adult settings are used (Brenner et al, 2003). An
18 astronaut may get about 2 - 3 mSv tissue-weighted effective dose on a typical 3-day space
19 shuttle mission, and about 50 mSv on a 60-day tour in the international space station
20 (NCRP, 2000). Estimated acute, neutron-weighted doses to the colon (weighted dose =
21 gamma dose plus 10 times neutron dose) from the atomic bombings of Hiroshima and
22 Nagasaki ranged from less than 1 mGy to nearly 6 Gy for survivors who were exposed
23 within 3 km of the explosions and who were still alive in October, 1950; among survivors
24 with estimated doses between 5 mGy and 4 Gy, the average was 200 mGy (RERF LSS
25 mortality data set, 2003). An acute, whole-body effective dose of 5 Sv is very likely to be
26 fatal without prompt medical attention, but radiation therapy for cancer usually involves
27 partial-body doses an order of magnitude higher. Fractionation or protraction of exposure
28 can allow higher doses to be tolerated in terms of acute effects. Cumulative occupational
29 exposures among monitored radiation workers were about 20 mSv in several major
30 studies (Gilbert, 2001) and the recommended upper limit for radiation workers is 20 mSv
31 per year averaged over 5 years, and no greater than 50 mSv in any one year (ICRP, 1991).
32 (However, yearly effective doses at the Mayak plutonium facility approached 1 Sv for
33 some workers during the earlier years of production (Akleyev and Lyubchansky, 1994;
34 Khokhryakov et al, 2000).)

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2.2.1 Existence of a dose response.

Dose-response data (e.g., pertaining to cancer morbidity) can be described in a number of ways, such as by arranging observations in order of dose, grouping them into consecutive dose intervals, and plotting cancer rates by dose interval (Figure 2.1). Sophisticated modeling of dose response is not strictly necessary to establish the existence of a dose response; that can be done by a test of increasing trend, usually obtained by fitting to the data a simple model, like one of the following:

$$\text{ERR}(D) = \alpha D, \tag{2-1}$$

$$\text{ERR}(D) = \exp\{\beta D\} - 1. \tag{2-2}$$

Here, ERR(D) is excess relative risk at radiation dose D, and α and β are unknown parameters. In testing for an increasing trend, the dose response is “statistically significant” when the statistical evidence is inconsistent with values of the parameter α or β less than or equal to zero. These simple models can be used in tests of overall tendency, or trend, and do not suffice to establish the shape of the dose response curve. In the example of Figure 2.1, in fact, neither of the fitted functions agrees particularly well with the plotted, dose-specific data points, especially at high doses, but both simple models serve to establish the existence of a dose response.

If statistical significance is not achieved by a trend test, it can be inferred that the evidence in favor of the existence of a dose response is not strong or that any dose response is too complex to be represented by such a simple parametric function. It *cannot* be inferred that there is *no* positive dose response, unless the trend is statistically significant in the negative direction; inadequate statistical power, because of inadequate sample size for the range of doses covered, can result in failure to achieve statistical significance in the presence of a positive dose response (see Section 2.4.2).

2.2.2 Estimating the dose response.

The information that can be derived from a dose-response analysis is always conditional upon assumptions about the functional relationship between radiation dose and exposure-related, excess risk. In Figure 2.1, the interval-based estimates are based on virtually no such assumptions; the different estimates are minimally correlated with each other, and that only because they share a common reference (i.e., the value for the zero-dose interval is constrained to be zero); thus, observations at any given non-zero dose

1 continuous function of dose, without threshold. Thus, cell killing is considered a
2 stochastic effect, the probability of which increases with increasing dose, and not a
3 deterministic effect, like tissue injury, which becomes noticeable when the proportion of
4 damaged cells exceeds some threshold level.

5 The general dose-response function (2-3) is not often used in epidemiological
6 research, mainly because the constrained parameters β and γ produce effects opposite in
7 curvature that may cancel each other out to some extent. While the model is used
8 successfully with very precise and numerous experimental data, most epidemiological
9 dose-response data lack the statistical power needed to support estimates for a model of
10 such complexity. This observation is illustrated here using the A-bomb survivor data of
11 Figure 2.1 for total solid cancers following a whole-body exposure, among the most
12 statistically powerful epidemiological radiation dose-response data in existence at the
13 time they were published (Thompson, 1994). The model fits these data reasonably well
14 (Figure 2.2, dashed line; Table 2.2), but statistically not significantly better than the linear
15 model of Figure 2.1 ($p = .11$). The estimated ERR per Gy at low doses (i.e., the estimated
16 value of α), 0.52 (90% confidence limits 0.16 - 0.83), does not differ markedly from that
17 according to the linear model, 0.57 (0.49 - 0.66); however, the confidence limits are
18 substantially wider for the more complex model, reflecting the wide range of
19 combinations of positive values of the parameters α , β and γ consistent with the data. The
20 analysis offers little evidence in support of a positive value of the (dose-squared)
21 parameter β ($p=.28$), but suggestive evidence in support of a non-zero value of the cell-
22 killing parameter γ ($p = .07$).

23 Less than 1% of the members of the Life Span Study Cohort for whom dose
24 estimates have been calculated have estimates greater than 2 Gy, and there are reasons to
25 believe that the dose estimates above 2 Gy may be biased upward (Pierce and Preston,
26 2000). Restriction of the dose-response analysis to subjects with doses under 2 Gy yielded
27 the linear-model parameter estimate $\alpha = 0.64$ (0.54- 0.74). Adding either the quadratic or
28 the cell-killing terms to the model produced zero or minimal change whereas adding both
29 of them yielded parameter estimates so uncertain as to be of no predictive value (Table
30 2.2).

31 In the remainder of this report, epidemiological risk estimates are based on linear
32 dose-response analyses.

33

1
2 **2.3 Inferences based on acute exposures in the moderate-to-high dose range**

3
4 **2.3.1 Modification of dose response by sex and age.**

5 The information obtained from studies of the A-bomb survivors and other
6 populations mentioned above is rich in detail. For many cancer sites and groups of sites,
7 we can estimate with some precision not only dose-specific risk of radiation-related
8 cancer, but also its variation by cancer site and by sex, age at exposure, attained age
9 and/or time following exposure. In general (but not always), radiation-related relative risk
10 is higher among women and following exposure at young ages. The relationship to age at
11 exposure is marked for thyroid cancer, acute leukemia, and female breast cancer (Ron,
12 1995; Preston, 1994; Preston, 2002; Land, 2003). Risk decreases somewhat, in relative
13 terms, with advancing age at observation, but increases in absolute terms because baseline
14 cancer risk tends to increase as a power of age, and faster than dose-specific ERR
15 decreases (Thompson, 1994; UNSCEAR, 2000; Pierce, 2002; Pierce and Vaeth, 2003).

16 The relative importance of exposure age and attained age as modifiers of radiation
17 dose response is uncertain, because in any epidemiological follow-up study the two
18 quantities are highly correlated and their effects are difficult to separate. With additional
19 follow-up as the major exposed populations are followed to the end of life span, the
20 importance of this question for lifetime risk will become moot because projection to the
21 end of life will no longer be required for subgroups exposed at young ages. However, the
22 dependence of radiation-related risk on exposure age and attained age probably will
23 remain complicated: one consideration is the presence of secular trends in baseline risk in
24 Japan during the period of follow-up for the atomic bomb survivors over the past half
25 century, the reasons for which are not entirely clear (Parkin, 2002).

26 Statistically stable descriptions can be obtained of the dependence of dose-specific
27 risk on sex, age, and time, for aggregations of cancer sites such as all cancers combined,
28 all solid cancers, all leukemia types, and other groupings. This is useful because radiation
29 protection is concerned with the totality of possible adverse consequences of exposure,
30 but also because overall patterns of dependence may emerge from such analyses that can
31 be incorporated into site-specific estimates, resulting in greater statistical precision
32 (Pierce and Preston, 1993; NAS/NRC, 2000; NCI/CDC, 2003).

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2.3.2 Modification by lifestyle and other individual factors.

There is a relatively small but growing amount of epidemiological information (Table 2.3) on modification of radiation-related risk by history of lifestyle factors such as tobacco smoking in the case of lung cancer (Prentice 1983; Kopecky, 1986; NAS/NRC, 1999; Lubin, 1995; Pierce, 2003), childbearing and breast feeding in the case of breast cancer (Boice, 1978; Shore, 1980; Land, 1994), ultraviolet light in the case of basal cell and squamous cell skin cancer (Shore, 2001, 2002; Ron, 1998), and disease history in the case of type C hepatitis infection and liver cancer (Sharp, 2002). Much more needs to be learned about interactions of ionizing radiation exposure with lifestyle factors and with exposures to other agents. It is not unlikely that some of our current inferences about dependence of radiation-related risk on exposure age, attained age, and sex may reflect secular changes in lifestyle, and in exposure to environmental agents, that have been associated with changes over time (and with successive birth cohorts) in both baseline and radiation-related risk.

2.3.3 Variation by population.

There does not appear to be an obvious, consistent relationship between baseline and radiation-related cancer risk, either across cancer sites within a single population or across populations for a single cancer site. In the female Japanese population generally, age standardized (world) rates per 100,000 per year are similar, at about 31 for gastric cancer and 34 for breast cancer (Parkin, 2002), whereas in the United States they are about 3 and 90, respectively. Among A-bomb survivors, the radiation-related excess relative risk at 1 Gy (ERR_{1Gy}) is 0.32 for gastric cancer and 1.6 for breast cancer (Thompson, 1994). Gastric cancer contributes a substantial proportion of total radiation-related risk, but that proportion is considerably less than the proportion of risk of *baseline* gastric cancer to total baseline cancer risk (about 22%) among A-bomb survivors (Thompson, 1994) and among Japanese generally (Parkin, 2002). In the United States, the ratio is 2% for males and 1% for females. For female breast cancer the opposite is true; the baseline rate in Japan is among the lowest in the world for developed countries whereas the total cancer rate is not much different from that in most other countries (Parkin, 2002) while, among A-bomb survivors, breast cancer contributes a disproportionately large fraction of the total radiation-related cancer burden (Thompson, 1994). In the United States, by contrast, baseline breast cancer rates are high but the

1 radiation-related excess risk (in absolute terms) per unit dose among medically-exposed
2 women is similar to that among the A-bomb survivors (Preston, 2002). That is, the dose-
3 specific, radiation-related component of total breast cancer risk is likely to be similar in
4 absolute magnitude for exposed Japanese and western populations but, in western
5 populations, smaller as a proportion of total breast cancer risk. For gastric cancer, on the
6 other hand, the US baseline rate is an order of magnitude lower than that in Japan,
7 whereas the limited information on dose-specific, radiation-related excess risk suggests
8 that, as a multiple of baseline risk, it may be comparable to that in the A-bomb survivors
9 (Griem, 1994; Carr, 2002).

10 The above information suggests that, for breast cancer, radiation-related excess
11 *relative* risk per Gy (excess risk per Gy expressed as a multiple of the Japanese baseline
12 risk) based on A-bomb survivor data would overestimate risk for an exposed US
13 population while, for gastric cancer, radiation-related excess *absolute* risk (the difference
14 between risk following exposure and the Japanese (baseline risk) would result in an
15 overestimate for the US population. For most other cancers we have almost no
16 information of a similar nature (Table 2.3). This is not a trivial matter, because any
17 transfer of a risk estimate from one population to another requires making an assumption,
18 explicit or implicit, about the relation between excess and baseline risk. Moreover, for
19 some sites (e.g., stomach, liver, and esophagus) baseline rates can differ markedly
20 between populations (Parkin, 2002).

21 It should not be surprising that the relationship between radiation-related and
22 baseline risk in different populations is not consistent for different cancer sites. There are
23 reasons, as yet poorly understood, why baseline breast cancer rates are high in the United
24 States, and why baseline gastric cancer rates are high in Japan. These reasons are almost
25 surely related to differences in lifestyle, since the descendants of immigrants to the United
26 States, for example, have tended to develop cancer rates that are typical of the general
27 U.S. population (Haenszel, 1968; Ziegler, 1993) and different from those of their
28 countries of ancestral origin. The lifestyle factors affecting the rates for breast and
29 stomach cancer are probably different, at least in part, and probably interact differently
30 with radiation dose.

31 Much of environmental, nutritional, and occupational cancer epidemiology is
32 concerned with identifying cancer risk factors that might account for some part of the
33 variation of site-specific baseline rates among populations. While there has been much
34 progress, the problem is vast and, as discussed in section 2.3.2, there is only limited

1 information on interaction between radiation dose and lifestyle factors in terms of cancer
2 risk. Thus, it is likely that, for the foreseeable future, the most useful information relevant
3 to transfer of radiation-related risk coefficients from one population to another will come
4 from multinational comparisons of site-specific radiation-related risk, rather than from
5 investigations of underlying cancer risk factors and their interactions with radiation dose.

6 7 **2.3.4 Radiation quality.**

8 Risk estimates for low-LET radiation protection purposes are based mainly on
9 epidemiological studies of populations exposed to substantial doses of medical x ray, or
10 to mixed gamma and neutron radiation from the Hiroshima and Nagasaki atomic bombs.
11 According to the DS86 dose reconstruction algorithm (Roesch, 1986) as represented by
12 public use RERF data sets (RERF, 2003), the correlation between neutron and gamma
13 dose within each city is greater than 95%, and the proportion of total absorbed bone
14 marrow dose contributed by neutrons is only 0.7 to 2.7% in Hiroshima and 0.3 to 0.7% in
15 Nagasaki, depending upon shielding and exposure distance (According to the as yet
16 unpublished DS02 dose reconstruction system, the neutron component is reduced slightly,
17 compared to DS86, in both Hiroshima and Nagasaki. In particular, an anticipated large
18 increase of the neutron component for low-dose survivors in Hiroshima did not
19 materialize (Preston et al, 2004).) Because of the relatively small contribution from
20 neutrons, there is minimal statistical power for estimating the relative biological
21 effectiveness (RBE) of the two radiation types based on the A-bomb survivor data.
22 Moreover, there are essentially no useful data on cancer risks in populations exposed
23 mainly to neutron radiation (IARC, 2000) and, therefore, the relative biological
24 effectiveness of neutron cf. gamma-ray dose can only be estimated from experimental
25 data. Risk coefficients for gamma ray dose are obtained from the A-bomb survivor data
26 through the use of a nominal weighting factor of 10 for the neutron component of dose
27 (Thompson, 1994). This weighting factor has been judged appropriate at A-bomb doses
28 of the order of 1 Gy; however, the variation in the estimated gamma-ray dose response
29 due to uncertainty in the weighting factor is not great, with 90% uncertainty limits¹ of
30 " 7% (NCRP, 1997).

¹ Here and elsewhere in this report, “confidence limits” or “confidence bounds” are used for statistical uncertainty in the classical sense, in keeping with conventional usage. “Uncertainty limits”, “uncertainty bounds”, “probability limits”, and “probability bounds” are used interchangeably for estimates that incorporate some information for which subjective or approximate assessments of uncertainty have been employed.

1 Cancer risks associated with alpha radiation exposure have been studied for lung
2 cancer among uranium miners exposed to inhaled radon decay products (NAS, 1999) and
3 in populations exposed to lower radon levels in residential settings, for bone cancer
4 associated with ingested ^{226}Ra and ^{228}Ra among former radium dial painters (Fry, 1998;
5 Stebbings, 1984; Carnes, 1997) and with injected ^{224}Ra in patients treated for benign
6 disease (Spiess and Mays, 1979; Nekolla 1999, 2000), and for cancers of the liver and
7 other organs in patients injected with x-ray contrast media containing thorium (Travis,
8 2003). Thus, estimates of cancer risk associated with exposure to alpha particle radiation
9 have a basis in direct observations, while estimation of risk associated with neutron
10 exposure is indirect, relying on scaled estimates of risk from low-LET radiation, using
11 experimentally-derived estimates of the effectiveness of neutrons compared to low-LET
12 radiation.

13 Epidemiological risk estimates based on exposure to gamma rays (photons with
14 energies of > 250 keV) and most medical x radiation (photons with energies in the 30-250
15 keV range) often are treated as interchangeable quantities (see, e.g., ICRP, 1991).
16 However, it has long been considered, based on biophysical considerations, that medical
17 x rays are more effective biologically than higher-energy gamma rays. This consideration
18 has been cited as a factor that may complicate inferences based on comparisons of cancer
19 risk associated with fractionated x-ray exposures and acute gamma ray exposures
20 (Brenner, 1999). Kocher et al (2002; also see NCI/CDC, 2003) have estimated uncertain
21 radiation effectiveness factors (REF), compared to gamma radiation, for 30-250 keV and
22 soft (<30 keV) x rays, assigning subjective uncertainty distributions with mean REF
23 values 2 and 2.7, respectively, and 95% uncertainty limits 1 – 4.7 and 1.1 – 6.4,
24 respectively for the two x-ray energy ranges. Electrons at energies like those of secondary
25 electron tracks induced by gamma-ray photons, i.e., above 30 keV, were assigned an REF
26 value of 1, while lower-energy electrons were assigned an uncertain REF with mean 2.6
27 and 95% limits 1.2 - 5.0.

28 29 **2.4 Estimation of risk at low doses and low dose rates**

30
31 Except for radiation therapy, where there is a recognized benefit from the
32 radiation dose itself, very few people are exposed to radiation effective doses of 0.2 Sv
33 and above. Most public concern is with exposures to less than 50 mSv, the historical
34 annual limit for radiation workers before a reduced level (20 mSv) was recommended in

1 ICRP Publication 60 (1991); that concern extends to effective doses well below 1 mSv,
2 the annual limit recommended by both ICRP (1991) and NCRP (1993), as well as the
3 annual dose from natural background radiation for most tissues other than the lung. As
4 previously mentioned, a chest x-ray delivers about 0.1 mGy to lung tissue; the dose to
5 breast tissue from a two-view mammography examination is about 3 mGy; and an
6 astronaut may get about 2.4 mSv tissue-weighted effective dose on a typical 3-day space
7 shuttle mission (NCRP, 2000).

8 9 **2.4.1 Difficulties of direct estimation of low-dose risk.**

10 Although such low-dose exposures (except, of course, the astronaut's) are very
11 common, it is extremely difficult to estimate the associated excess cancer risks by
12 studying populations with exposures limited to the low-dose range. This is because, at
13 low doses, the radiation-related excess risk, which is thought to be proportional to dose or
14 perhaps somewhat less when compared to risks at higher doses, tends to be dwarfed by
15 statistical and other variation in the background risk level in the absence of exposure.
16 Because of this, truly enormous sample sizes (e.g., millions) theoretically would be
17 required to obtain a statistically stable estimate of radiation-related risk, and even then the
18 estimate would be untrustworthy because we do not understand, and therefore cannot
19 control or adjust for, all of the sources of variation in baseline levels of risk (Land, 1980).
20 At higher dose levels there are fewer such problems because the excess risk tends to be
21 large relative to statistical variation in baseline risk, and we are more likely to understand
22 the causes of any substantial variation in baseline risk that might be confounded with
23 radiation dose.

24 25 **2.4.2 Illustrative example.**

26 Suppose (1) that baseline cancer risk in a given population, over a period of (say)
27 30 years, were known to be 10%, (2) that exposure to a whole-body effective dose of 1 Sv
28 would double risk over the same period, and (3) that excess risk were strictly proportional
29 to radiation dose over the interval 0-1 Gy. Suppose also that it were possible to find large
30 study populations with baseline risks known to be 10% and with uniform exposures to 1
31 Gy, 100 mGy, 10 mGy, or 1 mGy, and to observe them over 30 years. (This is a
32 simplified version of a study in which observed cancer frequencies in an exposed
33 population are compared with expected frequencies calculated on the basis of published
34 population rates.) The estimated excess cancer rate in such a population would be the

1 number of cancers divided by the population size, less the known baseline rate of 10%.
2 The estimate would be distributed approximately as a normal random variable with mean
3 equal to effective dose D , in Gy, times 10%, and variance equal to $(1 + D)$, times 10%,
4 divided by the population size, N . The population size needed to be able to detect the
5 excess risk associated with effective dose D , with probability 80% at the 5% significance
6 level, is shown in Table 2.4. The calculation is in fact an unrealistically optimistic one
7 since, as illustrated in a later example, we can never be that sure of the baseline rate in
8 any exposed population.

9 If an enormous study population is required to detect any excess risk associated
10 with exposure to a small radiation dose, it follows that, if we use a much smaller
11 population and fail to detect any excess risk, the implications are unexciting. A result
12 predictable under both of two opposing hypotheses supports neither of them against the
13 other. Thus, for example, failure of epidemiological studies to demonstrate a statistically
14 significant excess cancer risk associated with exposures on the order of 1 mGy does not
15 imply that there is no risk, although it does suggest that any such risk is small relative to
16 baseline cancer rates.

17 At low and very low radiation doses, statistical and other variation in baseline risk
18 tends to be the dominant source of error in both epidemiological and experimental
19 carcinogenesis studies, and estimates of radiation-related risk tend to be highly uncertain
20 both because of a weak signal-to-noise ratio and because it is difficult to recognize or to
21 control for subtle confounding factors. At such dose levels, and absent bias from
22 uncontrolled variation in baseline rates, positive and negative estimates of radiation-
23 related risk tend to be almost equally likely on statistical grounds, even under the LNT
24 hypothesis. Also, by definition, statistically significant positive or negative findings can
25 be expected in about one in twenty independent studies when the underlying true excess
26 risk is close to zero. Thus, even under the LNT hypothesis, the smaller the dose, the more
27 likely it is that any statistically significant finding will be a purely chance occurrence, and
28 that it will be consistent with either beneficial effects of radiation (hormesis) or a grossly
29 exaggerated risk (Land, 1980). Such estimates tend to be only a small fraction of the total,
30 but when selectively presented they can give the appearance of a substantial and even
31 overwhelming body of evidence in one direction or the other.

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2.4.3 Studies of low-dose exposures

i) Medical studies.

There is, in fact, some direct epidemiological evidence of excess cancer risk associated with radiation exposures on the order of a few tens of mGy. One example is increased risk of leukemia and solid cancer, which has been observed in several studies (Stewart, 1956; MacMahon, 1962; Monson, 1984; Harvey, 1985; Bithell, 1988) among children exposed in-utero to radiation from x-ray pelvimetry. The excess absolute risk coefficient calculated in this case was 6% per Gy (Doll and Wakeford, 1997).

A less direct, but nevertheless persuasive, example is increased breast cancer risk among young women exposed to high cumulative doses from multiple thoracic fluoroscopy examinations, delivered in fractions that were, on average, on the order of 10 mGy (Boice, 1991; Doody, 2001; Howe, 1995; Davis, 1987). Successive exposures were separated by a week or more, but were repeated often enough to yield cumulative doses of hundreds or even thousands of mGy. Excess (absolute) risks per unit of total dose (about 10 excess cases per 10,000 women per year per Gy at age 50, following exposure at age 25 (Preston et al, 2002)) were comparable to those associated with acute doses among atomic bomb survivors (Boice et al, 1979; Land et al, 1980; Little, 1999; Preston et al, 2002). A similar relationship for excess risk of lung cancer, compared to estimates based on high-dose, acute exposures, was not observed among fluoroscopy patients, even though lung doses were comparable to breast doses (Howe, 1995; Davis, 1987). Although excess lung cancer risk per unit dose of acute radiation is in general less than for breast cancer (Thompson, 1994), the difference between the breast and lung cancer findings among fluoroscopy patients suggests that there may be variation among cancer sites in terms of fractionation effects. It should be remembered, however, that exposure to tobacco smoke is by far the dominant risk factor for lung cancer. Among, for example, tuberculosis patients who underwent lengthy courses of lung collapse therapy associated with high cumulative radiation dose from fluoroscopic examinations, below-average exposure to tobacco smoke might mask a radiation-related increase in lung cancer risk.

A highly significant, dose-related excess risk of thyroid cancer was observed among 10,834 Israeli patients treated as children by x-ray depilation for ringworm of the scalp (*tinea capitis*), with estimated (fractionated) dose to the thyroid gland averaging 90 mGy (range 40 – 500 mGy), cf. 16,226 non-exposed comparison subjects (Ron et al, 1995). Estimated linear model ERR/Gy was 32.5 (95% CI 14 – 57), based on 44 cases

1 among the exposed and 16 among the non-exposed. No significant excess was observed
2 among 2,224 patients given similar treatment (average thyroid dose 60 mGy).in the
3 United States, cf. 1,380 given only topical ointment treatment; 2 thyroid cancers were
4 found in the x-ray group, consistent with general population rates, and none in the non-
5 irradiated group. The between-study difference in risk estimates was not statistically
6 significant, however (Shore et al, 2003).

7

8 **ii) Occupational studies.**

9 Except for (mainly historical) worker populations with fairly high levels of
10 exposure, such as uranium miners (BEIR VI), radium dial painters (Stebbing et al,
11 1984), Russian plutonium workers (Gilbert, 2002), and early radiologists (Matanoski,
12 1975; Smith and Doll, 1982), most occupational studies can be classified as low-dose and,
13 therefore, of low statistical power. Their main utility is to validate generally accepted
14 estimates in the sense that they are consistent with estimated radiation-related risks
15 among regulated radiation workers. For example, a large, combined analysis of cancer
16 mortality among nuclear workers in the United States, the United Kingdom, and Canada
17 found a statistically significant dose response for leukemia and a non-significant dose
18 response for all solid cancers which, although negative, had an upper confidence limit
19 consistent with linear extrapolation of estimates based on higher-dose data (Cardis, et al,
20 1995). Occupational radiation exposure and cancer mortality in the U.K. National
21 Registry for Radiation Workers were similarly associated, and consistent with estimates
22 based on the atomic bomb survivor studies (see below) (Muirhead et al, 1999). Patterns of
23 cancer mortality were inversely related to year of first employment among U.S.
24 radiological technicians, consistent with a radiation etiology given higher occupational
25 exposures to radiation in earlier compared to more recent times (Mohan, et al, 2002,
26 2003).

27

28 **iii) A-bomb survivor studies.**

29 It is sometimes forgotten that the vast majority of the exposed (as distinguished
30 from persons not present at the time of the bombings) Life Span Study (LSS) cohort of
31 atomic bomb survivors received radiation doses under 100 mGy (Table 2.5). For solid
32 cancer mortality between 1950 and 1997 (Preston, 2003), direct assessment of risks at
33 low doses obtained a statistically significant dose response when the analysis was
34 restricted to survivors with dose estimates less than about 120 mGy. The estimated ERR

1 per Gy over this range was 0.74 (90% CI 0.1 – 1.5). There was no indication that the
2 slope of the fitted dose-response curve differed significantly ($p > 0.5$) from the estimate
3 over the full dose range (ERR per Gy = 0.47), and no evidence of a threshold. As
4 discussed below, similar result was obtained from analyses of the same epidemiological
5 data using the DS02 dose estimates (Preston et al, 2004).

6 An earlier analyses of solid cancer incidence data from the LSS Tumor Registry
7 for 1958-1994 (Pierce and Preston, 2000) was focused on persons exposed at distances
8 under 3000 m, of whom about 10,000 had estimated neutron-weighted doses under 5
9 mGy and 41,000 had doses between 5 and 500 mGy. An analysis restricted to persons
10 exposed at distances less than 3000 m found a statistically significant linear dose response
11 which was not overestimated by linear-model risk estimates computed over the wider
12 dose ranges 0-2 Gy or 0-4 Gy (Figure 2.3). A statistically significant estimate was
13 obtained from an analysis restricted to the 0-120 mGy dose range; another finding was
14 that any threshold over 60 mGy would be statistically inconsistent with the data.

15 When cohort members exposed beyond 3000 m were included in the analysis, the
16 estimated slope of the fitted dose response was reduced slightly (by 3%), and the
17 statistical significance of the fitted linear dose response in the range 0-120 mGy was
18 reduced. Figure 2.3 shows a moving-average plot of dose-specific cancer rates over the 0-
19 500 mGy range, with uncertainty bounds corresponding to \pm one standard deviation (sd).
20 At 100 mGy the moving average estimate of relative risk is about 3.7 sd units above one
21 for an analysis restricted to survivors exposed at distances under 3000 m, and about 2 sd
22 units above the redefined baseline (represented by the dotted horizontal line at about
23 RR=1.04) using the less restricted data set.

24 Figure 2.4 is based on the same data as Figure 2.3, but shows linear regression
25 estimates of the ERR per Gy over dose intervals that are progressively trimmed of high-
26 dose data. Moving from right to left, the right-most estimate and its standard error are
27 based on observations over the dose range 0-2 Gy, the next on 0-1.5 Gy, and so on, while
28 the left-most one is based on data at 0-0.05 Gy. There is more variation between
29 consecutive estimates on the left-hand side of each graph than there is on the right-hand
30 side, and the \pm SE limits become progressively wider toward the left-hand side of each
31 panel as the dose range is further restricted at the high end (Donald Pierce, personal
32 communication).

33 The reference population used in the analyses of Figures 2.3 and 2.4 is the group
34 of “proximal” survivors (exposed within 3 km) in Hiroshima and Nagasaki with neutron-

1 weighted dose estimates less than 5 mGy. This choice was justified on the basis that the
2 “distal” population exposed beyond 3 km was more rural, and may have experienced
3 different cancer risk factors other than radiation, from those of the more urban proximal
4 survivors. The horizontal line in Figure 2.3, corresponding to a relative risk of 1.04,
5 represents the baseline if the distal survivors had been included in the analysis. Figure
6 2.5 repeats the analysis of the Figure 2.4 with the distal survivors included. While
7 estimates of ERR per Gy based on higher-dose data are little affected by the change, the
8 estimates at the left-hand side of Figure 2.5 are substantially lower than those at the left-
9 hand side of Figure 2.4, with similarly wide error bounds. Comparison of Figures 2.4 and
10 2.5 demonstrates the sensitivity of estimates, if based only on low-dose data, to the
11 influence of minor, and largely unknown or poorly understood, confounding factors.

12 The same overall patterns are seen in Figure 2.6, an analysis similar to Figure 2.5
13 (in that data for distal survivors contribute to the estimates) for LSS breast cancer
14 incidence, 1950-1990 (Land, 2003). Together, Figures 2.4 - 2.6 demonstrate that
15 regression estimates of dose-specific cancer risk for combined sites and for some single
16 sites are highly consistent with linearity, depend substantially on excess risk observed
17 among survivors with estimated doses under 200 mGy, and are statistically unstable when
18 based only on data pertaining to doses under about 100 mGy. These analyses provide no
19 strong evidence that excess risks per unit dose are substantially different at very low
20 doses than at doses up to 4 Gy.

21

22 **2.4.4 Extrapolation to low doses and dose rates.**

23 Epidemiological data are informative about radiation-related risks at acute doses,
24 on a logarithmic scale, in the moderately high (~ 1Gy), moderate (~100 mGy), and to
25 some extent, low (~10 mGy) dose ranges, but not in the very low (~1 mGy) and
26 extremely low (~0.1 mGy) ranges. Arguably the most important single problem in
27 radiation risk protection is how to extrapolate from statistically stable, and relatively
28 unbiased, risk estimates that pertain to higher-dose exposures, down to the lower dose
29 levels that are of greater concern in everyday life. The analyses of Figures 2.3 - 2.6
30 suggest that, for the 1958-87 LSS solid cancer incidence data at least, linear extrapolation
31 over one order of magnitude, e.g., from 2 Gy to 200 mGy, is justified. Dose-response
32 analyses for leukemia risk, on the other hand, support a linear-quadratic dose response
33 with approximate equivalence of the linear and dose-squared components of risk at bone
34 marrow doses around 1 Gy (Preston, 1994). Solid cancer mortality data (all sites

1 combined) for 1950-1997 (Preston, 2003) suggest linearity even for doses in the 0 – 150
2 mGy range; however, a later analysis, using the DS02 dosimetry, found a statistically
3 significant upward curvature over the restricted dose range 0-2 Gy, but the authors noted
4 that linear model dose-response analyses restricted to 0 - 1 Gy, 0 – 0.5 Gy, and 0 – 0.25
5 Gy gave substantially higher estimates of low-dose risk and they therefore did not
6 recommend using the linear-quadratic model to estimate low-dose risk (Preston et al,
7 2004).

8

9 **i) Dose and dose rate effectiveness factor (DDREF).**

10 The combined-site LSS solid cancer data support linearity of dose response down
11 to doses of low-LET radiation on the order of 200 and even 100 mGy. They provide no
12 evidence that linearity does not continue down to zero dose, nor do they rule out the
13 possibility of nonlinearity in the 10-mGy and lower range. The in-utero pelvimetry
14 studies, and the fractionated fluoroscopy study breast cancer data, suggest that radiation
15 doses on the order of 10 mGy are associated with excess cancer risk, while leaving room
16 for some dose-related variation in the amount (but not necessarily the existence) of excess
17 risk per unit dose. The curvilinearity of the LSS leukemia dose response is the main
18 epidemiological evidence in support of a reduced risk per unit dose at low and very low
19 doses (otherwise suggested by experimental observations (NCRP, 1980)), such as the
20 ICRP and UNSCEAR recommendation that extrapolated dose-specific risk estimates be
21 divided by a DDREF of 2 for chronic exposures and for acute doses less than 200 mGy
22 (ICRP, 1991; NCRP, 1993; UNSCEAR, 1993). A DDREF greater than 2 would in fact, in
23 the context of a linear-quadratic dose-response model, be statistically inconsistent with
24 the 1958-87 LSS solid cancer incidence data (Pierce and Preston, 2000).

25 An independent analysis of the 1958-87 tumor registry data by Little and
26 Muirhead (2000) used a linear-quadratic model to assess possible overestimation of low-
27 dose risk based on use of a linear dose-response model with these data, taking into
28 account random errors in DS86 neutron and gamma dose estimates, and systematic errors
29 in Hiroshima neutron dose estimates. They concluded that, for all solid tumors combined,
30 there was some indication of upward curvature over the 0-2 Gy dose range, but felt that
31 uncertainties in likely adjustments to the Hiroshima DS86 neutron dose estimates called
32 for a cautious interpretation of their findings (a prescient judgement in view of the later
33 mortality findings of Preston et al (2004) discussed above).

1 A DDREF would not be applied to the estimated linear-quadratic dose response
2 for leukemia, since it is already included in the model.

3
4 **ii) Site-specific differences.**

5 The analyses of Figures 2.3 – 2.5 are based on the numerous data for all solid
6 cancers combined, and that of Figure 2.6 is based on female breast cancer, for which the
7 radiation-related signal-to-noise ratio is high in the sense that dose-specific, radiation-
8 related risk tends to be high compared to the level of, and unexplained variation in, age-
9 specific baseline breast cancer rates. Risk estimates for thyroid cancer and leukemia are
10 based on far fewer cases, but signal-to-noise ratios tend to be high on a dose-specific
11 basis, especially for exposures at young ages. For these three cancer types, there is
12 evidence of radiation-related excess risk at doses below 200 mGy, and for all except
13 leukemia there is little evidence for departure of the dose response from linearity. For
14 most other cancer sites, however, numbers of cases and/or radiation-related signal-to-
15 noise ratio are too low to support strong statements about low-dose risk, although it also
16 can be said that there is little or no evidence of departure from linearity (e.g., Thompson,
17 1994).

18 The latter category of cancers includes some sites for which there is little or no
19 epidemiological evidence that radiation exposure either is or is not associated with
20 increased risk; examples include small intestine, prostate gland, female genital organs
21 other than ovary, squamous cell skin cancer, and chronic lymphocytic leukemia
22 (NCI/CDC, 2003). Rectal cancer falls into this category with respect to A-bomb exposure
23 but has been shown to be significantly associated with high-dose, partial body exposure
24 among patients given radiation therapy for cervical cancer (Boice, 1988). Cancer of the
25 small intestine, which is very rare in most populations (Parkin, 2002) can be induced in
26 experimental animals by high-dose irradiation of exteriorized intestinal loops (Osborne,
27 1963, as discussed by Watanabe, 1986) and the small intestine therefore is a susceptible
28 organ. However, the small intestine appears to have characteristics that render it highly
29 resistant to carcinogenesis at low-to-moderate levels of exposure to radiation and other
30 environmental carcinogens (Cairns, 2002; Potten, 2002; see Section 3.2.1). Thus,
31 inferences based on all cancers as a group, or on certain cancers for which there is
32 substantial information about dose response and its modification by other factors, need
33 not *necessarily* apply to all site-specific cancers, or even to all histological subtypes of
34 cancers of any given site. Nevertheless, there is evidence of some degree of commonality,

1 with respect to dose effects and their modification by sex and age, for cancers of many
2 different sites (Pierce, 1996), and it is therefore useful and informative to examine
3 radiation-related risk for groups of cancer sites.

4 5 **2.5 Thresholds cf. the linear, no-threshold hypothesis**

6
7 The so-called linear, no-threshold (LNT) hypothesis (see, e.g., Brenner and
8 Raube, 2001) is part of the current basis for risk-based radiation protection. The
9 hypothesis assumes proportionality between radiation dose and subsequent cancer risk,
10 usually with allowance for a DDREF to reduce risk per unit dose of low-LET radiation at
11 dose levels below 200 mGy (ICRP, 1991). However, at doses at which the DDREF
12 applies fully, excess risk is assumed to be proportional to dose. A consequence of the
13 LNT hypothesis is that exposures resulting in very small average doses to very large
14 populations are assumed to be associated with excess numbers of cancers that, although
15 undetectable by epidemiological study, might be numerous.

16 The threshold hypothesis is a competing hypothesis that, if generally accepted,
17 might make it easier to ignore possible consequences of very low dose exposures.
18 According to the hypothesis, there is some “threshold” dose below which there is either
19 no radiation-related health detriment or a radiation-related health benefit that outweighs
20 any detriment. If the threshold were a universal value, for all individuals and all tissues, a
21 consequence of the hypothesis is that, at some point, a very low dose to any number of
22 people would have no associated risk and could be ignored. Much, of course, depends
23 upon the value of the assumed threshold dose, since even under the LNT hypothesis there
24 must be a level of estimated risk so low that it is not worth the trouble to avoid. If,
25 however, thresholds existed but were known or believed to differ widely among
26 individuals and/or tissues, the effect of this knowledge on radiation practice and
27 philosophy might be much less, and radiation protection might be even more complex
28 than it is under the LNT hypothesis.

29 One argument made against the LNT hypothesis is that there is little or no direct
30 epidemiological evidence of excess cancer risk in populations exposed to less than 50
31 mGy or so. That isn't quite true, as discussed above, but it is true that there is no direct,
32 credible epidemiological evidence of a radiation-related risk associated with exposures on
33 the order of 1 mGy, for example. Nevertheless, as also discussed above, the argument is a

1 specious one; failure to detect a risk that (if it exists) is very small is not evidence that the
2 risk is zero.

3 A more subtle, and statistically more sophisticated, argument is to demonstrate
4 that a dose-response model with a threshold, such as a linear model for dose-specific
5 excess relative risk with a fitted negative intercept at zero dose, can fit a data set as well
6 as a linear or linear-quadratic model constrained to have a zero intercept (Hoel and Li,
7 1998 with critique by Little, 1999). The approach has the potential for showing
8 disproportionality between excess risk and dose, consistent with a threshold (and usually,
9 but not necessarily, also consistent with a linear-quadratic dose response), and could
10 conceivably provide more substantial evidence of a threshold. That strong support for a
11 threshold hardly ever is found in this way is more a reflection of low statistical power in
12 the low-dose region than of statistical evidence against the existence of a threshold. In a
13 more recent paper, Baker and Hoel (2002) modified the then-current DS86 A-bomb doses
14 for presumed systematic error in estimates of the neutron component of dose from the
15 Hiroshima bomb, and a dose-dependent relative biological effectiveness (RBE) for
16 neutrons compared to gamma rays, finding that an improved fit to morbidity data for solid
17 cancers and leukemia was obtained by introducing a threshold. However, their
18 assumptions about underestimation of the neutron dose for low-dose survivors of the
19 Hiroshima bombing, on which their conclusions depended, have not been borne out by
20 subsequent measurement data (Straume et al, 2003; Preston et al, 2004).

21 It is clear that epidemiological studies are very unlikely to establish the presence
22 or absence of a threshold at some low dose level, although they can place limits upon the
23 likely value of any possible threshold (Pierce and Preston, 2000). Radiobiological
24 evidence presented elsewhere in this report identifies the induction of double-strand DNA
25 breaks and more complex clustered DNA damage as probably the most important
26 mechanism by which ionizing radiation exposure contributes to radiation carcinogenesis.
27 Such events have been demonstrated by calculation (Brenner and Ward, 1992; Goodhead,
28 1994) and by experiment (Boudaiffa, 2000a, 2000b) to result from a single low-energy
29 electron track produced by an x-ray or photon interaction. At low doses and low dose
30 rates, the occurrence of such events is proportional to radiation dose and to the number of
31 cells irradiated (Kellerer, 1985). Current research on development of timely assays for the
32 presence and repair of DSBs may someday lead to findings that resolve the question of
33 low-dose thresholds vs. the LNT hypothesis. As discussed in Section 4.5 below, the
34 question is still very much in doubt.

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2.6 Conclusions: Implications for low-dose risk

Epidemiological data from studies of human populations exposed to ionizing radiation provide direct evidence that such exposure is associated with increased risk of cancer, and reason to believe that excess risk is not confined to persons exposed to very high radiation doses. Our knowledge of radiation-related risk is highly quantified, more so than for any other common environmental carcinogen, and we have learned much about factors that modify that risk. Our understanding of risks associated with doses commonly encountered in daily life is not insignificant; we know, for example, that such risks are far lower than those observed in populations exposed to hundreds or thousands of mGy. However, the problem of quantifying risks that are so low as to be practically unobservable, and then recommending policies based on that quantification, is very difficult.

It is highly likely that there will always be uncertainty about low-dose risk, and that we will have to come to terms with that uncertainty. One way to do that is to quantify the uncertainty in a manner consistent with mainstream scientific information, and to evaluate actions and policies in terms of plausible probability distributions of risks associated these actions and policies. An example of this type of approach is given in Chapter 6 below.

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3. LOW DOSE RISK - BIOLOGY

3.1 Introduction

The fundamental role of radiation-induced DNA damage in the induction of mutations and chromosome aberrations and the apparent critical involvement of aberrations and mutations in the pathogenesis of cancer provides a framework for the analysis of risks at low radiation doses and low dose rate exposures. Several key questions are important in considering the impact of exposure to low dose and low dose rate radiation at the cell and molecular level with respect to subsequent development of chromosome aberrations, mutations, and cancer. These questions relate to the nature of radiation-induced damage, the nature of repair and damage response pathways, and their role and impact on induction of chromosome aberrations, mutations and cancer. In this regard the fundamental questions at the cell and molecular level to be considered for understanding risks at low doses are: 1) whether the damage caused by radiation is similar or distinct from endogenous damage; and 2) does damage occur at low doses/dose rates by ionizing radiation that cannot be repaired accurately; 3) is damage induced under low dose and/or low dose rate conditions repaired by distinct mechanisms from damage induced at higher doses; and 4) are the signal transduction pathways activated by low dose and/or low dose rate conditions and what impact do these pathways have in determining the propagation or elimination of radiation damage in cells and tissues. Early studies in biology related to radiation-induced cancer were largely descriptive in nature. This was mainly related to technical limitations in biological research. As such the ability to directly study low dose effects was limited. However, recent advances in techniques in cell and molecular biology are increasing the ability to directly approach these important questions.

3.2 Damage caused by radiation.

It has long been known that radiation produces a broad spectrum of DNA lesions including damage to nucleotide bases (base damage), DNA single and double-strand breaks (SSBs and DSBs). Certain types of DNA base damage such as 8-hydroxydeoxyguanosine and thymine glycols have significant biological importance, but the available data suggest that such isolated base damage by itself probably plays a minor

1 role in radiation mutagenesis (Ward, 1995). It is generally accepted that unrepaired or
2 misrepaired DBSs are the principal lesions of importance in the induction of
3 chromosomal abnormalities and gene mutations (Goodhead, 1994; Ward, 1995).
4 However, recently it has become recognized that an important feature of radiation
5 damage is not the presence of any of these damages individually but instead their close
6 association creating “clustered damage”. Such clustered damage can arise from the
7 combination of direct damage induced by the original radiation track plus damage
8 generated from secondary reactive species arising from subsequent ionization events
9 (indirect damage) (Nikjoo *et al.*, 1999). Recent evidence has, in fact, shown that
10 substantial yields of DSBs may result from secondary electrons, with energies below the
11 ionization threshold, generated from the ionized nucleotides (Boudaiffa *et al.*, 2000).
12 Clustered damage may involve a SSB or DSB associated with base damage but can
13 involve far more complex associations including multiple closely spaced DSBs. Both the
14 frequency and complexity of clustered damage depend upon the linear energy transfer
15 (LET) of the radiation. Using sophisticated modeling and track structure methods, it has
16 recently been shown that nearly 30% of DSBs induced by low LET radiation are of a
17 complex form involving two or more DSBs. This value is 70% for high-LET radiation.
18 When breaks associated with base damage are included, then the complex proportion
19 becomes 60% and 90% for low and high-LET radiation respectively (Nikjoo *et al.*, 2002;
20 Nikjoo *et al.*, 2000; Nikjoo *et al.*, 1999; Nikjoo *et al.*, 2001). It is likely that as the
21 complexity of the damage increases, the damage will become less repairable and more
22 likely to lead to biological consequences (see below for further discussion). An important
23 aspect in considering the impact of exposure to low doses of ionizing radiation (IR), is
24 whether such damage is similar to that encountered endogenously. It is clear that a
25 significant level of oxidative damage can arise in cells from the generation of reactive
26 oxygen species (ROS) during normal cellular metabolism. In comparing ROS induced
27 damage with that induced by IR there appears to be similarities but also important
28 differences. One aspect of ROS and IR-induced DSBs, which can impact upon repair, is
29 the nature of their termini. Breaks induced by restriction enzymes have 3'-hydroxyl and
30 5'-phosphate moieties at their termini, a prerequisite for enzymatic ligation while the
31 majority of breaks generated by ROS and IR have ‘damaged’ termini, most frequently
32 3'-phosphate or 3' phosphoglycolate end groups (Ward, 1998). Some 5' termini with
33 hydroxyl end groups are also generated. Such termini require processing prior to ligation.
34 Excision of a damaged nucleotide will also frequently result in base loss at the break.

1 Recent evidence concerning the repair of such lesions will be considered below. These
2 aspects of the breaks are similar between ROS and IR induced damage although they
3 differ from DSBs induced during such metabolic processes as V(D)J recombination and
4 meiosis.

5 The predominant forms of ROS-induced damage are base damages and SSBs.
6 The frequency of DSBs generated by ROS depends upon the particular reactive species
7 but typically is less than 0.5% of the damage induced. More, importantly, these DSBs are
8 distributed relatively uniformly throughout the DNA. In contrast, due to non-
9 homogeneous energy deposition, the damage from even low doses of IR occurs in clusters
10 producing complex lesions. *It is unlikely that such damage will arise endogenously at*
11 *any appreciable frequency.* The impact of this difference on repair will be discussed
12 below.

13 UNSCEAR have explored the proposition that data on the high abundance of
14 spontaneously arising DNA damage could be used to argue that ‘a further small
15 increment of DNA damage from low doses of radiation will not impose significant risk;
16 that risk only becomes significant at relatively high doses when at a given level of
17 genomic damage, DNA repair capacity is exceeded’ (see UNSCEAR 2000). The principal
18 conclusion from UNSCEAR, which generally accords with that of the Task Group, is that
19 differences in the complexity (as discussed above) and repair characteristics (see later in
20 this chapter) of spontaneously arising and radiation-induced DNA lesions render this
21 proposition untenable.

22
23 **3.3 Damage Response Pathways.**
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25 The cellular responses to DNA damage include pathways of DNA repair, the
26 operation of cell cycle checkpoints and the onset of apoptosis. The latter two responses
27 overlap significantly and utilize, at least to some extent, the same sensor molecules or
28 complexes involved in damage recognition and signal transduction. There is mounting
29 evidence that the damage recognition complexes that control cell cycle checkpoint arrest
30 also influence or interact with the DNA repair machinery although the interplay between
31 the DNA repair pathways and between DNA repair and checkpoint control/apoptosis is
32 currently unclear. The operation of these responses serves two functions: one aim is to
33 enhance survival and a second is to maintain genomic stability. These are not necessarily
34 compatible outcomes. The principal evolutionary pressure for a lower organism such as

1 yeast is the survival of individual cells whereas in multicellular organisms a strong
2 selective pressure is the survival of the organism. Since the propagation of genetically
3 altered cells has the potential to kill higher organisms by tumor formation, mechanisms
4 have developed to prevent the growth of damaged cells. However, to achieve this, the
5 survival of individual cells may be compromised. The role of apoptosis for this purpose
6 has been evident for some time; the function of checkpoint control in this context is just
7 beginning to emerge. Thus, for radiation protection, it is necessary to evaluate not only
8 the mechanisms that repair DNA damage and enhance survival but also the mechanisms
9 that serve to limit the propagation of damaged cells. Below we consider first the
10 processes that repair damage induced by IR and then the steps involved in the responses
11 leading to checkpoint arrest and apoptosis. Since DNA double strand breaks (DSBs)
12 represent the major biologically significant lesion following radiation exposure, the focus
13 of the discussion will be on damage response mechanisms triggered by DNA DSBs.

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3.3.1 DNA DSB repair

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i) Non-homologous end joining (NHEJ)

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Five core proteins that function in NHEJ have been identified in mammalian cells (Fig 1). These include the three components of the DNA-PK complex, (Ku70, Ku80 and the DNA-PK catalytic subunit (DNA-PKcs)), together with XRCC4 and DNA ligase IV (for reviews see (Jeggo, 1998; Kanaar *et al.*, 1998; Lees-Miller and Meek, 2003; Lieber *et al.*, 2003). Mutations in any of these core components confer dramatic radiosensitivity and an impaired ability to rejoin DNA DSBs as monitored by pulse field gel electrophoresis (PFGE). Cells lacking these NHEJ components are also impaired in their ability to carry out V(D)J recombination, a process that involves the rejoining of site specific DSBs (see below). Patients with hypomorphic mutations in DNA ligase IV display immunodeficiency and defective mice, when viable, display severe combined immunodeficiency phenotypes (SCID) ((O'Driscoll *et al.*, 2001) for reviews see (Jeggo, 1998; Jeggo and Concannon, 2001; Schwarz *et al.*, 2003)). Recently, a further

1 component, Artemis, has also been shown to function in NHEJ and V(D)J recombination
2 (Moshous *et al.*, 2001). Artemis was identified as the protein defective in a class of SCID
3 patient. Cell lines derived from these patients are sensitive to IR but, in contrast to lines
4 defective in the other NHEJ components, Artemis defective cell lines are proficient in
5 DSB rejoining (Nicolas *et al.*, 1998). Artemis is able to function as a single strand
6 specific nuclease and its function in V(D)J recombination depends upon its ability to
7 cleave a hairpin intermediate generated during this process (Ma *et al.*, 2002). The role of
8 Artemis in rejoining IR induced breaks is less clear but it has been speculated that it may
9 function in modifying double stranded ends with additional DNA damage (Jeggo and
10 O'Neill, 2002). Finally, analysis of cell lines from human SCID patients has provided
11 evidence for a further factor required for NHEJ (Dai *et al.*, 2003).

12 In yeast, a range of additional proteins appear to be required for NHEJ. Mre11,
13 Rad50, Xrs2p are required for NHEJ in *Saccharomyces cerevisiae* but are dispensable
14 for end-joining in *Schizosaccharomyces pombe* (for a review see (Jeggo, 1998)). In higher
15 organisms, cell lines derived from Nijmegen Breakage Syndrome (NBS) carry mutations
16 in Nbs1, a functional homologue of Xrs2p (Carney *et al.*, 1998; Varon *et al.*, 1998). NBS
17 cell lines are proficient in their ability to carry out V(D)J recombination and do not show
18 the characteristic DSB rejoining deficiency of NHEJ defective cell lines although they do
19 show radiosensitivity (Yeo *et al.*, 2000). Sir2p, Sir3p and Sir4p are also required for
20 NHEJ in *S. cerevisiae* (Tsukamoto *et al.*, 1996). Current evidence suggests that their role
21 may be regulatory and recently it has been established that in *S. cerevisiae*, NHEJ is
22 regulated in a cell-type specific manner by Nej1p/Lif2p (Frank-Vaillant and Marcand,
23 2001; Kegel *et al.*, 2001; Valencia *et al.*, 2001). Consistent with this model, Nej1p is
24 repressed in *sir* strains. This regulation ensures that NHEJ only functions in haploid yeast
25 cells and demonstrates that the role of the *sir* proteins in NHEJ is to regulate Nej1p. There
26 is no data to indicate whether NHEJ is regulated in a similar manner in mammalian cells
27 although the process clearly functions in diploid mammalian cells. An Nej1p homologue
28 has not been identified in mammalian cells.

29 The heterodimeric Ku protein, consisting of 83 and 70 kDa subunits, has DNA
30 double stranded end-binding activity and its binding to DNA ends is likely to represent an
31 early step in the repair process. The binding of Ku to dsDNA ends serves to recruit DNA-
32 PKcs and activate its catalytic activity. DNA-PKcs is a member of a sub-family of
33 phosphoinositol (PI) 3-kinases, termed PI 3-K related protein kinases (PIKK), that have
34 protein rather than lipid kinase activity (Hartley *et al.*, 1995), which potentially provides

1 the cell with a signal transduction pathway to alert the presence of a DNA DSB.
2 However, the function and physiological targets of DNA-PK activity is currently unclear.
3 It does not appear to be involved in p53 activation nor for cell cycle checkpoint arrest
4 (Jimenez *et al.*, 1999). There is mounting evidence that DNA-PK may serve to auto-
5 regulate the process of NHEJ and one clear *in vivo* substrate of DNA-PK activity is the
6 protein Artemis, which is stimulated to cleave hairpin junctions by DNA-PK dependent
7 phosphorylation (Ma *et al.*, 2002; Merkle *et al.*, 2002). Autophosphorylation of DNA-PK
8 also appears to be essential for NHEJ (Ding *et al.*, 2003).

9 XRCC4 and DNA ligase IV co-associate strongly and depend on each other for
10 stability (Critchlow *et al.*, 1997; Grawunder *et al.*, 1997). XRCC4 has no obvious
11 domains or motifs (Li *et al.*, 1995). The crystal structure of XRCC4 reveals a globular
12 head domain and two coiled coil tails (Sibanda *et al.*, 2001). DNA ligase IV has a
13 catalytic domain at its N terminus plus two BRCT domains at its C-terminus and
14 interaction with XRCC4 occurs via the region between the two BRCT domains
15 (Grawunder *et al.*, 1998). It is the only mammalian ligase identified so far that can rejoin
16 double strand DNA ends. An emerging model is that Ku serves to recruit the DNA ligase
17 IV/XRCC4 complex to the DNA end and then translocates inwards to allow LX access to
18 the DNA end (Kysela *et al.*, 2003).

19 *Role of NHEJ in V(D)J recombination.* During B and T cell development, the V,
20 D, and J segments become rearranged into contiguous units by a process that involves the
21 introduction of site specific DSBs by two recombination activating genes (RAG1 and 2)
22 (for reviews see (Fugmann *et al.*, 2000; Gellert, 2002; Hesslein and Schatz, 2001). In
23 germ line cells, each V, D or J segment, termed a coding segment, is juxtaposed to a
24 recombination signal sequence (RSS). The DSBs are introduced at the junctions between
25 an RSS and its adjacent coding sequence. This process involves the introduction of a
26 single strand nick and a transesterification reaction generating a blunt ended RSS end and
27 a hairpin coding end. Rejoining yields accurately rejoined signal junctions and coding
28 junctions that frequently bear deletions or insertions. This rearrangement process coupled
29 with inaccurate rejoining of coding junctions plays a significant role in enhancing the
30 diversity of the immune response. Thus it appears that the cell utilizes the same DNA
31 NHEJ machinery to effect rearrangements during the V(D)J recombination process and to
32 rejoin radiation induced DNA DSBs.

33 The genetic requirements for signal and coding joint formation are distinct and
34 provide insight into the nature of the rejoining process. Rejoining of the blunt ended

1 signal junctions requires only Ku70, Ku80, Xrcc4 and DNA ligase IV. Thus, Artemis and
2 DNA-PKcs are largely dispensable for RSS rejoining. In contrast, all six proteins are
3 required for coding join formation (Moshous *et al.*, 2001). Recently, it has been
4 demonstrated that Artemis is activated by DNA-PKcs, and following activation is able to
5 cleave the hairpin coding junctions (Ma *et al.*, 2002). This neatly explains the
6 requirement of both DNA-PKcs and Artemis for coding join formation. What is the
7 likely role of Artemis and DNA-PKcs in the rejoining of radiation induced breaks? In
8 unphosphorylated form, Artemis has 5' to 3' exonucleolytic activity with single strand
9 (ss) DNA specificity (Ma *et al.*, 2002). Upon phosphorylation by DNA-PK, its activity
10 changes and Artemis gains single strand specific endonucleolytic activity on both 5' and
11 3' overhangs as well as the ability to cleave hairpins. It is, therefore, possible that
12 Artemis functions to modify the ends of radiation induced breaks (Jeggo and O'Neill,
13 2002).

14

15 **ii) Homologous Recombination (HR).**

16 HR is a high fidelity and efficient mechanism to repair DNA DSBs that utilizes
17 information on the undamaged sister chromatid or homologous chromosome to retrieve
18 information lost at the break site. In yeast, genes involved in HR belong to the Rad 52
19 epistasis group (Rad50, Rad51, Rad52, Rad54, Rad55, Rad57, Rad59, Mre11 and xrs-2);
20 see (Helleday, 2003; West, 2003) for recent reviews. Homologues of some of these
21 proteins (e.g. Rad50, 51 52, 52, 54 and Mre11) have been identified in higher organisms.
22 The yeast proteins Rad55 and Rad57 display sequence similarity to Rad51 and in
23 mammalian cells further members of a Rad51 family (termed Rad51-like genes) have
24 been identified, including XRCC2, XRCC3, Rad51L1, Rad51L2 and Rad51L3 (Thacker,
25 1999). Steps involved in HR have been characterized in yeast and in *E. coli* and involve
26 processing of the DNA ends, strand invasion, the formation of heteroduplex DNA and a
27 step involving resolution of the cross-over junction (Holliday junction) (outlined in
28 Figure 1) (Kanaar *et al.*, 1998). RecA, Rad51p and human Rad51 (hRad51) polymerize
29 on DNA to form a nucleoprotein filament that promotes ATP-dependent homologous
30 pairing and DNA strand exchange. hRad52 stimulates homologous pairing by hRad51
31 suggesting that it functions in an early stage of Rad51-mediated recombination that
32 precedes homologous pairing (Benson *et al.*, 1998; New *et al.*, 1998; Shinohara and
33 Ogawa, 1998). *In vitro*, the homology searching and strand exchange reaction is
34 facilitated by RPA, Rad55 and Rad57 although their precise roles are unknown.

1 Resolution of the Holliday junction complex is carried out by RuvABC in *E. coli* and
2 requires Rad51C and XRCC3 in mammalian cells (Liu *et al.*, 2004). Mre11, Rad50 and
3 xrs2 may play a role in early nucleolytic processing to produce ends suitable for the
4 exchange reaction (Tauchi *et al.*, 2002). There is also increasing evidence for roles of
5 BRCA1, BRCA2 and BARD1 in homologous recombination. Specifically, BRCA2 can
6 bind to Rad51 via its Brt domains and potentially plays a role in delivering Rad51 to
7 single stranded DNA (Pellegrini *et al.*, 2002; Yu *et al.*, 2003a). BARD1 interacts with
8 BRCA1 and loss of either prevents HR taking place (McCarthy *et al.*, 2003; Westermarck
9 *et al.*, 2003).

10

11 **iii) Single strand annealing (SSA).**

12 A third process for DSB rejoining identified in yeast is SSA, a mechanism that
13 uses short regions of homology, possibly to stabilize ends prior to rejoining. The analysis
14 of sequences at the break junctions in mammalian mutants arising after radiation in higher
15 organisms has suggested that this mechanism also functions in mammalian cells (Morris
16 and Thacker, 1993). This mechanism is inherently inaccurate since it involves loss of
17 sequences around the short regions of homology. This may be the mechanism utilized
18 when HR or NHEJ fail and could thus potentially contribute to error prone DSB repair.
19 Unfortunately, little is known about the genetic requirement for this process in
20 mammalian cells.

21

22 **iv) Contribution of HR and NHEJ to DNA DSB repair in mammalian cells.**

23 Yeast mutants defective in Rad51p, Rad52p or Rad54p are dramatically
24 radiosensitive; yeast NHEJ null mutants display little or no sensitivity but double mutants
25 defective in both HR and NHEJ are slightly more sensitive than single mutants defective
26 in HR. Thus, in yeast HR is the major mechanism for DSB repair and NHEJ functions in
27 its absence. Two factors may account for this. Firstly, Nej1p appears to regulate NHEJ in
28 yeast resulting in repression of the process in *MATa/MAT α* diploids (Frank-Vaillant and
29 Marcand, 2001; Kegel *et al.*, 2001; Ooi *et al.*, 2001; Valencia *et al.*, 2001). Additionally,
30 NHEJ appears to be regulated in some additional way allowing it to function primarily in
31 G1 phase. The situation in mammalian cells is quite different. The major radiosensitivity
32 of NHEJ defective mammalian cells attests to the importance of NHEJ in the repair of
33 DNA DSBs in higher organisms. However, HR does function in higher organisms and

1 radiosensitivity is a feature of some HR defective cell lines. Increasing evidence suggests
2 that in higher organisms HR functions to repair breaks in late S and G2 phases and that
3 NHEJ plays its major role in G1 phase (Fukushima *et al.*, 2001; Rothkamm *et al.*, 2003).
4 In mammalian cells, HR utilizes sister chromatids rather than a homologous chromosome
5 as the source of undamaged template. HR, therefore, plays a particular role in the repair
6 of breaks at the replication fork and also appears to be essential for the efficient repair of
7 breaks that arise from interstrand cross-links.

8

9

3.3.2 Cell cycle checkpoint control.

10 Perturbation to DNA metabolism, arising either endogenously or through
11 exogenous DNA damaging agents causes arrest at one of several cell cycle checkpoints,
12 collectively called DNA integrity checkpoints. Progression from one cell cycle phase to
13 the next occurs by phosphorylation or dephosphorylation of cyclin dependent kinases
14 (Cdks) and checkpoint arrest is effected by controlling the activity of the DSBss. In
15 addition to checkpoint controls that operate at the boundary between cell cycle phases
16 there is also an S phase checkpoint that presumably recognizes a stalled replication fork.
17 These checkpoint responses have been most widely studied using *Saccharomyces*
18 *cerevisiae* or *Schizosacchromyces pombe* as model systems but the operation of
19 checkpoints is also evident in mammalian cells and homologues of most of the yeast
20 checkpoint proteins have now been identified. The checkpoint responses involve three
21 stages; damage recognition, signal transduction and effector proteins. A brief overview of
22 the process in yeast will be given first followed by a discussion of the available
23 knowledge in mammalian cells.

24

25 **i) DNA integrity checkpoints in yeast.**

26 In yeast there are several points where cell cycle delay or arrest can occur: (a)
27 G1/S that serves to prevent replication of damaged chromosomes, (b) intra-S phase which
28 slows down or delays replication, and (c) G2/M which prevents transition from G2 into
29 M. In addition, there is a distinct response that monitors the replication status of the DNA
30 and prevents mitosis if replication has not been completed. SpRad3
31 (*Schizosaccharomyces pombe* Rad3) or ScMec1 (*Saccharomyces cerevisiae* Mec1) are
32 the phosphatidyl inositol 3-kinase-like kinases (PIKKs) that initiate the signal
33 transduction process by phosphorylating key proteins involved in cell cycle regulation
34 (see (Furuya and Carr, 2003; Osborn *et al.*, 2002; Rouse and Jackson, 2002). Both kinases

1 have partner proteins, SpRad26p and ScLcd1p/ScDdc2p, which most likely function to
2 target the kinase to the site of damage with recent evidence indicating that recruitment of
3 the proteins to the break site requires initial binding of RPA to single stranded regions of
4 DNA (Cortez *et al.*, 2001; Zou and Elledge, 2003). Activation of the kinases, however,
5 requires additional complexes. One is an RFC-like protein or protein complex represented
6 by ScRad24p and SpRad17p. The second complex contains PCNA-like proteins
7 (ScRad17p/ScDdc1p/ScMec3p and SpRad1p/SpRad9/SpHus1). The RFC-like proteins
8 can target damaged sites independently of the PIKKs and are required to load the PCNA-
9 like proteins. Downstream phosphorylation of transducer proteins in cell cycle checkpoint
10 control, such as the Chk1p and Rad53/Cds1 kinases, requires all the proteins described
11 above. Through effector proteins that include the Wee1 kinase, Cdc25 phosphatase and
12 Mik1 kinases, key Cdks that control cell cycle progression are activated or deactivated.
13 These include the mitosis-inducing kinase Cdc2.

14

15 **ii) Checkpoint responses in mammalian cells.**

16 Although the steps are less well understood in mammalian cells, the checkpoint
17 responses are clearly conserved between organisms (for reviews see (Durocher and
18 Jackson, 2001; Rouse and Jackson, 2002)). However, whereas in yeast, nearly all
19 checkpoint signaling is carried out by the ScMec1/SpRad3 kinases, which respond to a
20 range of different DNA damages, in mammalian cells there appears to be some
21 divergence of function with two PIKK kinases, ATM (ataxia telangiectasia mutated
22 protein) and ATR (ataxia telangiectasia and Rad3-related protein), both contributing to
23 damage-dependent phosphorylation events (Abraham, 2001; Bradbury and Jackson, 2003;
24 Shiloh, 2001). ATM appears to respond primarily to DNA DSBs and, therefore, is the
25 PIKK activated by IR. ATR, in contrast, appears to be activated by single stranded
26 regions of DNA arising during stalled replication forks or during processing of bulky
27 lesions (Zou and Elledge, 2003). A further significant difference in higher organisms is
28 the role of p53 in the signal transduction process, for which there is no functional
29 homologue in yeast. Mounting evidence suggests that recognition complexes similar to
30 those found in yeast, sense damage and by phosphorylation initiate signal transduction
31 pathways (Rouse and Jackson, 2002). In mammalian cells, these pathways also target
32 p53. The result of this is that in mammalian cells checkpoint activation, in addition to
33 inducing transient delays at cell cycle transitions, can also mediate permanent cell cycle
34 arrest or apoptosis (outlined in Figure 2).

3.3.3 Early sensors of DNA damage.

i) Role of ATM

Ataxia-telangiectasia mutated (ATM) is the protein defective in ataxia-telangiectasia (A-T), a multi-system disorder associated with diverse characteristics that include cancer predisposition and clinical radiosensitivity (Taylor *et al.*, 1996). A-T cell lines are defective in a range of damage responses following IR including an inability to arrest at the G1/S, S and G2/M cell cycle checkpoints (Goodarzi *et al.*, 2003; Shiloh, 2001; Shiloh, 2003). Significantly, p53 levels are not elevated following radiation in A-T cell lines suggesting that ATM functions upstream of p53 potentially as part of an early damage sensor mechanisms (Kastan *et al.*, 1992; Lu and Lane, 1993). ATM is a member of the PIKK family with homology to SpRad3 and ScMec1 although the yeast homologue of ATM is Tel1 (Savitsky *et al.*, 1995). ATM can function as a ser-thr protein kinase both *in vivo* and *in vitro* and specifically can phosphorylate the serine 15 residue of p53. (Banin *et al.*, 1998; Canman *et al.*, 1998; Khanna *et al.*, 1998). This residue of p53 fails to become phosphorylated in irradiated A-T cells demonstrating that ATM functions as the major, if not the only, kinase phosphorylating this residue of p53 after irradiation. This was initially thought to provide the explanation underlying p53 induction following irradiation. However, this is clearly an over-simplification; firstly phosphorylation of this residue does not appear to be a key factor controlling p53 stability, secondly ATM can phosphorylate other sites on p53, and thirdly it can phosphorylate other kinases such as Chk1 and Chk2, which themselves phosphorylate p53 on serine 20, which is required to stabilize p53 (see section on p53 below). Furthermore ATM can also phosphorylate MDM2, an event that could itself influence p53 stability. Added to this complex picture, other kinases including DNA-PK and ATR, can, at least *in vitro*, phosphorylate the S15 residue of p53. Thus, a complex picture of p53 regulation by phosphorylation emerges in which ATM clearly plays an important role either directly or indirectly. Taken together, these data suggest that ATM plays a key role in sensing DNA DSBs and by phosphorylation initiating signal transduction pathways that control cell cycle arrest. ATR probably serves the same role for UV induced lesions and stalled replication forks and overlaps to some degree with ATM for DNA DSBs.

1 **ii) Role of Nbs1, hMre11 and hRad50.**

2 Nijmegen Breakage Syndrome (NBS) is another syndrome associated with cancer
3 predisposition and radiosensitivity that is distinct from, but overlaps with A-T
4 (International Nijmegen Breakage Syndrome Study Group, 2000; Shiloh, 1997). In
5 contrast to their somewhat distinct clinical characteristics, cell lines derived from A-T and
6 NBS display similar phenotypes including radiosensitivity, cell cycle checkpoint defects
7 and decreased ability to stabilize p53. The gene defective in NBS has been shown to
8 encode a protein, Nbs1 or p95 (Carney *et al.*, 1998; Varon *et al.*, 1998). Nbs1 interacts
9 strongly with hMre11 and hRad50. In yeast Mre11 and hRad 50 interact with a third
10 protein, Xrs-2p, and mutants defective in any of these proteins share identical phenotypes
11 (Johzuka and Ogawa, 1995). Nbs1 appears to be a functional homologue of Xrs-2p
12 although the two proteins share only limited sequence homology. Like other DNA repair
13 proteins, Nbs1 has a fork-head associated (FHA) domain and a BRCT domain, which
14 appears important for function (Cerosaletti and Concannon, 2003). The link between A-T
15 and NBS has been even further strengthened recently by the finding that a milder variant
16 form of A-T called A-T like disorder (ATLD) has mutations in hMre11 (Stewart *et al.*,
17 1999). hMre11 and hRad50 null mice show embryonic lethality and the mutations in
18 hMre11 in ATLD impair but do not inactivate hMre11 function, a feature consistent with
19 the milder clinical features of this variant class of A-T. hMre11, hRad50 and p95 (called
20 the MRN complex) co-localise in nuclear foci which form at the sites of DSBs
21 (Kobayashi *et al.*, 2002)]. The precise role of the MRN complex is still hotly debated. In
22 yeast and vertebrates, there is evidence that MRX functions in both HR and NHEJ
23 (Tauchi *et al.*, 2002). In mammalian cells it is not an essential component of the NHEJ
24 machinery, however (O'Driscoll *et al.*, 2001). Importantly, current evidence also shows
25 that MRN is required either directly for ATM activation or to aid ATM-dependent
26 phosphorylation events (Girard *et al.*, 2002; Uziel *et al.*, 2003). Taken together the
27 findings suggest that MRN acts in concert with ATM in an early sensor complex that
28 activates by phosphorylation a number of damage response mechanisms that include p53-
29 dependent and independent processes.

30
31 **iii) BRCA1 and BRCA2.**

32 Germline mutations in these genes confer a high risk of breast and ovarian tumors
33 and both have been identified as genes defective in familial breast cancer patients (Miki *et al.*
34 *et al.*, 1994; Wooster *et al.*, 1995). Recent evidence points to the involvement of both gene

1 products in damage response mechanisms and cells carrying mutations in either protein
2 show pronounced genomic instability (see (Venkitaraman, 2002) for a review). *BRCA1*
3 has an N-terminal RING finger domain that mediates protein-protein interactions and a
4 tandem BRCT motif at its C-terminus, which appears to represent a phospho-protein
5 binding module (Manke *et al.*, 2003; Yu *et al.*, 2003b). *BRCA1* defective cells show
6 marked genomic instability and impaired checkpoint responses including impaired S and
7 G2/M checkpoint arrest (Xu *et al.*, 2001). *BRCA1* is also localised to H2AX foci after
8 DNA damage and thus co-localises with MRN, 53BP1 and MDC1 (Paull *et al.*, 2000).
9 *BRCA1* is phosphorylated after DNA damage and emerging evidence suggests that it is
10 required to facilitate at least some ATM-dependent phosphorylation events, a feature also
11 displayed by other proteins that localise to the H2AX foci (Foray *et al.*, 2003; Lee *et al.*,
12 2000). However, following irradiation *BRCA1* also co-localizes with Rad51 to nuclear
13 foci, which are distinct from the H2AX foci (Zhong *et al.*, 1999). Consistent with this
14 finding, *BRCA1* defective cells are impaired in HR (Moynahan *et al.*, 1999). Taken
15 together, these results suggest that *BRCA1* may have two independent functions, one in
16 checkpoint signaling and another in promoting HR. Thus, like p53, *BRCA1* has a
17 "caretaker" role.

18 *BRCA2* defective cells do not appear to show cell cycle checkpoint defects but
19 they are impaired in homologous recombination (Moynahan *et al.*, 2001). Rad51 foci do
20 not form in *BRCA2* defective cells and it has been suggested that *BRCA2* is required for
21 the delivery of Rad51 to the sites of single stranded DNA (Pellegrini *et al.*, 2002; Yang *et al.*,
22 2002). The link with DNA repair has been further strengthened by the surprising
23 recent finding that *FANCD1*, a gene involved in cross-link repair and defective in some
24 patients with Fanconi anaemia, is in fact *BRCA2* (Howlett *et al.*, 2002).

25

26 **iv) Role of H2AX.**

27 H2AX is a variant form of the histone H2A, which becomes phosphorylated in response
28 to DNA damage and plays a critical role in the retention of repair factors at the site of
29 double strand breaks (Celeste *et al.*, 2003; Paull *et al.*, 2000). Mice lacking H2AX are
30 viable but show genomic instability and radiosensitivity (Celeste *et al.*, 2002). H2AX
31 phosphorylation is a rapid response following the introduction of DSBs and
32 phosphorylation rapidly extends to H2AX molecules located up to 3 megabase pairs
33 within the region of the DSB (Rogakou *et al.*, 1999). Using phosphospecific antibodies,

1 phosphorylated H2AX (termed γ -H2AX) can be observed as discrete foci and current
2 evidence suggests that all DSBs are marked by the presence of such foci (Rothkamm and
3 Lobrich, 2003). The analysis of such foci is promising as a tool to monitor the formation
4 and repair of DSBs (see also section 3.5).

6 **v) MDC1, 53BP1 and SMC1.**

7 Recent data have led to the identification of additional proteins that accumulate at
8 the site of γ -H2AX foci and are required for an efficient checkpoint response. Lack of
9 these proteins confers at least some level of radiosensitivity. 53BP1 was originally
10 identified through its ability to bind to p53 via C-terminal BRCT repeats present in
11 53BP1 (Mochan *et al.*, 2003; Wang *et al.*, 2002). MDC1 was identified simultaneously by
12 several laboratories, one of which identified it as a binding partner of the Mre11 complex
13 (Goldberg *et al.*, 2003; Lou *et al.*, 2003; Stewart *et al.*, 2003). Both proteins form foci
14 that co-localise with H2AX and MRN foci after irradiation (Abraham, 2002; Fernandez-
15 Capetillo *et al.*, 2002; Goldberg *et al.*, 2003; Lou *et al.*, 2003; Stewart *et al.*, 2003).
16 SMC1 is also required for normal cell cycle checkpoint arrest and for radioresistance
17 (Kim *et al.*, 2002; Yazdi *et al.*, 2002). SMC1 also localises at H2AX foci after DNA
18 damage.

20 **3.3.4 Signal transduction after irradiation.**

21 **i) Role of p53.**

22 An early response of mammalian cells that occurs within minutes of a cell
23 sustaining DNA damage is an increase in the levels of p53 (Kastan *et al.*, 1991). In
24 addition to changes in p53 levels, its ability to function as a transcriptional activator may
25 also be increased (see (Ashcroft *et al.*, 1999; Lakin and Jackson, 1999) for reviews). In
26 combination, these changes in p53 result in the transcription of key proteins involved in a
27 number of distinct damage response mechanisms (see below). The role of p53 in the
28 response to radiation damage is complex since it affects some aspects of DNA repair, cell
29 cycle checkpoint arrest and the onset of apoptosis (see (Fei and El-Deiry, 2003) for a
30 review). The importance of p53 and the significance of the damage response mechanisms
31 it controls is underscored by the dramatically elevated cancer predisposition in patients
32 with mutations in p53 (Li-Fraumeni syndrome patients), and in p53 knock-out mice

1 (Donehower *et al.*, 1992; Malkin *et al.*, 1990; Srivastava *et al.*, 1990). Additionally,
2 mutations in p53 are found in around 40% of tumors covering all the cancer types.

3 Since p53 is so critical to the cell and to the whole organism, it is not surprising
4 that it is subjected to stringent regulation, the complexity of which is ever increasing (see
5 (Ashcroft *et al.*, 1999; Deb, 2003; Lakin and Jackson, 1999) for reviews). A key protein
6 controlling p53 is Mdm2 (Deb, 2003). Mdm2 binds to the amino-terminus of p53 and
7 targets it for ubiquitination and subsequent degradation by ubiquitin controlled
8 proteosomes (Kubbutat *et al.*, 1998). Thus, in undamaged cells p53 is maintained at low
9 levels via Mdm2 binding and ubiquitin-dependent degradation. Following radiation
10 exposure, changes to p53 and/or Mdm2 decrease their binding potential with consequent
11 increase in the half life of p53. Additionally, however, Mdm2 binding represses the
12 ability of p53 to act as a transcription activator (Momand *et al.*, 1992). Thus, Mdm2
13 negatively regulates both stabilization of p53 and its function. Knock-out mice for Mdm2
14 are embryonic lethal due to high endogenous levels of p53 but double mutant p53/Mdm2
15 knock out mice are viable. More significantly, mutations in Mdm2 are frequently found in
16 tumors, particularly those tumors without p53 mutations. Mdm2 is also itself subject to
17 controlling mechanisms which include multi-site phosphorylation and sumoylation (Meek
18 and Knippschild, 2003). Another factor influencing Mdm2 function is the tumor
19 suppressor protein called p19^{ARF} in humans, which is derived from an alternative reading
20 frame of INK4a. p19^{ARF} binds directly to Mdm2 in a region distinct from the p53 binding
21 domain. It does not inhibit p53/Mdm2 binding but does inhibit p53 degradation probably
22 by sequestering Mdm2 into the nucleolus. The major mechanism regulating MDM2 bind
23 to p53 is phosphorylation, both of p53 and MDM2 itself. As discussed above, ATM plays
24 a role in both of these events.

25

26 **ii) G1/S arrest.**

27 Careful analysis has demonstrated that two types of G1/S arrest can occur in
28 mammalian cells: prolonged arrest which is a p53-dependent response, and a more
29 transient response (Di Leonardo *et al.*, 1994; Little, 1968) . The latter appears to be
30 similar to the G1/S response observed in yeast. The major p53 response protein required
31 for G1/S arrest is p21 (Wahl and Carr, 2001). Whilst p21 is transcriptionally regulated by
32 p53, there is also recent evidence that p53 regulates the stability of p21 via another p53
33 protein, p53RPF (Ng *et al.*, 2003). p21 is an inhibitor of cyclin dependent kinases and
34 plays its major role in G1/S arrest by binding to the cyclin D/Cdk6 complex and

1 inhibiting its ability to phosphorylate pRb, which in turn inhibits the release of pRb from
2 E2F, an essential step that triggers S phase progression (see Ko and Prives (1996) for a
3 review). Consistent with this model, neither p53 nor ATM null cells show prolonged
4 radiation induced G1/S arrest. A-T cells are, however, capable of arresting at the G1/S
5 boundary following UV irradiation, demonstrating the specificity of the upstream signal
6 transduction mechanism. However, the operation of this checkpoint does not necessarily
7 serve to elevate survival to IR since transformed fibroblasts (which normally lack this
8 response due to p53 inactivation) as well as p53 null cell lines display elevated
9 radioresistance compared to primary or p53^{+/+} cells (Lee and Bernstein, 1993).

11 **iii) S phase arrest.**

12 Replication in mammalian cells is also inhibited following irradiation, which can be
13 observed by decreased ability of replicating cells to incorporate radioactive precursors
14 into DNA. Cells from ATM and NBS display a phenotype called radioresistant DNA
15 synthesis (RDS), which is believed to be due to a failure to undergo S-phase delay
16 (Jackson, 2002). Current evidence suggests that early S phase arrest after irradiation is
17 ATM-dependent but at later times S phase arrest is mediated via ATR (Zhou *et al.*, 2002).
18 Chk2 and possibly Chk1 represent strong candidate proteins involved in mediating S
19 phase arrest via Cdc25A degradation (Iliakis *et al.*, 2003; Xiao *et al.*, 2003). S phase
20 arrest encompasses inhibition of ongoing replication forks, stabilisation of replication
21 forks and the inhibition of late firing replicons (Feijoo *et al.*, 2001; Tercero *et al.*, 2003).

23 **iv) G2/M arrest.**

24 Progression from G2 to M is controlled largely by the DSBs-cyclin B complexes.
25 Activation or inhibition of these complexes is controlled by opposing kinases and
26 dephosphatases affecting the phosphorylation status of the Thr14 and Tyr15 residues of
27 Cdk. Currently, the prevailing evidence suggests that ATM phosphorylates Cds1 and/or
28 Chk1, which in turn phosphorylates and inactivates Cdc25, the event that prevents
29 dephosphorylation and activation of Cdc2-cyclin B. G2/M arrest after γ -irradiation,
30 though ATM dependent is p53 independent. In earlier studies, confusion arose concerning
31 the G2/M checkpoint due to the ability of cells to arrest in two distinct ways in G2.
32 Normal cells in G2 at the time of irradiation show a delay in entry into mitosis, which
33 represents the operation of a G2/M checkpoint. A-T cells in G2 at the time of irradiation

1 show a reduced delay compared to normal cells showing that this arrest is at least
2 partially ATM dependent (Beamish and Lavin, 1994). However, following higher doses
3 asynchronous A-T and control cells can show a permanent arrest at G2/M which has
4 recently been shown to be ATR-dependent (Wang *et al.*, 2003). The contribution of G2/M
5 arrest to survival following radiation exposure is unclear, although the prevailing view is
6 that arrest enhances survival and reduces the probability of genomic alterations.

7 8 **v) Apoptosis.**

9 Apoptosis is a process utilized to balance cell proliferation and cell death. It is
10 crucial to certain developmental processes and is for example used during immune
11 development to remove cells that have failed to undergo productive rearrangements (Sohn
12 *et al.*, 2003). It is also utilized to remove cells damaged by exogenous DNA damaging
13 agents. The onset of apoptosis in normal cells by radiation is p53-dependent although
14 p53-independent routes to apoptosis have also been described (Adams, 2003).
15 Additionally, there are significant differences between cells lineages in their propensity to
16 undergo apoptosis following irradiation.

17 The signaling processes leading from p53 induction to apoptosis are complex and
18 diverse. All the pathways, however, converge in the activation of proteases termed
19 caspases (Adams, 2003; Thornberry and Lazebnik, 1998). Caspases exist as pro-enzymes
20 that require activation and finally effect apoptosis by protein degradation that results in
21 disassembly of cell structures such as nuclear lamin, degradation of DNA repair proteins
22 such as PARP, ATM and DNA-PKcs, and by enhancing DNA fragmentation via the
23 cleavage of I^{CAD}, an inhibitor of a nuclease capable of fragmenting DNA. One pathway
24 leading to apoptosis, and probably the pathway that plays the major role following IR
25 involves the bax/bcl2 family (Adams, 2003), of which at least 15 members have been
26 described. Bcl2 itself, first identified by its presence at a chromosomal translocation break
27 site in B-cell lymphomas, is an anti-apoptotic protein whilst, in converse, Bax, with
28 which it can dimerise is a pro-apoptotic protein. Bcl2 family members together regulate
29 the release of cytochrome C from the mitochondria, which serves to activate caspases
30 through an interaction with Apaf1 (Cory and Adams, 2002). Other routes to apoptosis
31 involve death receptor proteins that activate death ligands, which in turn activate caspases
32 (Ashkenazi, 2002).

3.4 Fidelity of DSB repair.

A crucial consideration for radiation protection is the level of fidelity with which DSBs breaks are rejoined and the impact of error-prone rejoining. In this context, several factors are important: a) the inherent fidelity achievable by the distinct DSB rejoining mechanisms, b) the fate of unrejoined and misrejoined breaks and c) the ability of radiation damage to undergo accurate repair compared to other forms of DNA damage, particularly endogenous damage. These three issues are discussed below.

3.4.1 The fidelity achievable by HR and NHEJ.

HR is clearly a high fidelity process utilizing sequence information from an undamaged template to repair coding information lost at a break site. The level of fidelity achievable by NHEJ, for either simple breaks or complex breaks, is still an open question. One difficulty in evaluating the studies on fidelity is that frequently restriction enzymes are used to induce DSBs, and these may be repaired with different fidelity to radiation induced DSBs. Studies in *S. cerevisiae* have examined the fidelity of rejoining simple restriction-enzyme induced breaks in the presence and absence of the individual NHEJ components. From these studies it has been concluded that Ku-dependent NHEJ is an accurate process that can act as a barrier to an alternative error prone end-joining mechanism (Boulton and Jackson, 1996). Recently, a study examining repair of a transposase-induced DSB in mammalian cells also concluded that NHEJ was normally accurate (van Heemst *et al.*, 2004).

As discussed previously the NHEJ pathway is also used during V(D)J recombination. The rejoining of these V(D)J breaks can also provide information on the accuracy of the process in mammalian cells. Although the coding joints generated during V(D)J recombination are rejoined inaccurately due to specific processing unique to lymphoid cells, the signal junctions are rejoined accurately (Gellert, 2002). In cell lines lacking components of the NHEJ machinery, both the frequency and fidelity of signal joint formation is dramatically reduced (O'Driscoll *et al.*, 2001; Riballo *et al.*, 2001; Taccioli *et al.*, 1993). This suggests that for these types of breaks, if rejoining is compromised, then the ends are subjected to nuclease digestion and repair is inaccurate. This suggests that NHEJ has the ability to rejoin a blunt ended break accurately and indeed, does so predominantly.

1 However, the repair of radiation induced breaks may be more demanding than the
2 repair of the breaks discussed above. Many of the radiation induced breaks may represent
3 non-ligatable ends or ends that require additional processing prior to ligation. One
4 approach that has been used to assess the fidelity of radiation-induced breaks during
5 NHEJ is a technique that monitors the mis-repair of DSBs by pulse field gel
6 electrophoresis (PFGE), a procedure that separates large DNA fragments (of the order of
7 10^6 base pairs) on the basis of size (Rothkamm *et al.*, 2001). To assess the fidelity of
8 rejoining the gels were probed using a large unique DNA fragment generated by digestion
9 of genomic DNA using a rare-cutting restriction enzyme. Following radiation exposure,
10 the unique restriction fragment became a smear of smaller size due to the presence of
11 DSBs within it. Following incubation to allow repair, the band was recovered
12 representing accurate repair. It was argued that whilst fragments smaller than the
13 anticipated size could arise from either inaccurate rejoining or lack of rejoining,
14 fragments larger than the anticipated size could only arise by mis-rejoining. When the
15 experiment was carried out following exposure to a high dose (80 Gy), significant mis-
16 repair could be seen. Although a limitation of this technique is that it necessarily involves
17 the use of high doses, the results, nevertheless, demonstrate that, under such conditions,
18 NHEJ has the potential to rejoin breaks inaccurately. Studies with NHEJ deficient cells
19 further suggest that the observed mis-repair is, in fact, mediated by the NHEJ pathway.
20 Using the same technique, mis-repair was also examined following 80 Gy delivered at a
21 low dose rate where radiation induced DSBs would be less likely to be in close proximity
22 to one another in both space and time. Under these conditions, there was much less
23 detectable mis-repair observed. Taken together, these findings suggest that the accuracy
24 of NHEJ may be influenced by the presence of neighboring breaks and suggest that the
25 process has the potential to be of higher fidelity when few breaks are present in any one
26 cell but that its fidelity may be compromised when many breaks arise independently.
27 Interestingly, similar experiments carried out following exposure to alpha particles
28 showed that there was no reduction in mis repair with increasing fractionation (Kuhne *et*
29 *al.*, 2002). These data are consistent with dose and dose rate data for the induction of
30 chromosome alterations following exposure to IR where effects are significantly reduced
31 for low dose rate exposures. It is important to note, however, that these data show that
32 misrepair can occur at high dose rates. They leave open still the question of whether
33 misrepair can also occur at low dose rates and doses. Exposure of mammalian cells to IR
34 causes a linear dose dependent increase in chromosome breaks, gaps and rearrangements

1 at relatively low doses and low dose rates. Making the reasonable assumption that
2 chromosomal rearrangements represent erroneous DSB rejoining events, such data would
3 argue for mis-repair mediated via the NHEJ machinery even under conditions where the
4 distribution of radiation induced DSBs are not in close proximity in space and time. This
5 argument, while strong, cannot be directly tested experimentally at this time.

6 Cells lacking components of the NHEJ machinery (for example *xrs-6* cells) show
7 elevated radiation-induced chromosomal aberrations relative to normal cells (Darroudi
8 and Natarajan, 1989; Kemp and Jeggo, 1986). This suggests that in the absence of Ku, a
9 lower fidelity rejoining process takes place. Although this finding does not directly
10 address NHEJ fidelity, it does strongly suggest that there is elevated infidelity in the
11 absence of Ku. In other words, Ku serves to promote accurate rejoining.

12 Finally, recent studies have also provided evidence for a process of rejoining
13 DSBs that involves rejoining of the breaks to dysfunctional telomeres (Bailey *et al.*, 2004;
14 Latre *et al.*, 2003). These studies thus open a new pathway for misrepair that represents
15 DSB-telomere fusions and could represent an important cause of genomic instability
16 induced by radiation (Urushibara *et al.*, 2004).

17 18 **3.4.2 The fate of unrejoined and misrejoined breaks.**

19 The role of cell cycle checkpoints is to prevent the proliferation of damaged cells.
20 It has been argued that a single unrejoined DSB is lethal to a cell. From the perspective of
21 a multi-cell organism this may not be unduly harmful. A misrejoined break, may not,
22 however, be recognized by a cell and therefore may pose a bigger threat as a potential
23 oncogenic lesion. Failure of cell cycle checkpoint control coupled with impaired DSB
24 repair will, however, pose a particular risk. In this light, patients such as A-T and NBS
25 patients, where the defects impair both DNA repair mechanisms as well as cell cycle
26 checkpoint control display significant cancer predisposition. Similarly, the combination
27 of p53 mutations with mutations in essential DNA repair genes (such as DNA ligase IV)
28 promote survival at the expense of elevated tumor predisposition (Zhu *et al.*, 2002). In
29 this light, the ability of low doses of radiation to affect cell cycle checkpoint control is
30 particularly important to evaluate.

31 32 **3.4.3 The impact of the nature of DNA damage on repair.**

33 As described above (section 3.2), the damage induced by IR is distinct from
34 endogenous ROS induced damage in its complexity. Single strand breaks (SSBs) are

1 repaired accurately and rapidly, and there are an array of glycosylases that recognize and
2 initiate the excision of specific damaged bases (for review see (Scharer and Jiricny, 2001:
3 Slupphaug, 2003). It is important to point out, however, that although ROS induced
4 damage may not directly induce DSBs, it is likely that DSBs do arise endogenously
5 potentially through the processing or replication of other lesions. It is also likely that such
6 breaks will have ends that require processing prior to rejoining. The major evidence
7 suggesting that DSBs arise spontaneously, is that cells lacking either NHEJ or HR
8 components display elevated instability (Difilippantonio *et al.*, 2000; Karanjawala *et al.*,
9 1999).

10 The repair of complex lesions induced uniquely by IR, may, however, pose a
11 problem for the DNA repair machinery. Studies are now emerging on how one type of
12 damage influences the repair of another. The current evidence suggests that the ability of
13 glycosylases to recognize and remove a damage base is impeded by the presence of a
14 nearby nick on the opposite strand (David-Cordonnier *et al.*, 2000; David-Cordonnier *et*
15 *al.*, 2001). Since clustered base damage arises frequently after irradiation, a considerable
16 number of additional DSBs could arise if clustering of base damage inhibits repair
17 (Gulston *et al.*, 2004). How the presence of a nick or damaged base affects DSB repair is
18 entirely unknown.

19 Classical analysis of post-irradiation cell survival has also provided evidence that
20 the highly complex lesions induced by high LET radiation are less repairable than low
21 LET radiation. Most specifically, cells lacking Ku (e.g. xrs-6), are relatively more
22 sensitive when compared to wild type cells to low LET radiation than to high LET
23 radiation, consistent with the notion that high LET radiation has a higher non-reparable
24 component (Thacker and Stretch, 1985). These studies have been discussed in detail in
25 previous ICRP and UNSCEAR reports and will not be discussed further here.

26 As mentioned above, ROS-induced damage as well as the damaged induced by IR
27 frequently have damaged termini, precluding their repair by direct ligation. Recently,
28 polynucleotide kinase (PNK), a protein that has both DNA kinase and DNA phosphatase
29 activities has been found associated with Xrcc1, one of the proteins involved in SSB
30 repair (Whitehouse, 2001). Furthermore, Xrcc1 can stimulate the activity of PNK. Thus,
31 the damaged 3' end, which cannot be subjected to direct ligation, is first processed by
32 PNK in the presence of Xrcc1, which then coordinates gap-filling, if necessary, by an
33 interaction with DNA polymerase β followed by subsequent ligation (Caldecott, 2002).

1 This important finding demonstrates how cells use their resources to coordinate repair
2 involving several distinct steps. However, these damaged termini arise frequently
3 endogenously which has likely provided a strong selective pressure to drive the evolution
4 of this co-coordinated repair process. This may not be the case for other, more complex,
5 lesions unique to IR.

7 **3.5 Impact of defects in DNA repair, checkpoint control and apoptosis.**

9 Disruption of the NHEJ components in mice result in varied phenotypes; loss of
10 XRCC4 and DNA ligase IV causes embryonic lethality, Ku defective mice senesce
11 prematurely whereas DNA-PKcs defective mice grow and develop normally although
12 manifesting severe combined immunodeficiency (Bosma *et al.*, 1983; Frank *et al.*, 1998;
13 Gu *et al.*, 1997; Nussenzweig *et al.*, 1996). Significantly, however, DNA-PKcs defective
14 mice have only a small elevated spontaneous cancer incidence. The situation with Ku is
15 exceptionally unclear; Ku80 defective mice display no elevated tumour incidence
16 whereas Ku70 defective mice develop a high incidence of lymphomas. Thus the impact of
17 the loss of NHEJ on tumor incidence in mice remains to be resolved. A defect in DNA
18 ligase IV has been identified in a leukemia patient who was normal until the onset of
19 leukemia at age 14 (Riballo *et al.*, 1999). The mutation identified in this patient conferred
20 significantly decreased but not ablated ligation activity. This suggests firstly that
21 impairment of NHEJ can be compatible with life, and confers significant radiosensitivity
22 without overt immunodeficiency. More importantly, the defect may confer leukemia
23 predisposition. The fact that this patient has decreased activity rather than totally ablated
24 activity may be significant.

25 Haploinsufficiency of ligase IV has been shown to result in an increased incidence
26 of sarcoma in *ink4a/arf*^{-/-} mice (Ferguson *et al.*, 2000; Sharpless *et al.*, 2001). Decreased
27 but not ablated DNA PKcs activity has also been associated with increased sensitivity to
28 radiation induced lymphomas (Mori *et al.*, 2001) and mammary tumors (Yu *et al.*, 2001)
29 in mice and lung and colon cancer in humans (Auckley *et al.*, 2001; Rigas *et al.*, 2001). It
30 has been hypothesized that because of the importance of the NHEJ pathway, complete
31 loss of function of one of the components in this pathway may result in a low frequency
32 of tumors because of significant problems with genomic integrity and stability. Cells
33 with such significant problems would manifest substantial genomic damage and would
34 likely be eliminated by the cell cycle and apoptotic response pathways before having the

1 opportunity to progress to become tumors. On the other hand, with less severe defects in
2 this pathway the cellular effects would be less severe and it would be more likely that
3 cells with less severe forms of damage could escape elimination (Ferguson et al., 2000).

4 Recent evidence suggests that defects in checkpoint control or apoptosis confer a
5 very different phenotype with significantly elevated cancer predisposition. Mice defective
6 in p53 display elevated spontaneous tumor formation both in the homozygous and
7 heterozygous state. Recently CHK2 was identified as the germ line tumor suppressor loci
8 of a small number of Li-Fraumeni families that did not have TP53 mutations. Both A-T
9 and NBS patients display significantly elevated tumor incidence. BRCA1 and BRCA2
10 defects are associated with cancer predisposition. Taken together, this suggests that whilst
11 lack of repair may simply enhance sensitivity, failure to arrest at a cell cycle checkpoint
12 or failure to undergo apoptosis may result in elevated carcinogenesis. The impact of these
13 processes for radiation protection is two fold. Firstly, although the effect of low dose
14 irradiation on DNA repair has been investigated almost nothing is known about the
15 impact of low doses on cell cycle checkpoint arrest. Secondly, the variation in these
16 responses between individuals is not known.

17 It has been proposed recently that a low-dose threshold could result, not from the
18 absence of DSBs and complex lesions at very low doses, but from the absence of repair;
19 i.e., affected cells are unable to replicate, and therefore do not contribute to
20 carcinogenesis. That is, the affected organism, or tissue, might be genetically
21 programmed to tolerate a certain amount of cell loss as a means of minimizing the risk of
22 mutation and cancer due to DNA misrepair. A recent study by Rothkamm and Lobrich
23 (Bonner, 2003; Rothkamm and Lobrich, 2003) involved the irradiation of cultures of
24 nondividing primary human lung fibroblasts with 90 kV x rays at doses ranging from 2
25 Gy down to 0.1 mGy. Numbers of DSBs formed were measured by immunofluorescence
26 of foci of the phosphorylated histone, (γ -H2AX). The investigators found that the number
27 of DSBs formed was linear with radiation dose, but that DSBs induced at 1.2 mGy (0.1
28 foci per cell cf. 0.05 per cell among controls) remained unrepaired for many days, in
29 contrast to efficient DSB repair following exposure at higher doses (0.66 and 0.22 per cell
30 at 20 mGy and 5 mGy, declining to 0.1 per cell after 24 hours). However, there is some
31 question about the extent to which the assay can be relied upon to quantify DSB
32 frequency following radiation exposure. For example, in this study the assay indicates a
33 surprisingly high frequency, and persistence over time, of DSBs in control cells, and a

1 high persistence of radiation-related DSBs following high-dose exposure compared with
2 findings from split dose experiments. It has been demonstrated that those proteins
3 involved in DSB rejoining, including H2AX, translocate substantial distances along the
4 DNA from the break, implicating other functions for these proteins (Rogakou *et al.*,
5 1999). Also, Petrini and Stracker (2003) note that, although late foci of DSB repair
6 proteins and γ -H2AX appear to be genuine reflections of DSB metabolism, it is
7 problematic to use them to draw inferences about recruitment to DSB sites because the
8 vast majority of DSBs are repaired by 90 minutes after their induction. There is also some
9 question whether the DSBs examined by Rothkamm and Lobrich were direct or indirect
10 effects of radiation exposure (Seymour and Mothersill, 2004). Thus, the implications of
11 this intriguing study for low-dose risk are at present unclear.

14 **3.6 Conclusions.**

17 Ionizing radiation is able to produce a unique type of damage in which multiple
18 lesions are encountered within close spatial proximity. Even a single track of IR through a
19 cell is likely to induce these unique clustered damages. This type of damage is unlikely to
20 be generated frequently endogenously or by other exogenous agents, and thus, there may
21 not have been a strong selective pressure driving efficient repair. Although cells have a
22 vast array of damage response mechanisms that facilitate the repair of DNA damage and
23 the removal of damaged cells, these mechanisms are not fool proof. Moreover, clustered
24 radiation-induced lesions pose a particular problem and current emerging evidence
25 suggests that closely spaced lesions can compromise the repair machinery. On this basis,
26 there is not any strong evidence for a radiation dose below which all radiation-induced
27 damage can be repaired with fidelity. Whilst many of the cells containing such radiation-
28 induced damage may be eliminated by damage response pathways involving cell cycle
29 checkpoint control and apoptotic pathways, it is clear from analysis of cytogenetics and
30 mutagenesis that damaged or altered cells are capable of escaping these pathways and
31 propagating. This further argues against the likely possibility of a threshold for radiation-
32 induced cellular effects.

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14

1 **4. CELLULAR CONSEQUENCES OF RADIATION-INDUCED DAMAGE**

2
3 The mis-repair of radiation-induced DNA double-strand breaks and other lesions
4 is believed to be the principal pathway for the induction of chromosome and gene
5 alterations responsible for the killing, mutagenic, and carcinogenic effects of ionizing
6 radiation. Studies focusing on cytogenetic damage and mutagenesis were among the
7 earliest quantitative measures of the cellular effects of ionizing radiation (Sax, 1938).
8 On a practical level such studies have provided considerable information on dose
9 response relationships over a wide range of doses and on the effects of dose rate and
10 fractionation (NCRP, 1980). On a more fundamental level these studies have provided a
11 substantial amount of information relevant to DNA damage after radiation, repair kinetics
12 and on underlying mechanisms. Because of the close mechanistic relationship between
13 chromosome aberrations, mutations, and cancer (UNSCEAR, 2000) such studies also
14 have particular relevance to radiation risks and the question of risks at low doses.

15 16 **4.1 Radiation-induced chromosome aberrations.**

17
18 The first documented account of the cytogenetic effects of x rays described the
19 production of dicentrics, centric rings and deletions in plant microspores irradiated in the
20 extended G₁ phase (Sax, 1938). It was very difficult using the standard staining technique
21 to observe reciprocal translocations and inversions; these aberrations are the most
22 frequently present in tumors of different types. This latter fact is the consequence of
23 reciprocal translocations and inversion being transmissible from cell generation to
24 generation, whereas dicentrics, centric rings and deletions are cell lethal as the result of
25 the loss of genetic material at cell division. The ability to analyze all types of
26 chromosome aberrations has been greatly enhanced by the use of Fluorescence In Situ
27 Hybridization (FISH) that is discussed in more detail below.

28 The early studies by Sax and his colleagues also demonstrated that the dose
29 response curve for dicentric aberrations fit a linear-quadratic model ($Y = \alpha D + \beta D^2$),
30 suggesting that some dicentric exchanges were produced by one ionization track (αD) and
31 some by two independent tracks (βD^2). Neutrons induced the same types of chromosome
32 aberrations, but in contrast the dose-response curve for dicentrics was linear indicating a
33 one-track mechanism of formation. The prediction, based on the proposed mode of
34 formation of aberrations, was that chronic exposures to x rays would produce all types of

1 aberrations linearly with dose, and that split doses would lead to lower aberrations
2 frequencies than the same dose given as a single exposure. These predictions were borne
3 out in experiments with *Tradescantia* microspores (Sax et al, 1955) and have, of course,
4 been subsequently confirmed in an expansive range of studies covering many cell types
5 and species. Some of the most comprehensive studies examining low doses of radiation
6 were those of Lloyd and co-workers (1992). A further prediction from these studies is that
7 over a low dose range (and for low dose rates) the dose-response curve for chromosome
8 aberrations is linear (αD) and time (i.e., dose rate) independent because the one-track
9 mechanism dominates the response. Thus, the linear slope for low dose and low dose rate
10 exposures in this dose range would be the same. This has been borne out in careful
11 studies of the induction of chromosome aberrations over a range of dose rates by
12 Cornforth et al. to specifically test the prediction of a limiting slope at low doses and dose
13 rates (Cornforth et al, 2002).

14 While details of mechanisms involved in the formation of chromosome
15 aberrations remain under investigation, the current view is that the majority of radiation-
16 induced chromosome aberrations are produced by the misrepair of DNA double-strand
17 breaks (DSB) quite possibly those involved in complex DNA lesions (multiply damaged
18 sites). The observations presented above with respect to low dose linearity would support
19 this view. The repair of DSBs (described in Chapter 3) is performed by nonhomologous
20 end-joining and homologous recombination; the former is the prevalent mechanism in
21 mammalian cells. In some cases the pairs of DSB required for the formation of
22 chromosome aberrations by misrepair are produced by one or more electron tracks from a
23 single photon and in others by two or more tracks from different photons. While DSB are
24 generally presumed to be produced linearly with dose for low LET radiations, the
25 probability of conversion into chromosome aberrations is not established. The probability
26 of conversion will depend upon the probability of misrepair and the overall kinetics of
27 DSB repair, and it is likely to be linear with dose given the predictable one-track/two-
28 track nature of the dose-response curve for chromosome aberrations.

29 The development of FISH techniques has allowed for the assessment of the
30 nonlethal reciprocal chromosomal events, i.e. reciprocal translocations and pericentric
31 inversions, as well as complex events involving multiple chromosomal exchanges that
32 would not typically be identified by conventional staining. The dose response for
33 reciprocal translocations is quite similar to that for dicentrics, discussed above, and
34 involves a one-track and a two-track process (Camporoto et al., 2003). Thus, the effects

1 of dose rate and dose fractionation are also similar to those described above for dicentrics.
2 Low dose linearity is observed for acute and chronic low-LET exposures.

3 The “complex exchanges” observed with FISH are often considerably more
4 complex than previously thought. These complex exchanges can involve multiple
5 interactions among several chromosomes. Such complex aberrations constitute a large
6 fraction of aberrations observed after exposure to high LET radiations and that fraction
7 does not appear to vary with dose. For low LET radiation, the fraction of aberrations that
8 are complex is more dose dependent. At relatively high doses (2 to 4 Gy) the fraction is
9 high but at low doses the fraction of aberrations that are complex is substantially lower
10 but still present. More precise data on dose response and dose rate effects for these
11 complex aberrations at low doses will be forthcoming from ongoing studies over the next
12 several years. The mechanisms underlying these complex aberrations are not clear at
13 present and are under investigation. They do appear to involve interactions between sites
14 of complex DNA damage of the type particularly prevalent after exposure to high LET
15 radiation. Such damage is much less prevalent as a result of low LET irradiation, but is
16 still present even at low doses. The significance of these complex exchanges in
17 mutagenesis and carcinogenesis is also unclear. Many of them are probably lethal and
18 therefore not likely to impact such endpoints. However, certain complex aberrations are
19 potentially transmissible and could have a significant impact on mutagenesis,
20 carcinogenesis and the initiation of genomic instability. As a result, the understanding of
21 the mechanisms involved in the development of these complex aberrations may provide
22 important information relevant to low dose risks.

23 Thus, the prevailing view is that chromosome aberrations of all types result from
24 the interactions of pairs (or greater number) of DNA lesions. These lesions can be
25 induced by a single track or by combinations of two or more tracks. However, there is a
26 possible exception to this general rule. Griffin et al. (1996) assessed the efficiency of 1.5
27 keV aluminum x rays at inducing complex chromosome aberrations (requiring three or
28 more interacting lesions for their formation). Based upon the rather high efficiency of
29 this process, the authors suggested that damaged DNA could interact with undamaged
30 DNA to produce some of the aberrations. A proposed mechanism, similar to the
31 production of recombinations during meiosis (Szostak et al., 1983), is not supported by
32 data developed by Cornforth (1990) who concluded that a one-hit exchange probably did
33 not occur, although it could not be ruled out at low doses. The impact of a one-hit
34 exchange process on the shape of the dose-response curve at high doses and the exchange

1 yields at low doses is readily apparent; a steeper slope than that described by the αD
2 component of the linear-quadratic equation would be predicted. The question of its
3 likelihood requires further study.

4 Additional details of the mechanisms of formation of chromosome aberrations and
5 the relevance of their distribution among and within cells to low dose responses can be
6 found in NCRP Report No. 136 (2001). These data and those presented above support
7 the conclusion that at low doses of high- or low-LET radiations the dose-response curves
8 for chromosomal aberrations are linear. Predictions can be made for threshold responses,
9 but the existing data do not support or refute them. The same conclusion applies for
10 supralinear low-dose responses.

11 **4.2 Radiation-Induced Somatic Cell Mutations**

12
13
14 Radiation is capable of inducing a wide spectrum of mutations, from point
15 mutations in single genes to deletions that encompass several physically linked genes
16 (UNSCEAR, 2000). The nature of mutation assays limits the ability to detect large
17 deletions in certain genes because of their close linkage with sequences that are essential
18 for survival of the cell. With this complicating factor in mind, most molecular evidence
19 indicates that DNA deletions resulting in gene loss are the primary events responsible for
20 the mutagenic effects of ionizing radiation (UNSCEAR, 2000). It is also important to
21 note, in this regard, that when data are available, a close relationship between radiation-
22 induced mutations and chromosome aberrations has been found (UNSCEAR, 2000). This
23 spectrum differs from spontaneous mutations, mutations induced by ultraviolet light and
24 many chemical mutagens where the majority of mutations are a result of point mutations
25 (UNSCEAR, 2000). Interestingly, radiation-induced point mutations tend to occur
26 randomly throughout a gene while spontaneous mutations tend to be clustered at specific
27 sites ((Grosovsky et al., 1988; Nelson et al, 1994). The data indicating a predominance of
28 deletion type mutations, and the distribution of point mutations, suggest differences
29 between underlying damage induced by ionizing radiation compared with that from
30 endogenous processes.

31 Mutagenesis is essentially a result of the attempts of the cell to repair damage and
32 analyses of induced mutations can provide clues about mechanisms involved. Sequence
33 analyses of radiation-induced deletion-type mutations have revealed that, as in the case of
34 radiation-induced chromosome aberrations, the mutations are much more complex than

1 originally thought. Deletions often include inversions and insertion of genetic sequences
2 from other chromosomes and frequently involved short direct or inverted DNA repeat
3 sequences ((Morris and Thacker, 1993;Morris et al. 1993;Thacker, 1986). Overall, these
4 analyses support double strand breaks as an important initiating lesion in the pathogenesis
5 of the large deletions characteristic of ionizing radiation and the involvement of DNA
6 DSB repair pathways in the mutagenic process (UNSCEAR, 2000). The presence of
7 repeat sequences suggest that illegitimate recombination associated with double strand
8 breaks is often responsible for the mutagenic process when large deletions are involved.
9 Limited studies with cells defective in specific repair pathways also suggest an important
10 role for DNA DSB repair in the mutagenic effects of ionizing radiation (UNSCEAR,
11 2000). While double strand breaks are more difficult to repair with fidelity than base
12 damage, radiation-induced base damage is also important. It is clear that base damage
13 can often lead to base substitutions (point mutations) and that certain repair pathways
14 involved in base damage repair can be mutagenic.

15 Quantitative studies on dose response relationships for the induction of
16 mutagenesis can be more complicated than studies of chromosome aberrations with
17 considerable variation depending upon the nature of the mutations that can be assayed in
18 each system, genetic background, tolerance for large genetic changes such as deletions,
19 and sensitivity of the system. In systems that have sufficient sensitivity to examine
20 effects at relatively low doses, either linear or linear quadratic dose responses have been
21 reported when a wide dose range has been examined (UNSCEAR, 2000). In either case,
22 in the low dose region, data are consistent with a linear dose response. This linear
23 response is consistent with current models of mechanisms of mutagenesis involving DNA
24 damage and its processing. Such a linear dose response has been observed down to
25 ~200mGy by Thacker et al (1992).

26 Studies of dose rate effects are more complex. In most systems, the effectiveness
27 of low-LET radiation at doses greater than 1 Gy is reduced, at low dose rates, by a factor
28 of 2-4; however there are data in which the effectiveness has remained the same or even
29 increased after low dose rate exposures (Thacker et al , 1992). For example, no dose rate
30 effect or even an inverse dose rate effect is observed in TK6 and other DNA-repair
31 deficient human cells and in many rodent cell lines at very low dose rates (Amundson and
32 Chen 1995, 1996; Vilenchik and Knudson 2000). These dose rate data are consistent with
33 expectations when repair plays a major role in mutagenesis. Cells defective in DNA
34 repair capacity are likely to have little dose rate effect and in fact inverse dose rate effects

1 might be anticipated in cells with defects in damage response pathways at low dose rates
2 (UNSCEAR, 2000; Thacker et al, 1992). A systematic study of this hypothesis would be
3 important in clarifying mutagenic risks following protracted exposures.

4.2.1 Summary

6 The processing and mis-repair of radiation-induced DSBs, particularly complex
7 forms, are principally responsible for chromosome/gene alterations that manifest as
8 chromosome aberrations and mutations. Current understanding of mechanisms and
9 quantitative data on dose and time-dose relationships support a linear dose response at
10 low doses with no compelling evidence for the existence of a threshold dose below which
11 there would be no effect.

4.3 Bystander Effects, Genomic Instability and Adaptive Responses

15 Recently, studies on the induction of so called “bystander effects” in cells not
16 directly irradiated, and the development of genomic instability in the non-irradiated
17 progeny of irradiated cells many generations after exposure, have served to challenge the
18 conventional view that only those cells directly traversed by radiation are targets for
19 cellular effects of radiation including cell killing, and the induction of chromosomal
20 aberrations and mutations. In addition, the assumption that multiple exposures at low
21 doses are additive has come into question as a result of studies demonstrating an adaptive
22 response in certain cells following low dose radiation exposures. The concept of
23 additivity is a result of the view that, following repair, a cell will respond similarly to a
24 second exposure as it did to the first. Studies demonstrating an adaptive response,
25 however, suggest that this may not always be the case; the induction and/or activation of
26 genes likely involved in damage response pathways can influence, positively or
27 negatively, the response to subsequent exposures. If these three phenomena occur *in vivo*,
28 they could impact in particular on the shape of the dose-response curve for low dose, low
29 dose-rate exposures in human populations.

4.3.1 Adaptive Response

32 The adaptive response was first described for chromosomal aberrations (Olivieri
33 et al, 1984). It was observed that pre-exposing cells to a low “priming” dose of radiation
34 appeared to protect these cells from the effects of a second, larger “challenging” dose.

1 This effect was demonstrated most clearly in human lymphocytes, where a decrease of up
2 to 50% in the frequency of aberrations induced by the challenging dose has been observed
3 in cells pretreated with a small priming dose (Wolff, 1996; Sugahara *et al*, 1992). Since
4 the appearance of the initial report over 20 years ago, literally hundreds of reports have
5 been published describing this phenomenon in various experimental systems and for
6 various endpoints including micronucleus formation, mutations and neoplastic
7 transformation; many of these were reviewed in 1994 by the UNSCEAR Committee
8 (UNSCEAR Report, 1994). Despite all of this research, the mechanisms for this
9 phenomenon remain unclear, in contradistinction to the adaptive response to alkylation
10 damage (Lindahl *et al*, 1988). The effect is not consistently seen in all cell types, and
11 there has been considerable donor variation in studies with human lymphocytes.

12 In the earlier studies of the adaptive response to chromosomal aberrations in
13 lymphocytes, low dose-rate exposure from tritiated thymidine was used as a priming
14 dose, though it was later shown that an acute exposure to x-rays would also trigger the
15 effect (Shadley and Wiencke, 1989). Priming doses of 5-100 mGy are generally required
16 to induce the protective effect (Shadley and Wiencke, 1989; Sasaki, 1995). These doses
17 are high enough to produce significant damage in all cells irradiated. Adaptation takes
18 place within 3-6 hours when the cells become resistant to the larger challenge dose,
19 usually 1 Gy or higher. Gap junction mediated intercellular communication has been
20 implicated in this process (Ishii and Watanabe, 1996). The magnitude of the effect
21 depends on many factors including dose, dose-rate, cell and tissue type and the endpoint
22 measured.

23 The mechanisms for the effect remain unclear. It is now known that low doses of
24 radiation can modulate the expression of a variety of genes (e.g., Hallahan *et al*, 1991;
25 Leskov *et al*, 2001; Sasaki *et al*, 2002). Sasaki *et al* (2002) found that p53 appeared to
26 play a key role in the adaptive response while the DNA-PKcs, ATM and FANCA genes
27 were not involved. They proposed that the adaptive response and apoptosis constitute a
28 complementary defense mechanism. It has also been reported that the induction of heat
29 shock proteins may be involved in the adaptive response (Lee *et al*, 2002; Kang *et al*,
30 2002).

31 While it has been hypothesized that the phenomenon reflects the induction of
32 some type of DNA repair process that requires a certain level of damage in the cell, no
33 such inducible DNA repair mechanism for DNA strand breaks has been clearly
34 demonstrated in mammalian cells. Restriction enzymes that produce DNA double strand

1 breaks will induce adaptation in human lymphocytes (Wolff, 1996), and the rate of repair
2 has been reported to be more efficient in adapted cells (Ikushima *et al*, 1996). Evidence
3 has been presented to suggest the involvement of DNA repair in the adaptive response in
4 yeast (Dolling *et al*, 2000), and Haber and colleagues (personal communication) have
5 shown that when a single DSB is introduced in budding yeast cells synchronized in G₁,
6 the cells become significantly resistant to a challenge dose of MMS applied during the
7 discrete period approximately 6 hours later when repair is taking place. It is of interest in
8 this context that the inducible repair of thymine glycols by the base excision repair
9 process has been described (Le *et al*, 1998). Generally, however, DNA base damage is
10 not thought to be the principal mechanism for the induction of mutations and
11 chromosomal aberrations by ionizing radiation. It has also been proposed that the
12 priming dose may lead to persistent free radical activation as part of the post-irradiation
13 cellular stress response that includes the up-regulation of genes associated with signal
14 transduction and cell cycle control (Bravard *et al*, 1999).

15 A number of reports have presented evidence for an adaptive response for the
16 induction of specific gene mutations (Sanderson and Morley, 1986; Kelsey *et al*, 1991;
17 Zhou *et al*, 1994; Rigaud *et al*, 1995). In general, the mutation frequencies induced by
18 relatively high radiation doses have been shown to be decreased by approximately 50% if
19 the exposure is preceded by a priming dose of approximately 10 mGy 5 to 24 hours
20 previously. These experiments have been carried out in various different systems, though
21 generally but not exclusively with cells of lymphoid origin (lymphocytes, established
22 lymphoblastoid cell lines and a human T-cell leukemia cell line). The adaptive exposure
23 to radiation may also decrease the frequency of neoplastic transformation either arising
24 spontaneously or induced by a subsequent high radiation dose (Azzam *et al*, 1994;
25 Redpath and Antoniono, 1998; Redpath *et al*, 2001; 2003). Adaptive responses have
26 been described in human tumor cells with irradiation protocols closely resembling clinical
27 applications (Smith and Raaphorst, 2003).

28 Evidence is emerging for the occurrence of adaptive phenomena *in vivo*. These
29 include the induction of leukemia and lymphoma (Ishii *et al*, 1996; Bhattacharjee and Ito,
30 2001; Mitchel *et al*, 1999; 2003), as well as teratogenic effects and the development of
31 heritable germline mutations (Somers *et al*, 2002). In one study (Bhattacharjee, 1996),
32 pre-irradiating mice with five repeated exposures of 10 mGy a day appeared to reduce
33 significantly the incidence of thymic lymphoma induced by a challenge dose of 2 Gy. It
34 has been reported that short-term low dose occupational exposures may act as an *in vivo*

1 adaptive dose for the induction of micronuclei by *in vitro* irradiation of lymphocytes
2 (Thierens *et al*, 2002).

3 The adaptive response shares some similarities with the phenomenon of “low dose
4 hypersensitivity” described by Joiner and his colleagues (Joiner *et al*, 1996) based on the
5 multiphasic shape of the single dose survival curve for some mammalian cell lines. They
6 have observed a steep decline in cell survival in the low dose range, followed by a plateau
7 which they hypothesize represents induced radioresistance. In a recent study (Short *et al*,
8 2001), cells displaying a strong hypersensitivity response showed increased killing
9 following multiple low dose exposures. Similar to the adaptive response, it has been
10 proposed that the phenomenon may represent the manifestation of inducible processes
11 facilitating the repair of DNA damage (Joiner *et al*, 2001; Marples and Joiner, 2000). In
12 two quite different experimental systems for the study of malignant transformation *in*
13 *vitro*, evidence has been presented that the spontaneous transformation frequency is
14 actually reduced by very small doses of radiation (doses as low as 1 mGy) (Azzam *et al*,
15 1996; Redpath *et al*, 2001; 2003). The frequency of transformation rose rapidly at higher
16 doses.

17 Despite such provocative findings, there are still many questions concerning the
18 adaptive response (Wolff, 1998; Stecca and Gerba, 1998). The response for
19 chromosomal damage has been shown to vary with the donor, some individuals being
20 unresponsive and others showing a synergistic effect (Bosi and Olivieri, 1989). The same
21 is true for different cellular systems and for other biological endpoints such as cell
22 survival (Boothman *et al*, 1996; Short *et al*, 1999; Sorensen *et al*, 2002). In the absence
23 of firm knowledge of molecular mechanisms, it is difficult to evaluate the potential
24 significance of the adaptive response for the risk from exposure to ionizing radiation in
25 human populations. Clearly, the phenomenon appears to be a real one in many cellular
26 systems, one that could influence the response to protracted radiation exposure. It will be
27 important, however, to determine the extent to which it is active *in vivo* at relevant dose
28 and dose-rate levels for human exposures before it can be considered as a factor in risk
29 estimation.

30 Adaptive responses including those in relation to radiation induced cancer and
31 stimulatory effects on the immune system were comprehensively reviewed by UNSCEAR
32 in 1994 (UNSCEAR 1994) and some aspects were revised in UNSCEAR (2000). The
33 general conclusion from these reports was that there was insufficient information on the
34 role and mechanisms of adaptive responses to influence judgments on low dose cancer

1 risk. Recent animal carcinogenesis studies relating to adaptive responses (Mitchel et al
2 1999, 2003) raise the possibility that adaptive-like responses may increase tumour latency
3 whilst not affecting life-time risk. These data are of scientific interest but remain of
4 rather uncertain relevance to radiological protection.

6 **4.3.2 Radiation Induced Genomic Instability**

7 The term radiation-induced genomic instability refers to a phenomenon observed
8 in a number of different cellular systems whereby radiation exposure appears to induce a
9 type of instability in individual cells that is transmitted to their progeny, leading to a
10 persistent enhancement in the rate at which genetic changes arise in the descendants of
11 the irradiated cell after many generations of replication. The genetic endpoints studied
12 have included malignant transformation, chromosomal aberrations, specific gene
13 mutations, and cell survival. Typically, this phenomenon has been studied by examining
14 the occurrence of such genetic effects in clonal populations derived from single cells
15 surviving radiation exposure (Little, 2003), although some studies have relied upon the
16 post-irradiation analysis of cells in mass culture rather than clonal isolates..

17 Early evidence for the existence of such a phenomenon was derived from an
18 examination of the kinetics of radiation-induced malignant transformation of cells *in vitro*
19 (Sinclair, 1964; Kennedy *et al*, 1980; Kennedy and Little, 1984). These results suggested
20 that transformed foci did not arise from a single, radiation-damaged cell. Rather,
21 radiation appeared to induce a type of instability in 20-30% of the irradiated cell
22 population; this instability enhanced the probability of the occurrence of a second,
23 neoplastic transforming event. This second event was a rare one, occurring with the
24 frequency of approximately 10^{-6} , and involved the actual transformation of one or more of
25 the progeny of the original irradiated cells after many rounds of cell division. This
26 transforming event occurred with the constant frequency per cell per generation, and had
27 the characteristics of a mutagenic event (Kennedy *et al*, 1984). Thus, neoplastic
28 transformed foci did not appear to arise from the original irradiated cell but rather from
29 one or more of its progeny. These findings were consistent with the hypothesis that
30 radiation induces genetic instability in cells that enhances the rate at which malignant
31 transformation or other genetic events occur in descendants of irradiated cells after many
32 generations of cell replication.

33 This hypothesis has subsequently been confirmed in a number of experiment
34 systems for various genetic endpoints (Morgan *et al*, 1996; Little, 1998; Baverstock,

1 2000; Romney *et al*, 2001a; Morgan, 2003a). In terms of mutagenesis, approximately
2 10% of clonal populations derived from single cells surviving radiation exposure showed
3 a significant elevation in the frequency of spontaneously arising mutations as compared
4 with clonal populations derived from non-irradiated cells (Chang and Little, 1992; Little
5 *et al*, 1997). This increased mutation rate persisted for approximately 30 generations
6 post-irradiation then gradually subsided. Interestingly, the molecular structural spectrum
7 of these late-arising mutants resembles those of spontaneous mutations in that the
8 majority of them are point mutations (Grosovsky *et al*, 1996; Little *et al*, 1997),
9 indicating that they arise by a different mechanism from that of direct x-ray-induced
10 mutations which involve primarily deletions. An enhancement of both minisatellite (Li *et*
11 *al*, 1992) and microsatellite (Romney *et al*, 2001b) instability has also been observed in
12 the progeny of irradiated cells selected for mutations at the *thymidine kinase* locus, further
13 evidence that a subpopulation of genetically unstable cells arises in irradiated populations.
14 It is of interest that instability as measured in minisatellite sequences of x-ray-transformed
15 mouse 10T½ cells was markedly enhanced when the cells were grown *in vivo* as
16 compared to prolonged cultivation *in vitro* (Paquette and Little, 1994).

17 An enhanced frequency of non-clonal chromosomal aberrations was reported in
18 clonal descendants of mouse hematopoietic stem cells examined 12-14 generations after
19 exposure to alpha radiation (Kadhim *et al*, 1992). Persistent radiation-induced
20 chromosomal instability has since been demonstrated in a number of other cellular
21 systems (Sabatier *et al*, 1992; Holmberg *et al*, 1993; Marder and Morgan, 1993; Kadhim
22 *et al*, 1995; Little *et al*, 1997; Ponnaiya *et al*, 1997; McIlrath *et al*, 2003). Susceptibility
23 to radiation-induced chromosomal instability differs significantly among cells from
24 different strains of mice (Watson *et al*, 1996a; Ponnaiya *et al*, 1997), and similar
25 differences in genetic susceptibility to radiation-induced chromosomal instability have
26 been observed in primary human fibroblasts (Kadhim *et al*, 1998).

27 It is now clear that genomic instability, both chromosomal and mutational
28 instability, can be induced by high or low LET radiation (Little *et al*, 1997; Belyakov *et*
29 *al*, 1999; Limoli *et al*, 2000; Evans *et al*, 2001), and in most normal and transformed
30 human and rodent cases as described above. The fact that Dugan and Bedford (2003)
31 found no evidence for induced chromosomal instability in a normal human diploid
32 fibroblast strain may be related to genetic factors as described by Kadhim *et al* (1998),
33 who observed variability in the response of different strains of human diploid fibroblasts.
34 Furthermore, delayed reactivation of p53 and a persistent induction of reactive oxygen

1 species has been reported in normal human fibroblasts (Rugo *et al*, 2003) as well as in
2 human fibrosarcoma cells (Suzuki *et al*, 2003). Long-term instability can be induced by
3 irradiation of cells with single alpha particles from a focused microbeam (Kadhim *et al*,
4 2001), supporting earlier observations that the instability phenotype can be activated by
5 low radiation doses, becoming saturated at higher doses (Kadhim *et al*, 1995; Grosovsky
6 *et al*, 1996; Little *et al*, 1997).

7 Finally, a persistently increased rate of cell death has been shown to occur in cell
8 populations many generations after irradiation (Seymour *et al*, 1986; Chang and Little,
9 1991; Belyakov *et al*, 1999). This phenomenon has been variously referred to as
10 occurring as a result of “lethal mutations” or “delayed reproductive failure”, but has been
11 measured as a reduction in the ability of cells to attach and form macroscopic colonies in
12 a classic clonogenic survival assay. In some cellular systems, an increased rate of
13 apoptotic cell death has been shown to accompany this phenomenon (Jamali and Trott,
14 1996; Limoli *et al*, 1998; Belyakov *et al*, 1999). Persistent reproductive failure has been
15 linked to chromosomal instability (Limoli *et al*, 1998) and malignant transformation
16 (Lewis *et al*, 2001; Redpath and Gutierrez, 2001), and evidence presented to suggest that
17 DNA is at least one of the critical targets in the initiation of this phenomenon (Limoli *et*
18 *al*, 1999). Instability was attenuated by treating the irradiated cells with free radical
19 scavengers or allowing potentially lethal damage to be repaired by confluent holding prior
20 to analyzing the subsequent development of chromosomal instability (Limoli *et al*, 2001).
21 It has been proposed that oxidative stress perhaps consequent to enhanced, p53-
22 independent apoptosis may contribute to the perpetuation of the instability phenotype in
23 these populations (Limoli *et al*, 1998; Redpath and Gutierrez, 2001).

24 Of importance in terms of radioprotection is whether this phenomenon occurs *in*
25 *vivo* and thus may be related to the induction of cancer. A number of mouse models for
26 genetic instability have been described (Reliene and Schiestl, 2003). The transmission of
27 chromosomal instability *in vivo* has been reported in several distinct experimental models
28 (Pampfer and Streffer, 1989; Watson *et al*, 1996b; Watson *et al*, 2001; Ullrich and Davis,
29 1999), though not in others (Bouffler *et al*, 2001a), and *in vivo* aspects of transmissible
30 instability are addressed in detail later in this report. Evidence for transmissible
31 instability in irradiated human populations is inconsistent (Nakanishi *et al*, 2001;
32 Whitehouse and Tawn, 2001). While it has been suggested that instability induced in X-
33 irradiated mouse hematopoietic stem cells may be related to the occurrence of the non-
34 specific genetic damage found in radiation-induced leukemias in these mice (MacDonald

1 *et al*, 2001), other work from the same laboratory indicates that susceptibility to radiation-
2 induced leukemia/lymphoma is generally separable from sensitivity to induced genomic
3 instability (Boulton *et al*, 2001).

4 In the case of murine mammary tumors induced by radiation, the link
5 between genomic instability and early events in mammary cancer development appears
6 stronger (Ullrich and Davis, 1999; Okayasu *et al.*, 2000). In this instance, the instability
7 appears to be directly related to a defect in the function of the DNA repair enzyme DNA-
8 PKcs.

10 **4.3.3 The Bystander Effect in Irradiated Cell Populations**

11 The bystander effect of radiation refers to the evidence that damage signals may
12 be transmitted from irradiated to non-irradiated cells in a population, leading to the
13 occurrence of biological effects in cells that receive no radiation exposure. The use of
14 this term has been interpreted broadly, however, as is evidenced by the experimental
15 protocols employed to study such effects *in vitro*. The first protocol employs monolayer
16 cultures of mammalian cells in which a small fraction of the cells in the population are
17 irradiated, generally by alpha particles, and the biological effect examined in the non-
18 irradiated, neighboring cells. A corollary protocol involves mixing experiments in which
19 irradiated cells are mixed with non-irradiated cells and the biologic effect subsequently
20 measured in the non-irradiated cohort of the population. The second protocol involves
21 the harvesting of conditioned medium from irradiated cultures and incubating this with
22 non-irradiated cells; the bystander cells are thus not in physical proximity to the irradiated
23 cells. Both mixing and medium transfer techniques permit the examination of effects with
24 low LET as well as high LET radiations.

25 The experimental model employed in many of these studies has involved the
26 exposure of monolayer cultures of mammalian cells, often confluent or sub-confluent, to
27 very low fluences of alpha particles, fluences whereby only a very small fraction of the
28 nuclei in a cell population will actually be traversed by an alpha particle. This may be
29 accomplished by irradiation from an external source of alpha particles (Metting *et al*,
30 1995) or by use of precision microbeam irradiators whereby specific cells can be targeted
31 (Hei *et al*, 1997; Prise *et al*, 1998; Prise *et al*, 2000; Folkard *et al*, 2001; Shao *et al*
32 (2003a). A grid arrangement has also been employed to protect many cells in a
33 population exposed to relatively high fluences of alpha particles (Lorimore *et al*, 1998).

1 The first evidence for this phenomenon was derived from studies of the induction
2 of sister chromatid exchanges (SCE) by very low fluences of alpha particles from an
3 external source (Nagasawa and Little, 1992). It was observed that an enhanced frequency
4 of SCE occurred in 20-40% of the cells exposed to fluences whereby only about 1/1000
5 to 1/100 cell nuclei were actually traversed by an alpha particle. This finding was later
6 confirmed and evidence presented to suggest that the phenomenon involved secretion of
7 cytokines or other factors by irradiated cells leading to the up-regulation of oxidative
8 metabolism in bystander cells (Deshpande *et al*, 1996; Narayanan *et al*, 1997; Lehnert
9 and Goodwin, 1997; Narayanan *et al*, 1999). It has since been shown that an enhanced
10 frequency of specific gene mutations occurs in bystander cells in populations exposed to
11 very low fluences of alpha particles (Nagasawa and Little, 1999). As a result, the induced
12 mutation frequency per alpha particle track increases at low fluences where bystander as
13 well as directly irradiated cells are at risk for the induction of mutations. This leads to a
14 dose-response curve in which the slope is initially steeper than it is at higher doses.
15 Studies with microbeam irradiation have provided evidence for an enhanced frequency of
16 micronucleus formation, cell killing and apoptosis in bystander cells (Prise *et al*, 1998;
17 Prise *et al*, 2000; Belyakov *et al*, 2001; Schettino *et al*, 2003; Shao *et al*, 2003a), as well
18 as an enhanced frequency of mutations (Zhou *et al*, 2000; Zhou *et al*, 2001) and
19 malignant transformation (Sawant *et al*, 2001a).

20 It has also been shown that changes in gene expression occur in bystander cells in
21 monolayer cultures; the expression levels of p53, p21^{Waf1}, CDC2, cyclin-B1 and rad51
22 were significantly modulated in non-irradiated cells in confluent human diploid cell
23 populations exposed to very low fluences of alpha particles (Azzam *et al*, 1998). These
24 experiments were carried out by western blotting and *in situ* immunofluorescence staining
25 techniques utilizing confocal microscopy; although only about 1-2% of the cell nuclei
26 were actually traversed by an alpha particle, clusters of cells showed enhanced expression
27 of p21^{Waf1}. This phenomenon involved cell-to-cell communication via gap junctions
28 (Azzam *et al*, 1998; 2001), as has also been shown for micronucleus formation (Shao *et al*
29 *et al*, 2003b) and mutations (Zhou *et al*, 2001). It appears that radiation exposure itself can
30 enhance intercellular communication as evidenced by an up-regulation of Connexin 43
31 (Azzam *et al*, 2003a). Evidence for DNA damage in bystander cells was provided by
32 examining micronucleus formation, a surrogate measure of DNA damage; that the up-
33 regulation of the p53 damage response pathway in bystander cells was a consequence of
34 this DNA damage is supported by the observation that p53 was phosphorylated on serine

1 15 (Azzam *et al*, 2001). Interestingly, it has been hypothesized that the apparent
2 persistence of DNA double strand breaks after very low dose x-ray exposure might be the
3 result of such a bystander effect (Rothkamm and Lobrich, 2003).

4 DNA damage in bystander cells, however, appears to differ from that occurring in
5 directly irradiated cells; whereas the mutations induced in directly irradiated cells were
6 primarily partial and total gene deletions, over 90% of those arising in bystander cells
7 were point mutations (Huo *et al*, 2001). This would be consistent with the evidence that
8 oxidative metabolism is up-regulated in bystander cells (Narayanan *et al*, 1997; Azzam *et al*,
9 2002), and has led to the hypothesis that the point mutations are a result of oxidative
10 base damage occurring in bystander cells (Huo *et al*, 2001). A similar mechanism has
11 been proposed for the observation that localized cytoplasmic exposure from a microbeam
12 irradiator led to a significant increase in the frequency of point mutations which appeared
13 to involve the generation of reactive oxygen species (Wu *et al*, 1999; Shao *et al*, 2004)).
14 Bystander cells defective in the non-homologous end joining pathway including mouse
15 knockout cell lines for Ku80, Ku70 and DNA-PKcs are extremely sensitive to the
16 induction of mutations and chromosomal aberrations (Nagasawa *et al*, 2003; Little *et al*,
17 2003). Interestingly, the mutations in these repair deficient bystander cells were primarily
18 the result of partial and total gene deletions (Nagasawa *et al*, 2003), whereas those in wild
19 type bystander cells were predominantly point mutations. The marked sensitization of
20 repair-deficient bystander cells to the induction of mutations and chromosomal
21 aberrations may be a consequence of unrejoined DNA double strand breaks occurring as a
22 result of clustered damage arising from opposed oxidative lesions and single strand
23 breaks. Mutations in wild-type cells arise primarily from oxidative base damage.

24 In earlier studies, it was reported that alpha particle irradiation could induce the
25 intracellular generation of reactive oxygen species (ROS) including the superoxide anion
26 and hydrogen peroxide (Narayanan *et al*, 1997). This ROS response did not require direct
27 nuclear irradiation, as an ROS response was induced in non-irradiated cells with
28 conditioned medium from alpha irradiated cells. The various studies examining the role
29 of oxidative metabolism and gap junction mediated intercellular communication have
30 been summarized by Azzam *et al* (2003b). The role of oxidative stress in modulating
31 signal transduction and micronucleus formation in bystander cells was examined in
32 confluent monolayer populations of human diploid cells exposed to low fluences of alpha
33 particles (Azzam *et al*, 2002). The results support the hypothesis that superoxide and
34 hydrogen peroxide produced by flavin containing oxidase enzymes mediate the activation

1 of several stress inducible signaling pathways as well as micronucleus formation in
2 bystander cells. These include the p53 damage response pathway as well as the MAP
3 kinase family of signaling pathways. It has also been reported that nitric oxide may
4 initiate intercellular signal transduction pathways influencing the bystander response to
5 radiation (Matsumoto *et al*, 2001; Shao *et al*, 2002). It thus appears that ROS may be the
6 primary mediators of the bystander effect (Szumiel, 2003).

7 Interestingly, this up-regulation of oxidative stress in bystander cells is
8 reminiscent of the effect associated with radiation-induced genomic instability (Redpath
9 and Gutierrez, 2001; Limoli *et al*, 2001), and it has been proposed that the bystander
10 effect may be related to the induction of an inflammatory-type response *in vivo* (Lorimore
11 *et al*, 2001). The activation of MAP K proteins and their downstream effectors in
12 bystander cells (Azzam *et al*, 2002) is of particular interest in terms of the recent
13 observation that membrane signaling pathways are involved in the bystander effect in
14 monolayer cultures (Nagasawa *et al*, 2002; Shao *et al*, 2004).

15 Bishayee *et al* (1999) and Howell and Bishayee (2002) developed a three-
16 dimensional tissue culture model which utilized Chinese hamster V79 cells to study
17 bystander effects caused by non-uniform distributions of radioactivity. Cells labeled with
18 ¹²⁵I-Urd were mixed with unlabelled cells and multicellular clusters formed by
19 centrifugation. A decrease in clonogenic survival occurred among the unlabelled cells
20 which, based on inhibitor studies, appeared to depend upon gap junction mediated
21 intercellular communication (Bishayee *et al*, 2001). On the other hand, when cells
22 irradiated with carbon beams were co-cultured with non-irradiated cells, cloning
23 efficiency and proliferation of the non-irradiated recipient cells was increased (Shao *et al*,
24 2003c), reminiscent of the well known feeder layer effect. When a mixture of ¹²⁵I-labeled
25 and unlabeled human tumor cells were injected into nude mice, a distinct inhibitory effect
26 on the growth of the unlabeled cells was observed (Xue *et al*, 2002). Belyakov *et al*
27 (2003) have presented evidence for a bystander effect in a primary tissue explant model.
28 Watson *et al* (2000) transplanted a mixture of irradiated and non-irradiated bone marrow
29 cells in a mouse system that allowed the discrimination between irradiated donor stem
30 cell-derived cells and non-irradiated stem-cell derived cells *in vivo*. They were able to
31 demonstrate chromosomal instability in the progeny of the non-irradiated hematopoietic
32 stem cells, providing a link between a bystander effect of ionizing radiation and the
33 induction of genomic instability *in vivo*.

1 There is a long history of the apparent induction of clastogenic factors by
2 radiation, primarily as measured in the plasma of irradiated individuals. These studies are
3 reviewed in detail by Mothersill and Seymour (2001). These workers have reported that
4 the exposure of cells in culture or explants of tissue to gamma radiation doses as low as
5 10 mGy can lead to the release of factors into the medium by the irradiated cells; when
6 this conditioned medium is transferred to non-irradiated cells, their cloning efficiency is
7 reduced associated with increased levels of apoptotic cell death (Mothersill and Seymour,
8 1998). This phenomenon has been associated with early changes in mitochondrial
9 membrane permeability and the induction of reactive oxygen species (ROS) (Lyng et al,
10 2001).

11 Overall, however, a clear picture has yet to emerge from the experience with
12 medium transfer experiments. There is convincing evidence that factors are released into
13 the medium by irradiated cells that can lead to changes in the viability of non-irradiated
14 cells incubated with such conditioned medium. The results from different laboratories,
15 however, are not entirely consistent. Some workers report that incubation with
16 conditioned medium harvested from irradiated cultures leads to a reduction in cloning
17 efficiency of the recipient cells (Lyng *et al*, 2002; Sawant *et al*, 2002), while others find it
18 is enhanced (Iyer and Lehnert, 2002; 2002) or dependent on cell type (Mothersill and
19 Seymour, 1997). The effect of medium irradiation alone is particularly controversial
20 (Lehnert and Goodwin, 1997; Belyakov *et al*, 2001; Zhou *et al*, 2002). In terms of
21 genetic effects, one laboratory describes a bystander effect for sister chromatid exchanges
22 in conditioned medium transfer experiments (Lehnert and Goodwin, 1997), whereas
23 another finds little or no evidence for a bystander mutagenic effect under similar
24 conditions (Zhou *et al*, 2002). The effect appears likely to be mediated by cytokines or
25 reactive oxygen species, but the exact nature of the factor or factors responsible for the
26 biological effects in the non-irradiated, bystander cells remains to be elucidated.

27 In sum, the results of these studies of bystander effects indicate clearly that
28 damage signals can be transmitted from irradiated to non-irradiated cells. In confluent
29 monolayer cultures, this phenomenon involves gap junction mediated cell to cell
30 communication, and appears to involve both the induction of reactive oxygen species and
31 the activation of extra-nuclear signal transduction pathways. Preliminary evidence
32 suggests a role for membrane signaling. Multiple biological effects may occur in
33 bystander cells including cell killing, the induction of mutations and chromosomal
34 aberrations, and the modulation of gene expression. Some evidence suggests that

1 regulation of the p53 damage response pathway may be central to this phenomenon.
2 Damage signals may in addition be transmitted through the extracellular medium, also
3 appearing to involve the production of reactive oxygen species. Finally, preliminary
4 studies with tissue explant models and a mouse bone marrow stem cell transplant system
5 suggests that the effect may occur *in vivo*.

6 7 **4.4 Conclusions: Implications for Risk Assessment** 8

9 There is increasing evidence that the development of invasive metastatic cancer
10 involves a series of distinct genetic events some of which can be associated with specific
11 stages in the carcinogenic process (Fearson and Vogelstein, 1990). A question that arises
12 is how as many as six to eight such genetic events may accumulate in a single cell
13 lineage, given that the prevalence of most mutations is about 10^{-5} . Loeb *et al* (2003) and
14 others have postulated that early in the process of carcinogenesis a mutation may arise in
15 a gene that is important in maintaining genomic stability, yielding a cell lineage with a
16 mutator phenotype. This phenotype would enhance the frequency with which
17 spontaneous mutations arise in these cells, and thus facilitate the accumulation of the
18 requisite number of genetic events to produce a cancer. One such example is hereditary
19 non-polyposis colon cancer which is associated with a germline defect in DNA mismatch
20 repair. While genomic instability is a hallmark of tumor cells, most types of cancer have
21 not been associated with specific DNA repair defects.

22 The finding that radiation itself may induce an instability phenotype has thus
23 attracted considerable interest. It would suggest that the initial radiation-induced event
24 may be a frequent one involving as many as 10-20% of the population, rather than a rare
25 mutagenic event. This increased level of instability which is transmissible over many
26 generations of cell replication would thus enhance the rate at which multiple genetic
27 events important to the development of cancer would arise in the cell population. It is not
28 yet clear, however, the extent to which this radiation-induced phenomenon may be of
29 importance in carcinogenesis. The fact that it appears to saturate at fairly low doses (of
30 the order of 100-500 mGy) implies that it could influence the extrapolation to low dose
31 effects. On the other hand, as it may not represent an irreversible carcinogenic event such
32 as mutation, it might be susceptible to modulation by external factors. Clearly, additional
33 research is needed to determine the mechanisms involved in radiation-induced genomic
34 instability, in terms of both the initiating event and how the effect is transmissible for

1 many generations of cell replication, before its implications for the assessment of the
2 carcinogenic risk of low dose, low dose-rate exposure to ionizing radiation can be
3 clarified.

4 Another area where this phenomenon could well be of significance involves
5 potential transgenerational effects of irradiation. The sum of the available evidence
6 suggests that such instability is induced in the germ cells of irradiated parents and in the
7 offspring born to them (Niwa, 2003). If exposure to low levels of ionizing radiation thus
8 induces the instability phenotype in germ cells of the offspring of irradiated parents, it is
9 entirely feasible that this instability could increase their susceptibility to cancer or other
10 genetic effects. For example, Pils *et al* (1999) reported that genomic instability
11 manifested by lethal and teratogenic effects may be passed on to two successive
12 generations of offspring in mice after irradiation of the zygote, while Niwa and
13 Kominami (2001) and Dubrova and his colleagues (Dubrova *et al*, 1998; Dubrova and
14 Plumb, 2002) presented evidence for transmissible germline instability at mouse
15 minisatellite loci. There is preliminary experimental evidence to suggest that such
16 transmissible instability might lead to increased susceptibility to the induction of tumors
17 in the offspring of irradiated mice (Nomura, 1982; Lord *et al*, 1998). The question of
18 radiation-related transgenerational cancer risk in experimental animals and human
19 populations is discussed above, in Section 2.1.1; the induction of transmissible genomic
20 instability by radiation in germ cells would provide a mechanism for such
21 transgenerational effects.

22 The bystander effect has clear implications in terms of human exposures to very
23 low fluences of high LET particulate radiation, such as alpha particles from
24 environmental radon or densely-ionizing galactic cosmic rays in space (Brenner and
25 Elliston, 2001). In the case of radon, for example, only a small fraction of a person's
26 bronchial epithelial cells, the presumed target for lung cancer, will be hit each year by an
27 alpha particle arising from residential radon exposure. In the past, the genetic or
28 carcinogenic risk has been assumed to be related directly to the number of cell nuclei
29 actually traversed by an alpha particle, thus yielding a linear dose response relationship.
30 The evidence that irradiated cells may transmit damage signals to neighboring non-
31 irradiated cells that result in genetic alterations in these "bystander" cells would invalidate
32 this assumption. Rather, it would suggest that the dose-response curve may be non-linear
33 at low mean doses yielding a greater effect than that predicted on the basis of the dose
34 received by individual cells at low alpha particle fluences. Preliminary data, based

1 primarily on cell mixing experiments, are emerging to suggest that a bystander response
2 also occurs with low-LET radiation. However, these preliminary data are at present
3 insufficient to draw any conclusions concerning the significance of this effect at low
4 radiation doses, particularly at levels such that the track fluence is less than the number of
5 cells in the radiation field (J. B. Little, personal communication).

6 Evidence for the convergence of the three phenomena (adaptive response,
7 genomic instability, and bystander effects) is also of interest (Lorimore and Wright, 2003;
8 Morgan, 2003b). Several different studies involving both *in vitro* and *in vivo* assays have
9 shown, for example, that transmissible genomic instability may arise in bystander cells
10 (Lorimore *et al*, 1998; Watson *et al*, 2000), and that the bystander effect may be
11 modulated by the adaptive response (Iyer and Lehnert, 2002; Mothersill *et al*, 2002;
12 Sawant *et al*, 2002b; Zhou *et al*, 2003). Defects in the non-homologous end joining DNA
13 repair pathway have been associated with both radiation-induced genomic instability
14 (Okayasu *et al*, 2000) and the bystander effect (Little *et al*, 2003). It has been reported
15 that conditioned medium from certain (but not all) unstable clones harvested many cell
16 generations post-irradiation is highly cytotoxic to unirradiated cells (Nagar *et al*, 2003).
17 Finally, oxidative stress manifested by enhanced levels of reactive oxygen species has
18 been implicated in all three phenomena.

19 When considered as a whole, the emerging results with these three phenomena
20 raise the possibility that the dose response at low doses of ionizing radiation is uncertain,
21 and a simple extrapolation from high dose effects may not be appropriate. In some cases,
22 such as the induction of mutations by exposure to very low fluences of high LET
23 particles, or as reported for the cytotoxic effects of very low doses of x-rays, the effect
24 may be greater than predicted from a linear extrapolation. On the other hand, certain
25 studies of malignant transformation have revealed a reduced effect for very low doses.
26 Overall, however, these findings imply that the biological effects of radiation in cell
27 populations may not be restricted to the response of individual cells to the DNA damage
28 they receive, but rather that tissues respond as a whole. These three phenomena are of
29 importance as they may influence in particular the nature of the dose response
30 relationship at low doses and low dose-rates. However, a better understanding of the
31 mechanisms for these phenomena, the extent to which they are active *in vivo*, and how
32 they are interrelated is needed before they can be confirmed as factors to be included in
33 the estimation of potential risk to the human population of exposure to low levels of
34 ionizing radiation.

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2

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5. CARCINOGENIC EFFECTS OF IONIZING RADIATION

5.1 Mechanisms of Radiation-Induced Cancer

Studies on the cellular and molecular mechanisms of carcinogenesis over the last several years have provided substantial insight with respect to the complex multi-step nature of the process of neoplastic development (Hanahan 2000; UNSCEAR 2000). Such studies have identified a number of specific target genes and gene pathways and also important variations among different tumor types. From such studies tumor development is now generally viewed as a multi-step clonal process of cellular evolution and selection. The conversion of a normal somatic cell into a cell with neoplastic potential is generally referred to as initiation (UNSCEAR 2000; Knudson 2001) . Subsequent to initiation, the process of neoplastic development continues via the progression phase. This phase includes clonal selection and the development of additional mutational events. As such, this stage may be viewed as the early developmental and evolutionary phases of an initiated cell during neoplastic progression. Factors such as cell-cell communication, mitogenic stimulation, cellular differentiating factors, mutational processes and cell-tissue interactions all play a role in determining the probability of progression of initiated cells. Specific genetic changes involved in progression often differ among tissue types, although key related pathways are generally involved (Hanahan 2000; UNSCEAR 2000). The end phase in tumor progression is the conversion of a cell or cells to the malignant phenotype. Because of the high degree of instability associated with such cells, progression and evolution within a population of malignant cells will continue indefinitely (Loeb 1991). Overall, it is clear that only a small fraction of cells that enter the pathway of neoplastic development as initiated cells will complete the full sequence of events leading to malignancy, a process that can require years in the human being.

Although radiation-induced tumorigenesis in experimental animals and in humans has been the subject of intense study for many years, until recently direct evidence with respect to underlying mechanisms of radiation carcinogenesis has been lacking and models have relied heavily on indirect inferential data. For example, it has been suggested for many years that the primary effect of radiation is principally on early events, i.e. the primary effect of radiation is as a tumor initiating agent. This is based on several observations. First, animals and human beings are generally more sensitive to the tumorigenic effects of ionizing radiation at young compared to older ages. This suggests that radiation effects have more to do with tumor initiation than with promotional effects

1 that accelerate the development of pre-existing neoplasms (UNSCEAR 2000; Clifton
2 1986; Fry 1977; Fry 1987; Fry 1992). Second, experimental animal data from studies of
3 skin cancer development, specifically designed to examine the influence of radiation on
4 different stages of tumorigenesis, show that radiation only weakly promotes the
5 development and progression of chemically-initiated tumors but has significant initiating
6 activity (Jaffe 1987). Finally, it is observed in humans and animals that single acute
7 doses of low-LET radiation are sufficient to produce a dose-dependent increase in cancer
8 risk and that, in quantitative animal studies, dose protraction decreases that risk. The last
9 observation also supports the inference that the major effect of radiation is on early events
10 in the carcinogenic process (IARC 2000; Hanahan 2000). While this inference appears to
11 be logically based, until recently there has been no direct evidence in support of it.

12 Advances in cell biology, cytogenetics, molecular biology and mouse genetics
13 over the past several years have enabled more direct investigation of events in the
14 tumorigenic process following radiation exposure. Such studies, by linking specific cell
15 and molecular effects directly to the tumorigenic process, provide valuable insights into
16 mechanisms as well as a better understanding of potential radiation-related risks. Of
17 particular importance in this regard have been animal studies using newly developed
18 models, both in inbred strains of mouse and rat and in genetically engineered rodents.
19 Quantitative studies using mouse and rat models for radiation-induced mammary cancer
20 and for thyroid cancer in rats have now provided direct evidence to indicate that the
21 principle effects of ionizing radiation are on early events (Adams 1987; Bouffler 1996;
22 Bouffler 1997a; Ethier 1984; Jaffe 1987; Ullrich 1996; Domann 1994; Gould 1987;
23 Watanabe 1986; Mulcahy 1984). Cellular, cytogenetic and molecular data for AML,
24 intestinal tumors, and mammary tumors also provide evidence for clonal development of
25 radiation-induced pre-neoplasms, implying an initial, single-cell target (Bouffler 1997b;
26 Haines 2000; Ullrich 1996). Recent cytogenetic and molecular studies on the induction of
27 AML and mammary tumors in inbred mouse strains, and of a variety of tumors in
28 transgenic mouse models, have provided more specific information on the potential
29 nature of these early events (Bouffler 1996; Selvanayagam 1995a; Silver 1999; Haines
30 2000; Kemp 1994; Pazzaglia 2002). These studies provide direct support for the view that
31 the critical radiation-associated events in the tumorigenic process are predominantly early
32 events involving DNA losses targeting specific genomic regions harboring critical genes.
33 Since many of the radiation-associated DNA loss events in these tumorigenesis models
34 involve large chromosomal regions within the genome, mechanisms for radiation-induced

1 chromosome aberration induction appear to be of particular relevance to the
2 understanding of radiation effects at low doses. The predominant importance of DNA
3 DSB induction and post-irradiation error-prone NHEJ repair for the induction of
4 aberrations, and the apparently critical role for radiation-induced aberrations in the
5 pathogenesis of cancer in these experimental models, would tend to argue against the
6 proposition of a low dose threshold in the dose-response for the initiation of
7 carcinogenesis.

8 More recently, experimental studies have questioned whether the initiating events
9 produced by radiation are direct chromosomal or mutational effects or whether the
10 mutations and chromosomal rearrangements result indirectly as a consequence of
11 genomic instability induced by the radiation exposure (Little 1997; Little 1990; Morgan
12 1996; Selvanayagam 1995b; Yu 2001).

13 It is well known that the development of tumors is frequently accompanied by the
14 acquisition of genomic instability phenotypes that serve to promote the mutational
15 evolution involved in neoplastic progression. This form of genomic instability is
16 increasingly well understood and many of the responsible tumor gene mutations have
17 been identified (Loeb 2001). This instability, however, differs from radiation-induced
18 genomic instability described during the last decade (Selvanayagam 1995b). Evidence
19 has accumulated that, under certain experimental conditions, the progeny of cells
20 surviving radiation appear to express new chromosomal and gene mutations over many
21 post-irradiation cell generations. The details of radiation-induced genomic instability have
22 been discussed in detail earlier in this report. What may be unique about radiation-
23 induced instability with respect to its potential role in tumorigenesis is that, because of the
24 high frequencies of instability observed following radiation exposure (10-50% of
25 irradiated cells), such instability would not appear to be a result of radiation-induced
26 mutations in a specific gene or family of genes (Kadim 1991; Selvanayagam 1995b;
27 Wright 1995). On the basis of data discussed earlier on radiation-induced genomic
28 instability, and the previously reported high frequency of neoplastic cell transformation
29 (Kennedy 1980; Selvanayagam 1995b), it has been suggested that such events can serve
30 to de-stabilize the genomes of a substantial fraction of the progeny of irradiated cells, and
31 that it is the elevated post-irradiation mutation rates in cell progeny rather than gene-
32 specific initial mutations that act to drive radiation tumorigenesis (Selvanayagam 1995b).
33 The question then arises as to the impact of this type of mechanism on assumptions with
34 respect to low dose risks.

1 Instability associated with telomere dysfunction appears to be of particular
2 relevance to tumorigenesis (Mills 2003; Lo 2002a; Lo 2002b; Desmaze 1999b; Ducray
3 1999; Bouffler 2001; Morgan 1996). Such dysfunction can be manifest in several forms.
4 Telomeric repeat sequences (TTAGGG)_n cap the ends of mammalian chromosomes and
5 serve to protect against replicative erosion and chromosomal fusion; in normal human
6 cells in culture, telomere shortening and instability is a natural feature of replicative cell
7 senescence. Telomeric repeats are also found in subtelomeric and interstitial
8 chromosomal locations and there is some evidence that these loci may act as sites at
9 which radiation-induced and other forms of genomic damage are preferentially resolved.
10 There is also good evidence that telomeric instability is a recurrent feature of tumorigenic
11 development. Of particular relevance to the question of unstable translocation junctions
12 are the so-called segmental jumping translocations which have been well-characterized in
13 spontaneously arising human leukemias. With respect to radiation-induced leukemia,
14 detailed cytogenetic analyses suggest an excess of complex aberrations and segmental
15 jumping translocations in leukemias arising at old ages in high-dose A-bomb survivors
16 (Nakanishi 1999). Telomeric instability at radiation-associated deletion/translocation
17 breakpoints in mouse myeloid leukemia has also been reported but it is not a general
18 characteristic of such tumor-associated events. Interestingly, excess spontaneous
19 telomeric instability is often found to be associated with deficiencies in DNA repair or
20 damage response (Mills 2003).

21 Evidence for the involvement of telomeric sequences in the pathogenesis of at
22 least some forms of radiation-induced instability comes from several laboratories. Early
23 studies on the post-irradiation development of chromosomal instability in *in vitro*
24 passaged human diploid fibroblasts were among the first to suggest a link between
25 telomeres and instability. Initial studies using this in-vitro model were suggestive of
26 instability effects in a high proportion of irradiated cells (Sabatier 1989; Sabatier 1992).
27 Subsequent studies by the same research group have served to address issues related both
28 to the pathogenesis of instability as well as its frequency (Desmaze 1999a; Ducray 1999;
29 Lo 2002a; Lo2002b). Detailed cytogenetic analyses suggested that passage-dependent
30 instability in cultured human fibroblasts primarily represented telomeric events
31 expressing in cell clones naturally selected by growth rate during passage. Overall the
32 data obtained may be interpreted as evidence that initial radiation exposure brings
33 forward in time the natural process of clonal telomeric instability associated with cell
34 senescence and telomere shortening. Equally important is the suggestion that selection

1 processes lead to an overestimate with respect to the frequency of induction of instability
2 by radiation. Whether selection processes impact estimates of the frequency of instability
3 in other systems remains to be addressed

4 A different form of post-irradiation telomere-associated instability is expressed in
5 a hamster-human hybrid cell system where in some clones chromosomal instability is
6 persistently expressed at translocations that have telomeric sequences at their junction
7 (Morgan 1996). Similar unstable structures have been observed in non-irradiated hamster
8 cells undergoing gene amplification. Such data suggest that radiation induces genomic
9 structures that enhance the natural expression of instability. A number of other reports
10 have also suggested that radiation-associated chromosomal exchange can lead to the
11 formation of unstable junctions that undergo secondary change, leading to the formation
12 of complex chromosomal aberrations (Desmaze 1999b; Desmaze 2003; Lo 2002a; Lo
13 2002b; Morgan 1996).

14 The mechanistic role of instability in radiation tumorigenesis is not clear and the
15 two model systems used to study this question have yielded differing results. Radiation-
16 induced genomic instability in hematopoietic cells was first demonstrated in studies
17 showing a persistent excess of chromatid type aberrations in the progeny of mouse bone
18 marrow cells irradiated *in vitro* with alpha particles and subsequently grown in culture
19 (Kadim 1991). Alpha particles are considered to be substantially more effective than
20 low-LET radiation in inducing this form of genomic instability, which has also been
21 reported in the progeny of cells which had not sustained an alpha track traversal; i.e.
22 induced instability may occur as a bystander effect (Lorimore 1998). *In vivo* post-
23 transplantation growth of *in vitro* irradiated bone marrow cells was also reported to result
24 in excess chromatid aberrations. On the basis of these observations it was proposed that
25 such instability had a major role in radiation-induced murine acute myelogenous
26 leukemia (AML). More recent data have not supported this hypothesis, and in fact
27 suggest that radiation-induced instability is not involved in the initiating events in murine
28 AML (Bouffler 2001). Of particular importance in this regard were studies demonstrating
29 that susceptibility to radiation-induced instability in hematopoietic cells, and
30 susceptibility to radiation-induced AML, are not genetically linked phenotypes (Boulton
31 2001).

32 In contrast to these studies are data on instability and radiation-induced mammary
33 cancer. Differences in radiosensitivity and susceptibility to induced tumorigenesis among
34 inbred mouse strains are well recognized and there is good evidence that the BALB/c

1 mouse is unusually sensitive to the induction of tissue injury and mammary tumors, while
2 the C57BL/6 mouse falls into the radio-resistant category (Hanson 1987). Initial
3 cytogenetic studies showed that mammary epithelial cells cultured from irradiated
4 BALB/c mice persistently expressed substantially more chromatid aberrations during
5 passage than those derived from irradiated C57BL/6 animals (Ponnaiya 1997). In follow-
6 up investigations the chromatid instability phenotype of BALB/c was shown to be
7 associated with a partial deficiency in the NHEJ repair protein DNA PKcs together with
8 compromised post-irradiation DNA DSB repair (8 2000; Yu 2001). This study, which
9 included an intercomparison of inbred mouse strains, showed the deficiency of DNA
10 PKcs and DNA DSB repair to be restricted to BALB/c, suggesting genetic associations
11 with persistent genomic instability and with mammary tumor susceptibility. Molecular
12 genetic analyses showed that BALB/c mice carry a rare variant form of the gene (*Prkdc*)
13 encoding DNA PKcs. Subsequent analysis of recombinant mice provided strong evidence
14 that variant *Prkdc* directly determined DNA PKcs deficiency and post-irradiation
15 chromatid instability in mammary epithelial cells (Yu 2001). On the basis of these data it
16 was proposed that induced genomic instability and mammary tumor susceptibility were
17 genetically co-determined. Importantly these investigations provide genetic evidence that
18 a deficiency in the repair of DNA DSB is likely to determine persistent instability.
19 Interestingly, recent observations have suggested a link between DNA PKcs function,
20 telomeric integrity, and genomic instability. The question as to whether such instability is
21 a primary causal element in mammary tumorigenesis remains to be resolved (Bailey
22 1999; Bailey 2001).

23 While the role of radiation-induced genomic instability in radiation-induced
24 cancer is still a matter of investigation, there are several observations that provide a
25 framework for its potential role in cancer development following radiation exposure. In
26 the case of radiation-associated, persistent telomeric rearrangement and unstable
27 chromosome translocation junctions, a strong case may be made that a certain fraction of
28 misrepaired genomic damage after radiation may be prone to ongoing secondary change
29 in clonal progeny. Since there is evidence that such secondary genomic rearrangement
30 can be a normal component of tumor development, it is reasonable to assume that
31 instability of this type would be involved in the pathogenesis of some radiation-associated
32 tumors. It is unclear whether it plays a major role and, if so, for which tumor types. The
33 genetic evidence from mouse mammary studies, which implies that post-irradiation
34 instability can associate with mammary tumor development, supports a role for genomic

1 instability in this system. Thus in certain genetic settings, such as individuals harboring
2 specific types of DNA repair deficiencies, a role for post-irradiation instability in
3 tumorigenesis appears reasonable.

4 Interestingly, recent data in the SCID and in the BALB/c mouse strains, both of
5 which have defects in DNA-PKcs, suggest that telomeric instability may be the
6 underlying mechanism for the induction of instability, and that the resulting cytogenetic
7 instability plays an important role in early carcinogenic events in the mouse mammary
8 carcinogenesis model discussed above. In particular, it appears that dysfunctional
9 telomeres may tend to interact with sites of radiation induced DSBs, increasing the
10 probability of misrepair (Bailey 1999; Bailey 2001; Mills 2003). It would be predicted
11 that mechanisms involving DNA DSB and telomeric sequence interactions would be
12 particularly important at low doses where DNA DSBs are in relatively low abundance.
13 This appears to be consistent with observations that instability is induced in a dose-
14 dependent manner at radiation doses below 0.5 Gy, whereas no dose dependence is
15 observed at higher doses, at which the response appears to plateau. Importantly, the
16 emerging evidence suggests a role for radiation-induced DSBs in the induction of
17 instability and provides a mechanistic link between DSBs, chromosome aberrations, and
18 cancer not unlike that for more directly induced effects. This linkage would also suggest
19 that predictions of effects at low doses will be unaffected by the underlying mechanism
20 whether that mechanism involves direct effects of radiation or is mediated by radiation-
21 induced instability.

22 Observations of microsatellite instability in acute AML among A-bomb survivors
23 (Nakanishi 2001) appear to provide only weak evidence of involvement of this
24 phenomenon in radiation leukemogenesis, with significantly more instability among
25 exposed cf. non-exposed cases but with little evidence of a dose response among the
26 exposed, or of greater involvement in cases in which radiation exposure was more likely
27 to have played a causal role (Little 2002; Cox 2002; Plumb 2003; Little et al, 2003). The
28 question remains open, however, and studies with greater statistical power may some day
29 resolve the issue.

30 Microsatellite instability, observed in radiation-related pediatric thyroid cancers
31 associated with human population exposure to radioactive fallout from the 1986
32 Chernobyl accident, was significantly greater for tumors diagnosed within 6-8 years after
33 the accident compared to those with later onsets (9-11 years); however, without

1 individual radiation dose estimates it was not possible to evaluate the effects of dose on
2 instability (Nikiforov 1998, Richter 1999, Lohrer 2001).

3 4 **5.2 Tissue Modifying factors**

5
6 It is well known that the probability that individual initiated cells will progress to
7 become tumors can be modulated by interactions with surrounding cell and tissue
8 components as well as systemic host factors (Bissell 2001). Studies have also provided
9 evidence that radiation can influence these cell-cell, cell-tissue, and host factor
10 interactions (Barcellos-Hoff 1998; Barcellos-Hoff 2001; Barcellos-Hoff 2001b; Bissell
11 2001; Park 2003). There has been renewed interest in these effects as a result of recent
12 studies that have begun to identify potential underlying mechanisms involved in
13 modulation of tumorigenic progression and expression (Barcellos-Hoff 1996; Barcellos-
14 Hoff 2000; Barcellos-Hoff 2001b; Bissell 2001). Research in this area will be extremely
15 important in understanding the overall processes involved in neoplastic development but
16 a clear understanding of their potential impact on radiation-induced cancer remains to be
17 determined.

18 Two key points tend to support the view that factors involved in modulation of
19 tumor progression and expression are not likely to play a major role in determining low
20 dose risks. It has been demonstrated in a number of instances that an important early and
21 ongoing events in the process of neoplastic development is the acquisition of genomic
22 instability (Selvanayagam 1995a). This instability increases the rate of mutational and
23 chromosomal changes in the cells and increases the probability for mutations that will
24 allow initiated cells to escape from the inhibitory effects of cell, tissue, and host
25 modifying factors. Further, it is also known that with age, there are changes in the tissue
26 microenvironment which also tend to reduce inhibition by normal cells and tissues of the
27 ability of initiated cells to express their neoplastic potential. Over time and with
28 increasing age, therefore, it is highly likely that mutations in initiated cells and alterations
29 in tissue microenvironment will result in the emergence of a cell or population of cells
30 capable of escaping or overcoming these cell, tissue and host modulating factors.
31 Because of this, it seems prudent to focus on early initiating cell and molecular events as
32 the major determinant of risks at low doses.

33 Studies on in vivo tumor induction in mice and rats also suggest that early cell and
34 molecular events represent the principle determinant of radiation-related cancer risk in
35 tissues. In this regard, fractionation studies are particularly relevant. Comparisons of the

1 carcinogenic effects of fractionated exposures to effects of acute radiation exposures of
2 rat skin (Burns 1977; Burns 1975; Vanderlaan 1975) and mouse lung (Ullrich 1980;
3 Ullrich 1984; Ullrich 1987) have clearly demonstrated that the greatest reduction in the
4 carcinogenic effect is for fractions separated by times of 24 hours or less. Such time
5 periods are compatible with repair of initial damage. Longer times of up to 30 days
6 between fractions, which would allow for tissue effects to impact cancer risk, have not
7 been found to result in further reduction in risk.

8 9 **5.2.1 Target cells**

10 In hierarchical-type tissues, where less-differentiated precursor cells produce well
11 differentiated and mature functional cells, cancers are generally considered to originate
12 from tissue stem cells, which possess unlimited division capacity. These tissue stem cells
13 are transformed by carcinogenic agents, altering their differentiation patterns so that cell
14 renewal predominates over differentiation, leading to growth of the abnormal cell
15 population. Stem cells have been well characterized in haemopoietic, epithelial and
16 spermatogenic tissues (Potten, 1983, 1997). They have renewal and location
17 characteristics that are specific to a particular tissue. They renew themselves more slowly
18 than their dividing and differentiating daughter cells, and hence, in protracted irradiation
19 scenarios, receive more ionisations per cell cycle. Stem cells are often located at the
20 static end of a polarized system of cell production, for example near the bottom of
21 intestinal crypts, in the basal layer of epithelia, and more centrally in red bone marrow.
22 These locations can provide some protection against exposure from short-range
23 radionuclides deposited on (for example) epithelial surfaces or lumenally.

24 In the case of the colon, it has been suggested that tumours may originate in cells
25 on the intercryptal plate rather than, or in addition to, stem cells at the base of the crypt
26 (Shih et al., 2001). This study indicated that most early neoplastic lesions of the colon
27 contain dysplastic cells only at the orifices of crypts and on the luminal surface between
28 crypts. Analysis showed loss of the *APC* gene and high expression of β -catenin in such
29 dysplastic cells but not in cells with normal appearance within the crypts. Mutations in
30 the *APC* gene are the earliest genetic alterations in the genesis of colorectal tumours and
31 appear to be required to initiate clonal evolution, involving over-expression of β -catenin
32 (Fodde et al., 2001). This suggestion of target cells on the luminal surface is contentious
33 (Wright and Poulson, 2002; Preston et al 2003). In normal tissue, differentiated

1 epithelial cells on the intercryptal surface would have a very limited life-span of a few
2 days, and would be destined to be lost into the intestinal lumen in the normal process of
3 cell renewal. To develop into a tumour, these dysplastic cells would need to escape this
4 process completely to allow time for progression to malignancy, involving a number of
5 mutational events (Vogelstein et al., 1988; Goyette et al., 1992). Although this scenario
6 seems highly unlikely, the possibility cannot be excluded that daughter cells of the stem
7 cells, situated at higher cell positions in the crypt, are also target cells, perhaps to a lesser
8 degree. *For the purposes of the ICRP report on the Human Alimentary Tract (reference*
9 *to be added), doses are calculated to the estimated position of the stem cells.* However,
10 in considering uncertainties, the possibility that cells higher in the crypts may also be
11 targets has been addressed, including the extreme case of target cells on the intercryptal
12 luminal surface.

13 There are other protective mechanisms in stem cell systems, such as the selective
14 retention of template DNA strands in stem cells, providing protection of the stem cell
15 genome (Cairns 1975, 2002). An example of this is the stem cells in the crypts of the
16 small intestinal mucosa, which divide about a thousand times during the lifespan of a
17 laboratory mouse. Yet these cells show little evidence of any decline in proliferative
18 potential and rarely produce overt tissue abnormalities, suggesting that their genome is
19 extremely well protected. Protection against DNA replication-induced errors can be
20 achieved by the selective sorting of old (template) and new DNA strands with all template
21 strands retained in the stem cell line. Experiments have shown that the template strands in
22 the stem cells can be labeled during development or during tissue regeneration using
23 tritiated thymidine (3HTdR) (Potten et al 2002). Labeling newly synthesized strands with
24 a different marker (bromodeoxyuridine, BrdUrd) allowed segregation of the two markers
25 to be studied. It was shown that template strand label was retained (3HTdR), whereas
26 label in the newly synthesized strands (BrdUrd) was lost following the second division of
27 the stem cell. Random errors may still occur in the template strands owing to
28 environmental agents.

29 Another protective mechanism is apoptosis. Apoptosis is the non-inflammatory
30 and 'altruistic' cell suicide that involves characteristic molecular and cytological features.
31 It occurs naturally at a low level in many hierarchical tissues in the stem cell zone, and
32 the frequency is enhanced by irradiation. This type of cell death is very radiosensitive.
33 Hypotheses for the low rate of cancer in the small intestine have been proposed, based on

1 apoptosis which deletes mutated stem cells (Potten et al., 1992). These hypotheses
2 suggest that radiation-induced TP53-dependent apoptosis in the stem cell zone in the
3 small intestine prevents the propagation of mutated dividing progenitor cells. This is
4 consistent with the increased frequency of cancer in *Tp53*-null mice compared to wild-
5 type mice. Experiments in mice show that the level of apoptosis saturates after acute
6 doses above 100 mGy, there is no detectable dose-rate effect (Hendry et al 1982), and the
7 incidence of apoptosis is repeatable after each dose in a series of small radiation
8 exposures. This provides a potential mechanism in this tissue for the often purported
9 presence of a threshold dose for carcinogenesis. Higher doses indeed are capable of
10 inducing tumours, as found in rats given irradiation to a temporarily exteriorized loop of
11 small intestine (Osborne et al 1963). In the large intestine there is also natural and
12 radiation induced apoptosis. However, *Tp53* is not expressed in the stem cell zone, and
13 *bcl-2* expression promotes cell survival and allows the development of mutated
14 progenitor cells (Merritt et al., 1995). Hence this potential protective mechanism does not
15 operate in the colon. Also, carcinogenesis in the colon may be exacerbated by the longer
16 presence of fecal contents containing carcinogens.

17 In other organ systems such as lung and thyroid, cell renewal is very slow and a
18 much greater proportion of the total cell population may be target cells. In these cases the
19 above mechanisms are very unlikely to apply, and the long-lived target cells would
20 accumulate multiple mutations in the conventionally-described multistage process of
21 carcinogenesis (Vogelstein et al., 1988; Goyette et al., 1992).

22 An important question in with respect to protective mechanisms in target cells and
23 the removal of damaged cells via apoptosis is the persistence of radiation-initiated cells
24 once the initial damage has been produced. Hoshino and Tanooka examined the
25 persistence of latent carcinogenic damage in irradiated mouse skin (Hoshino and Tanooka
26 1975) and found that radiation initiated cells could persist as latent carcinogenic damage
27 for up to 400 days. Yokoro and his co-workers, in studies examining the interaction of
28 radiation and hormones in breast cancer development, also found that latent radiation-
29 initiated cells persisted for a substantial portion of the rats' lifetimes (Yokoro and others,
30 1977).

31

5.3 Radiation induced Cancer in Animals

On the basis of the discussion of *cellular and molecular mechanisms* above, it can be predicted that the dose response and time-dose relationships for radiation-induced cancer would be similar to those for radiation-induced chromosomal aberrations. Specifically, at low doses a linear dose response would be anticipated. There are, however, relatively few studies on animal carcinogenesis where the data are sufficient to address the issue of dose response relationships or the issue of dose rate effects, protraction, and/or fractionation effects and rigorously test these predictions. Those studies where such analyses are possible are mainly limited to rodent studies, principally studies in mice. A further caveat is the applicability of animal data to human risks. The pathogenesis of certain tumors in experimental animals appears to involve unique mechanisms for induction that do not appear to be compatible with known mechanisms of cancer development in humans. This section will describe the available data and its applicability to understanding of low dose risks and risks following low dose rate or protracted exposures. This is not meant to be a comprehensive review but is limited to those data sets which focus on effects at low doses (< 0.5 Gy) and low dose rate exposures following external irradiation. Data from studies using internal emitters are not included because of the dosimetric issues that complicate interpretation. Likewise, studies with low statistical power in the low dose range have also been excluded.

At first glance an examination of available animal data suggests a high degree of complexity in that a variety of dose responses have been observed ranging from threshold responses to linear or linear quadratic responses. However, a more systematic examination of the data with a view toward the underlying biology involved in the pathogenesis of individual tumor types reveals a clearer picture. In this regard it is useful to first separate the discussion of the data into that for induction of leukemias and solid tumors.

5.3.1 Leukemia

The induction of leukemia and lymphoma has been examined in two murine systems, thymic lymphoma and acute myelogenous leukemia. The dose response for induction of thymic lymphoma is complex and reducing the dose rate results in a large reduction in the effectiveness for radiation-induced thymic lymphoma (Ullrich 1979a). The applicability of these data to human risk estimates is unclear. The development of thymic lymphoma in mice following irradiation is an extremely complex process largely

1 mediated through indirect mechanisms (Kaplan 1964; Kaplan 1967). Importantly in this
2 regard, expression of thymic lymphoma can be substantially reduced or eliminated by
3 protection of a small fraction of bone marrow stem cells from radiation-induced cell
4 killing. The complex nature of the pathogenesis of murine thymic lymphoma involving
5 substantial bone marrow cell killing, and the lack of a comparable counterpart in humans
6 argues against thymic lymphoma as an appropriate model for the understanding of dose
7 response and time-dose relationships in humans.

8 In contrast, data on the biology and pathogenesis of murine acute myelogenous
9 leukemia (AML) suggest strong similarities between mouse and human. Such data
10 support its applicability to radiation-induced leukemogenesis in humans with respect to
11 studies of mechanisms and potential low dose risks (Rithidech 1999; Rithidech 2002;
12 Silver 1999; Tenen 2003). For murine AML the most comprehensive data on dose
13 response and dose rate or fractionation pertain to radiation-induced myeloid leukemia in
14 CBA mice and RFM mice (Mole 1983; Mole 1983; Ullrich 1987; Upton 1970). The
15 CBA mouse has also been used to dissect underlying radiation-induced molecular events
16 described previously (Bouffler 1996a; Bouffler 1996b; Bouffler 1997). Over the 0 to 3
17 Gy dose range (the lowest dose used was 250 mGy), the dose response for both strains
18 could be described by a pure quadratic dose response relationship, although linear-
19 quadratic and simple linear dose responses also provided an adequate fit to the data sets.
20 After fractionation or protraction of the dose there was a reduction in the leukemogenic
21 effects of radiation at doses of 1.5 Gy and higher resulting in a linear dose response over a
22 wide range of doses in both strains. Barendsen has analyzed the RFM data set including
23 acute high dose rate, fractionated and low-dose rate exposures and concluded that a
24 linear-quadratic model derived from the high dose rate data adequately predicted the low
25 dose rate and fractionation effects (Barendsen 1975). Importantly, these data and the
26 analysis by Barendsen are fully compatible with predictions based upon the known role
27 for aberrations/deletions in chromosome 2 in the pathogenesis of murine AML and
28 predictions based upon data for induction of chromosome aberrations by radiation.

29 **5.3.2 Solid Tumors**

30 Data from experimental studies examining dose response relationships following
31 whole body external exposures are also available for a limited number of solid cancers.
32 The tumor types for which sufficient data are available include Harderian gland, pituitary,
33 and ovarian tumors in female RFM mice (Ullrich 1979a; Ullrich 1979b), and lung and
34 breast cancers in female BALB/c mice (Ullrich 1983; Ullrich 1987). Data are also

1 available in female Sprague-Dawley rats for mammary tumors (Burns 1975; Burns 1977;
2 Finkel 1968; Hulse 1969; Shellabarger 1980) and for skin in mice and rats and bone
3 tumors in mice . The data for skin and bone tumors involve localized exposures since the
4 induction of these tumors generally requires radiation doses that are too high to be well
5 tolerated when given as whole body exposures.

6 The observation that high radiation doses are required for induction of skin and
7 bone tumors supports the view that a threshold might exist for induction of these tumors.
8 However, this does not imply that low doses of radiation cannot and do not result in the
9 initiation of skin and bone cells. Studies in mouse skin clearly demonstrate that low
10 doses of radiation can initiate cells that have the potential to progress to become tumor
11 cells (Jaffe 1987). Rather, these data suggest that for these tissues, factors influencing
12 tumor progression play an important role in determining whether or not initiated cells
13 progress and ultimately express their tumorigenic potential. The high doses required
14 suggest an important role for radiation-induced cell killing resulting in disruption of cell-
15 cell and cell-tissue interactions as well as the recruitment of growth factors all of which
16 may participate in the progression of initiated cells in these systems. It is important to
17 note that skin and bone are also not considered highly sensitive to radiation-induced
18 cancer in humans as well. By far the greatest contribution to estimates of radiation risk
19 comes from tissues that are more sensitive to tumor induction and for which risks at low
20 doses are of more concern.

21 The apparent lack of sensitivity of bone and skin at low doses does not mean that
22 risks can be ignored. Exposure to ultraviolet light has been shown to be an effective
23 promoting agent following exposure of the skin to ionizing radiation (Shore 1984). Such
24 exposure allows the expression of initiated cells that would not be expressed otherwise.
25 As a result the relationship between the dose of ionizing radiation and skin tumor
26 development shifts from one with an apparent threshold to a much more linear response.
27 This effect underscores the argument made previously in this section that it is important
28 to focus on early initiating cell and molecular events as the major determinant of risks.
29 An apparent threshold cannot be assumed to indicate that there is no increased risk to an
30 individual who might be exposed to other agents with promoting effects or for whom
31 intrinsic risk factors could exist which could allow expression of initiated cells that
32 would normally not be expected.

33 Data from the studies using RFM and BALB/c mice and Sprague-Dawley rats are
34 most applicable with respect to low dose and low dose rate effects because of the

1 sensitivity of these tissues to radiation-induced cancer and the dose range over which data
2 has been obtained. Again, caution must be exercised in the application of data derived
3 from all tumor types without regard to the underlying biology involved in tumorigenesis.
4 The most dramatic example is that for ovarian cancer in mice. Ovarian cancer in the
5 mouse following whole body irradiation appears to be a result of an indirect mechanism
6 involving oocyte cell killing, and subsequent alterations in the pituitary-ovarian hormonal
7 interactions leading to ovarian tumorigenesis (Foulds 1975). Because of the close
8 association between cell killing and ovarian cancer in mice and because mouse oocytes
9 are uniquely sensitive to the killing effects of radiation (LD50) for oocyte killing is
10 approximately 50 mGy, ovarian tumors at high frequencies are observed following very
11 low doses. Consistent with an indirect mechanism mediated by cell killing, a threshold
12 dose response has been observed for the induction of ovarian tumors. Lowering the dose
13 rate increased the threshold dose from approximately 110 mGy to 700 mGy (Ullrich
14 1979c; Ullrich 1979b). There is no evidence for similar indirect mechanisms for
15 radiation-induced cancer at any site in human studies and, therefore, radiation-induced
16 ovarian tumorigenesis will not be included in further discussions below.

17 Data for the induction of Harderian gland and pituitary tumors in female RFM
18 mice and lung and mammary cancer in female BALB/c mice generally support the linear-
19 quadratic model over a dose range from 0.1 - 2Gy (Ullrich 1979a; Ullrich 1979b; Ullrich
20 1987) while the induction of mammary tumors in Sprague-Dawley rats tend to be more
21 linear over this dose range (Shellabarger 1980). For these tumor types it has also been
22 found that reducing the dose rate or fractionating the dose into small fractions reduces the
23 risk for development of radiation induced cancer in the manner predicted by the linear-
24 quadratic model. At high doses (>1 Gy) the risk of cancer development is reduced
25 primarily as a result of the diminution of the quadratic portion of the dose response
26 resulting in a limiting linear slope over a wide dose range that is equivalent to the linear
27 slope of the high dose rate dose response in the low dose range. At lower total doses
28 radiation effects are time independent and therefore the incidence of tumors increase in a
29 linear fashion with dose.

30 **Overall, relevant animal tumor data tend to support a linear response with**
31 **no threshold at low doses.**

5.4 Life shortening

1
2
3 A large number of studies in mice and dogs have been conducted using life-span
4 shortening as a means to quantify late radiation effects (NCRP 1980; Carnes 1989;
5 Carnes 2002; Carnes 2003; Storer 1979; Storer 1982; Storer 1983; Thompson and Grahn
6 1988, 1989; Thompson et al 1981a, 1981b, 1983, 1985, 1986; Sacher et al 1958, 1976;
7 Leshner et al 1960, 1965; Grahn and Hamilton 1964; Grahn and Sacher 1957, 1958; Grahn
8 et al 1963). While it has been argued that life shortening can serve as an integrated
9 measure of the deleterious effects of radiation, the interpretation of these studies is not
10 straightforward. A large variation in life shortening is observed as a function of strain,
11 species, gender, and physiological status of the animals. This variation is largely a result
12 of differences in the spectra of spontaneous and induced disease, and the age distribution
13 of disease occurrence. For example, a high degree of life shortening is observed in
14 animals susceptible to the induction of radiation-induced cancers that tend to occur early
15 in life, such as thymic lymphoma or myelogenous leukemia. Studies using animals that
16 are not susceptible to such typically early-developing neoplasms but, rather, tend to
17 develop late-occurring solid tumors following radiation exposure have observed
18 considerably less life shortening at the same radiation dose. Regardless of the degree of
19 life shortening observed, however, analyses of experimental studies indicate that at low
20 doses of radiation and for radiation delivered at low dose rates, radiation-induced life
21 shortening is due almost entirely to radiation-induced cancer (NCRP 1980; Carnes 2002;
22 Storer 1979; Storer 1982; Leshner et al 1960). Life shortening attributable to non-
23 neoplastic effects has only been observed at single acute doses in the range of 500 mGy
24 and higher and no such effects have been observed following low dose rate or protracted
25 exposures to low LET radiation (Carnes 2002; Storer 1979; Storer 1982).

26 Experiments designed to address questions of risk following low dose rate or
27 protracted exposures have also been performed. With few exceptions dose response
28 relationships derived from data following single acute radiation doses, fractionated
29 exposures and terminated low dose rate exposures all suggest linear dose responses over a
30 wide range of doses (NCRP 1980; Carnes 2003; Storer 1979; Thomson and Grahn 1988,
31 1989; Thomson et al 1981a, 1981b, 1983, 1985, 1986; Tanaka et al 2003). This is not
32 surprising since the dose response for life shortening represents the integrated dose
33 responses for a variety of tumor types whose individual dose responses may vary widely.
34 The primary effect of fractionating the radiation dose or reducing the dose rate at which

1 the dose is delivered is to reduce the slope of the linear response. Importantly,
2 experiments using multiple low dose-rate, terminated exposures suggest a limiting linear
3 slope in all cases. Once this limiting linear response is reached, no further reduction in
4 effect is seen if dose rate is reduced further. Protracting exposures over the entire life-
5 span can result in a further reduction in life shortening per unit dose. There are two
6 confounding factors in protraction studies that must be considered. First, in such studies
7 the radiation injury induced very late in life often does not have sufficient time to be
8 expressed. Second, it is difficult to determine the dose at which specific effects have been
9 induced because the exposure continues even after the processes involved have been
10 initiated. Both factors tend to result in an overestimation of the dose required to produce
11 a specific degree of observed life shortening (NCRP 1980). This overestimation of the
12 dose reduces the slope of the dose effect relationship beyond the limiting slope obtained
13 following terminated exposures.

14
15

5.5 Summary

16 Studies on the cellular and molecular mechanisms of carcinogenesis over the last
17 several years have provided substantial insight with respect to the complex multi-step
18 nature of the process of neoplastic development and on radiation-induced cancer. These
19 studies provide direct support for the view that the critical radiation-associated events in
20 the tumorigenic process are predominantly early events involving DNA losses targeting
21 specific genomic regions harboring critical genes. Since many of the radiation-associated
22 DNA loss events in these tumorigenesis models involve large chromosomal regions
23 within the genome, mechanisms for radiation-induced chromosome aberrations appear to
24 be of particular significance. The predominant importance of DNA DSB induction and
25 post-irradiation error-prone NHEJ repair for the induction of aberrations, and the
26 apparently critical role for radiation-induced aberrations in the pathogenesis of cancer in
27 these experimental models, would tend to argue against the proposition of a low dose
28 threshold in the dose-response.

29 More recently, experimental studies have questioned whether the initiating events
30 produced by radiation are direct chromosomal or mutational effects or whether the
31 mutations and chromosomal rearrangements result indirectly as a consequence of
32 genomic instability induced by the radiation exposure. However, at this point the
33 mechanistic role of instability in radiation tumorigenesis is not clear. Data thus far
34 suggests that in certain genetic settings, such as individuals harboring specific types of

1 DNA repair deficiencies, a role for post-irradiation instability in tumorigenesis appears
2 reasonable but its general applicability and its impact on low dose risks remains a matter
3 of investigation.

4 Factors that modify the progression and persistence of initiated cells must also be
5 considered when addressing low dose risks. It is well known that the probability that
6 individual initiated cells will progress to become tumors can be modulated by interactions
7 with surrounding cell and tissue components as well as systemic host factors. Data to
8 date, however, suggest that such factors are not likely to play a major role in determining
9 low dose risks. Another important question is the persistence of radiation-initiated cells
10 once the initial damage has been produced. It has been hypothesized, for example, that
11 apoptosis could be a protective mechanism which removes potentially neoplastic cells
12 and could in effect result in a threshold at low radiation doses. Two studies using different
13 experimental systems (skin and mammary gland) have addressed this issue and found that
14 latent radiation initiated cells could persist for a substantial portion of the rats' lifetimes.
15 At present, therefore, it seems prudent to focus on early initiating cell and molecular
16 events as the major determinant of risks at low doses.

17 On the basis of the discussion of cellular and molecular mechanisms in this
18 chapter, it can be predicted that the dose response and time-dose (i.e., fractionation and
19 protraction) relationships for radiation-induced cancer would be similar to those for
20 radiation-induced chromosomal aberrations. Specifically, at low doses a linear dose
21 response would be anticipated. There are, however, relatively few studies on animal
22 carcinogenesis where the data are sufficient to address the issue of dose response
23 relationships or the issue of dose rate effects, protraction, and/or fractionation effects and
24 rigorously test these predictions. Those studies where such analyses are possible are
25 mainly limited to rodent studies, principally studies in mice. Overall, these animal tumor
26 data tend to support a linear response at low doses and dose rates with no threshold.

27 A large number of studies in mice and dogs have been conducted using life-span
28 shortening as a means to quantify late radiation effects and it has been argued that life
29 shortening can serve as an integrated measure of the deleterious effects of radiation.
30 Support for this argument comes from the observation that, regardless of the degree of
31 life shortening observed, radiation-induced life shortening is due almost entirely to
32 radiation-induced cancer. Life shortening experiments have examined risks following low
33 dose, low dose rate or protracted exposures. The primary effect of fractionating the
34 radiation dose or reducing the dose rate at which the dose is delivered is to reduce the

1 slope of the linear response. Importantly, experiments using multiple low dose rate
2 terminated exposures suggest a limiting linear slope in all cases adding further support for
3 the view that effects at low doses are consistent with a linear no threshold model.

5 **5.6 Conclusions: Implications for radiation-related cancer at low doses**

6
7 Models of radiation action as well as a wide range of molecular, cellular and
8 animal data have been used to argue that data on radiation-induced cancer in human
9 populations derived from studies following acute radiation exposures tend to overestimate
10 radiation risks at low doses and dose rates (NCRP 1980; ICRP 1991). In this regard, a
11 number of advisory groups have used a similar approach to quantify the degree to which
12 extrapolation of acute high dose data might tend to overestimate risks at low doses and
13 low dose rates. Essentially, the effectiveness per unit dose for acute exposures has been
14 determined using a linear interpolation of data between the 2-3 Gy dose range and control
15 data at 0 Gy. Effects per unit dose following low dose rate exposures were derived by
16 calculating the slope of the entire dose response (not just in the 2-3 Gy dose range). By
17 dividing the tumorigenic effectiveness per unit dose of acute exposures using the high
18 dose data and low dose rate exposures, an effectiveness ratio was obtained. This ratio has
19 been termed the Dose and Dose Rate Effectiveness Factor. The rationale for using only
20 the high dose data and not data at lower doses was based on the assumption that this
21 would simulate analyses of risks from epidemiological studies where most of the
22 available data were for single acute exposures at relatively high doses. Since the actual
23 dose response for most radiation-induced tumors following single acute exposures has
24 generally been found to be linear quadratic (see discussion above), this procedure would
25 tend to overestimate effects for low single acute radiation doses (in the dose range where
26 the response is predominantly linear) and for low dose rate exposures over a wide range
27 of total doses.

28 In spite of its apparent simplicity, the derivation and application of dose and dose
29 rate effectiveness factors (DDREF) must be performed with caution. Tumors for which
30 there is evidence (from knowledge of their mechanisms), that they are unlikely to be
31 applicable to radiation carcinogenesis in human populations, should not be considered.
32 This leaves a limited data set upon which to base DDREF calculations. These data sets
33 include myeloid leukemia, and a few solid tumors including Harderian gland (for which
34 there is no comparable tissue in humans), lung adenocarcinomas, and mammary tumors.

1 All the data sets for myeloid leukemia support a reduced carcinogenic effect when
2 comparing high and low dose rate exposures over the 0 to 3 Gy dose range. Calculation of
3 DDREF values using the procedures described above yield estimates on the order of 2 to
4 6 with most values in the range of 4-5. For lung adenocarcinomas and for Harderian
5 gland tumors DDREF values of approximately three have been calculated over the 0 to 2
6 Gy dose range. For mammary tumors all of the data suggest a DDREF value of less than
7 2 and more nearly close to a value of 1 when effects of high dose rate and low dose rate
8 exposures are compared in this 0-2 Gy dose range. Thus, it appears that myeloid
9 leukemia is probably more sensitive to dose rate effects than are solid tumors.

10 It should be emphasized that these values are based upon extrapolation of data from
11 acute doses of 2-3 Gy, and may represent maximum DDREF values (NCRP 1980). Total
12 dose-dependent dose-rate effects have also been reported and quantified for cytogenetic
13 endpoints by Sorensen and co workers (Sorensen et al 2000). The impact of dose range
14 must be considered when applying DDREF factors to human risk estimates for which
15 there are now reliable data at and below 1 Gy.

16 It has been argued that life shortening data may be a more appropriate measure of
17 overall risk, and therefore, the use of these data is a better approach to the derivation of a
18 single DDREF value. The complications of life shortening data have been described in
19 an earlier section including changes in disease spectrum as a function of dose and dose
20 rate, and complications associated with terminated versus life-time exposures. These
21 complications notwithstanding, DDREF values determined from terminated radiation
22 experiments indicate maximum DDREF values following extrapolation of acute effects in
23 the 2 Gy dose range on the order of 2. Protraction of the radiation exposure over a
24 significant portion of an animal's life-time tends to reduce the effectiveness of the
25 exposure more than that observed following a simple reduction of dose rate to specific
26 total doses. However, as discussed earlier this experimental approach makes the
27 determination of true effects per unit dose difficult if not impossible. Because of this, the
28 application of these large (i.e., >2) protraction factors to human risks is problematic.

29
30

5.7 References

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6. QUANTITATIVE UNCERTAINTY ANALYSIS

6.1 Overview

Chapter 2 described the epidemiological basis for estimation of radiation-related cancer risk in exposed populations, including various uncertain factors that must be considered when applying epidemiological risk estimates from one population to another, especially when the base data are as yet incomplete and must be projected forward to end of lifetime of the study population. The discussion was focused on uncertain biases introduced by random dose-reconstruction error in the first population, population differences in baseline cancer rates, and extrapolation of estimates, derived largely from moderate-to-high dose data, to situations of low-dose and very-low-dose exposure. The topic of the present chapter is quantitative uncertainty analysis of estimated cancer risk associated with low-dose, low-LET radiation exposure, illustrated in terms of the application of atomic bomb survivor risk coefficients to the population of the United States.

Quantitative uncertainty analysis (QUA) was developed in a decision-theoretic framework and has been extensively applied to nuclear reactor safety (U.S. Nuclear Regulatory Commission 1974, 1990) and ecological risk assessment (IAEA 1989; Warren-Hicks and Moore 1998; Gilbert 1994). It involves the application of Bayesian probability methods to estimates and decision rules based on uncertain statistical and subjective information. As stated by Warren-Hicks and Moore (1998), benefits of quantitative uncertainty analysis include improved transparency and credibility, avoidance of worst-case assumptions, focus on critical areas of uncertainty that might benefit from further data collection, and improved decision support. Limitations of the method include the practical inability to consider all possible sources of uncertainty, the possibility that the method may be used incorrectly, and lack of universal awareness and acceptance of the methodology.

The approach (i.e., QUA) is used here, not to reach a particular decision, but to illustrate the implications for radiation protection of the various types of (mostly uncertain) information that contribute to our estimates of radiation-related risk. The emphasis on uncertainty is appropriate because the need for radiation protection is driven by the likelihood and magnitude of exposure-related risks, because estimates based on statistical data and realistic assumptions are uncertain, and because radiation protection is

1 a political process which must take account of the diverse interests and viewpoints of
2 individuals and population subgroups affected by implementation of radiation protection
3 policies. The development of such policies, to be successful, requires accommodation and
4 consensus. It must be seen to be done fairly and openly, on the basis of facts and
5 assumptions accessible to and challengeable by all of those affected by implementation.
6 An important aspect of the information relevant to the political process of radiation
7 protection is the uncertainty of estimates of radiation-related risk derived from a
8 combination of statistical and largely subjective information sources.

9 Different people have different points of view about risk. For example, a risk-
10 averse person may tend to focus on how high the risk from exposure might reasonably be
11 (e.g., on its upper 90% uncertainty limit), while a person who is primarily averse to the
12 costs of exposure reduction may tend to demand proof that the risk is high enough to
13 worry about, e.g., may focus on its lower uncertainty limits. A complete uncertainty
14 distribution for estimated risk summarizes all the uncertainty information inherent in the
15 statistical data used and in the consensus estimates of crucial assumptions needed to apply
16 the statistical data to the matter at hand. That summary is highly relevant to both of these
17 points of view and to others as well.

18 Radiation-related cancer risk is among the subjects most suitable for QUA. It is
19 highly quantified, and a number of major sources of uncertainty have been explored (NIH
20 1985; CIRRPC 1988; Sinclair 1994; NCRP 1996, 1997; EPA 1999; NCI/CDC 2003).
21 Knowledge of uncertainty is highly relevant to radiation protection philosophy and
22 practice, and it can be at least as important as knowing the value of a single-valued “best
23 estimate”. For example, a point estimate of one lifetime excess cancer death per thousand,
24 with 90% probability (uncertainty) limits 0.5-2.0 per thousand, has different implications
25 for, say, a risk-benefit analysis than the same point estimate with probability limits 0.1-10
26 per thousand. In the second case, assuming a lognormal uncertainty distribution, the
27 likelihood that the risk per thousand is between 0.5 and 2.0 is only 38% and the likelihood
28 that it is greater than 2.0 is 31%.

29 Statistical analyses of epidemiological or experimental observations on radiation
30 carcinogenesis are usually concerned with quantifying risk in the context of a particular
31 study. Applications of the original risk estimates in other contexts, without adjustment,
32 may be misleading for a number of reasons discussed earlier in this chapter. Adjustment
33 requires other steps and assumptions, about which the original study may not be

1 informative. The incorporation from other sources of additional information, which may
2 be uncertain, may modify the resultant risk estimate and its uncertainty.

3 Uncertainty analysis is concerned with such changes and their implications for the
4 ultimate application of (in the present case) risk estimates. The approach has been
5 extensively applied in assessments of environmental contamination (NCRP 1996). The
6 1985 NIH radioepidemiological tables report (NIH 1985) was possibly the first formal
7 application to radiation-related cancer risk. The approach was subsequently taken a step
8 farther, at the request of the United States Department of Veterans Affairs (VA), by the
9 Committee on Interagency Radiation Research and Policy Coordination (CIRRPC 1988).
10 The following discussion is based primarily on the following sources: NCRP
11 Commentary 14 (1996) discusses uncertainty analysis applications to assessment of dose
12 and risk related to environmental contamination; NCRP Report 126 (1997) was derived in
13 part from Sinclair (1994) and is specifically concerned with applications of radiation-
14 related mortality risk estimates to low-LET radiation protection; an Environmental
15 Protection Agency report (1999) deals with the same subject; and a recent revision of the
16 1985 NIH radio-epidemiological tables report (NCI/CDC 2003) is concerned with
17 applications to adjudication of compensation claims for radiation-related cancer
18 morbidity.

19 When we estimate the radiation-related cancer risk associated with a particular
20 low-dose exposure history, what is it we are estimating? Some possibilities:

21 **a) An increase in lifetime cancer rate**, e.g., from r to $r' = r \times (1 + x)$, for a
22 particular population specified by age, sex, lifestyle, etc. Note that this increase
23 theoretically can be verified by observation of cancer rates among exposed and non-
24 exposed members of the population. Estimation requires information on:

25 *i)* Dose-related risk in some population (or group of populations), and the variation of that
26 risk by sex, age, etc. Generally, this information will pertain most directly to doses and
27 dose rates higher than those of immediate interest. For radiation-related risk, there is a
28 substantial body of epidemiological information, the most comprehensive of which is
29 based on follow-up of the survivors of the atomic bombings of Hiroshima and Nagasaki,
30 Japan.

31 *ii)* How to transfer risk estimates for the informative population to the population of
32 interest, which may differ from the first population in specified ways such as baseline
33 cancer rate, smoking prevalence, patterns of reproductive history, other possible dose-
34 response modifiers. Also, random and biased errors in dose reconstruction for the first

1 population, which should not affect risk estimates for members of the first population,
2 may bias the application of dose-specific risk estimates to the second population. A
3 similar problem exists for biased ascertainment of cancer cases, e.g., because of
4 inaccuracies of death certificates.
5 *iii*) How to extrapolate risk from high to low doses and from high to low dose rates,
6 including dose and dose-rate effectiveness factor (DDREF) and departures from the LNT
7 hypothesis such as hormesis and low-dose threshold.

8 ***b*) The likelihood that a particular individual will develop cancer as a result
9 of his or her exposure.** Note that this likelihood is not verifiable at the individual level;
10 the individual either will or will not develop cancer, and the estimate of the individual's
11 probability, or excess probability, of developing cancer is verifiable only if we assume
12 that information on a population also pertains to the individual.

13 Thus, *b*) reduces to *a*), and is addressed as follows:

14 *i*) Identify the individual as a member of some population with the exposure history and
15 other characteristics of the individual insofar as the relevance of these characteristics to
16 risk is known or estimated.

17 *ii*) Estimate the exposure-related increase in cancer rate for that population.

18 *iii*) Treat the individual as a randomly sampled person from the population, i.e., a possible
19 cancer event is treated as a Bernoulli random variable with probability $p = r'$ as given in
20 *a*) above. Note that r' is itself an uncertain quantity.

21 The several kinds of required information discussed under *a*) are qualitatively
22 different. Many of them are subjective in nature, requiring expert judgment.

23

24 **6.2 Sources of uncertainty**

25

26 **6.2.1 Statistical estimate of excess risk per Gy.**

27 The epidemiological information from a radiation-exposed population is
28 summarized by a statistical estimate, of excess absolute or excess relative risk (EAR or
29 ERR, respectively), the uncertainty of which can be expressed by confidence limits or,
30 more comprehensively, by a probability distribution derived from the statistical likelihood
31 contour of the estimate. This probability distribution defines likelihood-based statistical
32 confidence limits at all confidence levels, and may depend upon sex, exposure age,
33 attained age, and other identifiable risk modifiers. Figure 6.1 represents an example of a
34 likelihood-based statistical uncertainty distribution for excess relative risk of cancer at

1 ages 50 or older following a 1 Gy, whole-body, acute exposure at age 40. The estimate is
2 based on a linear-model dose-response analysis of LSS tumor registry cancer incidence
3 data (Thompson 1994) for males, reanalyzed in the context of adjudication of
4 compensation claims for possibly radiation-related cancer (NCI/CDC 2003). In that
5 analysis, it was found that most variation of ERR by exposure age was confined to ages
6 under 30, and that most variation by attained age occurred at ages under 50. A model was
7 used based on log-linear splines in exposure age and attained age such that there was no
8 variation in ERR per Gy by exposure age after 30 and by attained age after 50. The
9 resultant statistical uncertainty distribution for ERR per Gy at older exposure ages and
10 attained ages is approximately lognormal with 5th and 95th percentiles (90% confidence
11 limits) 0.18 and 0.43.

12 This statistical uncertainty distribution is the basis for the numerical
13 demonstration presented below. However, summary results are also given, later in this
14 chapter, for calculations based on the fitted estimate for a female population, with a
15 lognormal statistical uncertainty distribution and 90% confidence limits 0.45 and 0.72,
16 and for a population evenly divided by sex, for which the confidence limits are 0.33 and
17 0.53.

18 Estimates of excess absolute risk (EAR) for age-specific risk, or for lifetime risk
19 starting from age 50, can be obtained by scaling the ERR distributions by the appropriate
20 age-specific or lifetime baseline cancer rates. However, since in most applications the
21 population of interest is not the LSS population and the exposure of interest is not to an
22 acute dose of 1 Gy, it is computationally convenient to develop the ERR estimate for the
23 population and exposure of interest and then convert to EAR.

24

25 **6.2.2 Diagnostic misclassification.**

26 Based on autopsy-based analyses by Sposto et al (1992) of misclassification of
27 cancer as noncancer on death certificates, NCRP Report 126 (1997) introduced an
28 uncertain correction factor for combined-site cancer mortality risk estimates, subjectively
29 distributed as normal with 5th and 95th percentiles 1.02 and 1.18, respectively. No
30 correction factor was deemed necessary, however, for cancer morbidity as determined by
31 the RERF Tumor Registry, and none is applied in the present exercise. (Here, (or 90%
32 “probability limits”, here used as a general term to include statistical confidence limits
33 and uncertainty limits for distributions that have a subjective component - see footnote 1
34 to Chapter 2)

6.2.3 Dose-reconstruction errors.

Application of epidemiological information from one radiation-exposed population to a second population is problematic because errors in dose reconstruction for the first population are unlikely to be repeated in the second; therefore, dose-specific risk estimates should be corrected before being applied to the second population. Also, lifestyle, environmental, and other factors may differentially modify radiation dose response in the two populations.

NCRP Report 126 treated bias correction for dose-reconstruction error in the A-bomb survivor population, involving 5 different factors: random errors in individual dose estimates (following Pierce et al, 1991), uncertainty about the magnitude of the neutron component of dose in Hiroshima, uncertainty about the relative biological effectiveness weight, relative to gamma dose, applied to the neutron component of individual dose, uncertain neutron dose, and uncertain gamma dose. A full rationale is given in the NCRP report (NCRP 1977) to which the reader is referred for details. With the implementation of a new A-bomb survivor dose reconstruction system, designated DS02 (Preston et al. 2004), the details will change. For present purposes, it is enough to note that dose reconstruction is a source of bias and uncertain error, which can contribute to the uncertainties of risk estimates and should be taken into account. For illustration, we use the subjective uncertainty distribution of the combined correction factor described in Figure 3.6 of NCRP Report 126 and redrawn for Figure 6.2 of the present report, which was calculated as approximately normal with mean 0.84 and 90% uncertainty limits 0.69 to 1.0. The resulting corrected uncertainty distribution for ERR at 1 Gy is approximately lognormal with mean 0.26 and 90% limits 0.15-0.46 (Figure 6.3).

6.2.4 Transfer between populations

Also uncertain is the relationship between radiation-related excess risk and baseline cancer rates in the two populations. This is an important consideration if population baseline rates differ substantially. For example, current age-specific incidence rates for female breast cancer are substantially higher in the United States than in Japan, according to tumor registry data from Hiroshima and the U.S. SEER registry (Parkin, 1997) (Figure 6.4). In the figure, breast cancer risk among female A-bomb survivors exposed to a breast tissue dose of 1 Gy at age 15 is represented as a constant multiple of age-specific baseline risk beginning at age 25. The two dashed curves tracking the US

1 age-specific baseline rates represent two of many different ways of transferring the A-
2 bomb survivor estimate to a US population. The lower of the two dashed curves was
3 calculated as the sum of the US baseline rate plus the radiation-related excess (absolute)
4 rate in the A-bomb survivors (additive transfer). The higher curve was calculated as the
5 product of the US baseline rate times the estimated radiation-related relative risk among
6 the A-bomb survivors (multiplicative transfer). If the baseline rate curves were the same,
7 the additive and multiplicative transfer methods would give the same solution. Because
8 the baseline rates are so different, the lifetable-averaged (over age) estimates of excess
9 risk differ by three-fold.

10 In the case of breast cancer, there is epidemiological evidence that the additive
11 transfer model is more realistic than the multiplicative model (Preston, 2002; Little and
12 Boice, 1999; Land, 1980b), but there is not enough evidence to rule out alternatives. For
13 stomach cancer there are some data favoring multiplicative transfer (Carr, 2002; Boice,
14 1988; Inskip, 1990). For most other site-specific cancers, there is little or no relevant
15 information on transfer between populations. NCRP Report 126 considered only total
16 cancer mortality, which is about 40% and 80% higher in the US than in Japan for males
17 and females, respectively (Pisani, 1999). In Report 126, subjective uncertainty about
18 population transfer was expressed as an uncertain multiplicative correction factor,
19 distributed as lognormal with 5th and 95th percentiles at 0.70 and 1.65, respectively, to be
20 applied to the multiplicative transfer model estimate (NCRP, 1997).

21 For site-specific cancers a more detailed approach is needed because standardized
22 rates may differ between the two countries by as much as 10- to 15-fold in either direction
23 (e.g., for liver, stomach, prostate), although for most sites rates are more comparable. The
24 approach used for the updated NIH radioepidemiological tables report (NCI/CDC, 2003)
25 for most cancer sites was to weight equally all possible linear combinations of the
26 multiplicative (M) and additive (A) model estimates, $p \times M + (1-p) \times A$, by assuming p to
27 be a random variable distributed approximately uniformly over the unit interval. This
28 subjective approach was motivated by (1) the consideration that differences in baseline
29 rates might reflect differential exposure to both cancer initiators (consistent with additive
30 transfer) and cancer promoters (consistent with multiplicative transfer) and (2) an almost
31 complete lack of relevant epidemiological information for most cancer sites. The general
32 EPA approach for site-specific cancer risk was similar, but on a logarithmic scale: the
33 logarithm of the excess risk was assumed to be a linear mixture between the logarithms of
34 the multiplicative and additive transfer model estimates (EPA, 1999), where the uncertain

1 mixture parameter p was assumed to be uniformly distributed over the unit interval. The
2 EPA approach tends to yield somewhat lower risk estimates than the NCI/CDC approach.
3 For the few sites where information on population transfer was available, the NCI/CDC
4 approach was to favor one simple transfer model over the other, e.g., for breast cancer,
5 0.5 probability was placed on additive transfer and 0.5 on the uniform model; for stomach
6 cancer, probability 0.33 was placed on multiplicative transfer and probability 0.67 on the
7 uniform model.

8 For all cancers except skin, as a group, the sex-age-standardized ratio of American
9 to Japanese rates was assumed to be 1.3 (Parkin, 1997). Multiplicative transfer of LSS-
10 based excess relative risk would involve applying the same ERR to U.S. baseline rates,
11 whereas for additive transfer the LSS-based ERR would be divided by 1.3 to obtain the
12 same absolute excess in the two countries. The resulting uncertainty distribution for ERR
13 at 1 Gy in a U.S. population, after application of the NCI/CDC approach, is
14 approximately lognormal with 90% limits 0.13-0.41, and mean 0.25 (Figure 6.5).

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6.2.5 DDREF

17 In general, epidemiological estimates of overall and site-specific cancer risk
18 related to radiation exposure are statistically consistent with a linear dose response
19 (leukemia, with a linear-quadratic dose response, is an exception). For the same reasons
20 that data restricted to low doses tend to be uninformative about radiation-related excess
21 risk, this apparent linearity does not rule out, on statistical grounds, the possibility of
22 increased, decreased, or even absent excess risk per unit dose at very low doses. For
23 various reasons discussed in Chapters 2 and 3, linear-model estimated excess risk is often
24 divided by a dose and dose-rate effectiveness factor (DDREF) at low doses and low dose
25 rates. The ICRP (1991) recommended a DDREF of 2 for radiation protection purposes,
26 and the United Nations Scientific Committee on Effects of Ionizing Radiation
27 (UNSCEAR, 1993) recommended that the chosen DDREF be applied to chronic
28 exposures at dose rates less than 6 mGy per hour averaged over the first few hours, and to
29 acute exposures at total doses less than 0.2 Gy. This recommendation was adopted by the
30 EPA (1999). Continuous, subjective uncertainty distributions for DDREF were used in
31 uncertainty analyses carried out for NCRP Report 126 (NCRP, 1997), the EPA (1999),
32 and by an expert committee advising the Colorado Department of Public Health and
33 Environment (Grogan, 2000) (Figure 6.6). The Grogan uncertainty distribution differs
34 from the NCRP distribution mainly in allowing a small probability that risk per unit dose

1 might increase at very low doses. Thus, the NCRP and EPA distributions allowed for the
2 possibility of DDREF values between 1 and 5, while the Grogan et al distribution
3 included DDREF values as low as 0.2. The uncertainty analysis for the revised NIH
4 radioepidemiological tables report postulated a discrete subjective uncertainty distribution
5 for DDREF, with non-zero probabilities assigned to 0.5, 0.7, 1.0, 1.5, 2.0, 3.0, 4.0, and
6 5.0 (Figure 6.7).

7 Application of a DDREF factor greater than 1 reduces estimated risk, and an
8 uncertain DDREF introduces additional uncertainty in estimated risk. Applying the
9 different DDREF assumptions summarized in Figures 6.6 and 6.7 to the adjusted
10 uncertainty distribution for risk in Figure 6.5 resulted in roughly lognormal uncertainty
11 distributions for ERR per Gy at low doses and dose rates, with mean values substantially
12 less than the mean value 0.25 for ERR at 1 Gy for acute exposures corresponding to the
13 uncertainty distribution in Figure 6.5, and upper 95% uncertainty limits somewhat less
14 than the value 0.41, also from Figure 6.5. By model, means and upper limits were 0.12
15 and 0.20, respectively, for the EPA DDREF, 0.11 and 0.23 for the NCRP model, mean
16 0.12 and upper limit 0.28 for the Grogan et al model, and mean 0.17 and upper limit 0.36
17 for the NCI/CDC model (Figure 6.8).

18

19

6.2.6 Variation by sex

20 The above results apply to males. Carrying out the same calculations based on the
21 statistical uncertainty presented in Section 6.2.1 for a female population yields an ultimate
22 uncertainty distribution, using the NCI/CDC DDREF model, with mean 0.355 and 95th
23 percentile 0.69. For a population divided equally by age and sex, the mean is 0.26 and
24 the upper limit is 0.50.

25

26

6.2.7 Expression of excess risk in absolute terms

27 For U.S. males, the lifetime baseline cancer risk, tabulated by National Cancer
28 Institute's SEER program (http://seer.cancer.gov/faststats/html/dev_all.html), from age
29 50 given cancer-free survival to age 40, is 45.3%. Thus, the estimated lifetime excess
30 cancer risk per Gy associated with a low-dose, low-LET, whole-body radiation exposure
31 is roughly lognormal with mean $0.17 \div 45.3\% = 7.7\%$ and 95th percentile $0.36 \div 45.3\% =$
32 16.3% ; the 5th percentile is $0.066 \div 45.3\% = 1.1\%$. For females, the corresponding
33 baseline risk is 35.5%, and the uncertainty distribution for lifetime EAR per Gy has mean

1 0.355 H 35.5% = 12.6% and 90% uncertainty bounds (0.146, 0.69) H 35.5% = (5.2%,
2 24.5%). For a population evenly divided by sex, the baseline risk from age 50 given
3 survival to age 40 is 40.4%, and lifetime EAR per Gy is roughly lognormal with mean
4 10.5% and 90% bounds (3.8%, 20.1%).

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6.2.8 Gradualism in DDREF and threshold effects.

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A rule that a DDREF should apply at acute doses below (say) 0.2 Gy and not at 0.2 Gy and above, or at dose rates less than 6 mGy per hour but not at dose rates marginally higher than that value, is contrary to experience with stochastic phenomena, and would be difficult for practical applications, e.g., in adjudicating compensation claims for radiation-related cancer. Accordingly, in the recent revision of the NIH radioepidemiological tables report (NCI/CDC 2003), DDREF was gradually phased in, from 1 to its (uncertain) full value, over an interval of decreasing dose of acute exposure. Similarly, a threshold dose, below which there is presumed to be no radiation-related risk, is generally not thought of as a value associated with the abrupt disappearance of risk, but a (possibly uncertain) value greater than zero Gy at which the gradual disappearance of excess risk with decreasing dose becomes complete. Thus, a threshold or possible threshold would, like the DDREF, be phased in gradually with decreasing dose.

For simplicity of presentation, phasing in DDREF and/or a threshold is ignored in the following discussion.

6.3 Allowing for the uncertain possibility of a threshold.

The threshold concept has practical importance only if the threshold dose is high enough to justify ignoring, for radiation protection purposes, a substantial range of exposures that would otherwise be of concern. A reasonable way to include the threshold concept in an uncertainty analysis is to multiply the uncertain dose-specific excess relative risk, adjusted for the DDREF and other factors discussed in the preceding paragraphs, by a threshold factor distributed as a Bernoulli random variable taking value zero with probability $p(D)$ and value one with probability $1 - p(D)$, where $0 \leq p(D) \leq 1$ and p is a possibly uncertain, decreasing function of radiation dose D . Some examples will illustrate the impact of uncertainties regarding whether a threshold exists or the dose level of that threshold.

Example 1--Threshold and dose level certain. Known threshold at 10 mGy: $p(D) = 1$ for $D \leq 10$ mGy and $p(D) = 0$ for $D > 10$ mGy (for simplicity, the threshold is not phased in as a function of D). Thus, the uncertainty distribution for excess risk assigns probability 1 to the value zero, below 10 mGy, and is the same as that without a threshold (e.g., the NCI/CDC distribution in Figure 6.8) above 10 mGy. The mean and 95% upper probability limit on ERR per Gy are unchanged above 10 mGy, but they are both zero below that dose level. This example represents the common conception held by those who believe there is a threshold, albeit the putative threshold dose level may differ from 10 mGy.

Example 2--Threshold uncertain but threshold dose level certain. A threshold may exist at 10 mGy; this possibility is assigned subjective probability \mathbf{p} , where \mathbf{p} is a known value such as 5%, 20%, 50%, or 80%. The uncertainty distribution of ERR/Gy risk below 10 mGy assigns probability \mathbf{p} to zero and, for all other possible values of ERR/Gy, $1 - \mathbf{p}$ times the probability that would be assigned if there were no threshold. For doses below 10 mGy, the mean of the uncertainty distribution is $1 - \mathbf{p}$ times the mean of the uncertainty distribution for ERR/Gy if there were no threshold (i.e., if \mathbf{p} were zero). The 95% upper uncertainty limit is given by $\text{limit} = F^{-1}((.95 - \mathbf{p}) / (1 - \mathbf{p}))$ for $\mathbf{p} < 0.95$, and $\text{limit} = 0$ for $\mathbf{p} \geq 0.95$, where F^{-1} is the inverse cumulative distribution function of the uncertainty distribution in the absence of a threshold (Land, 2002). Plots of the mean and

1 upper 95% limit, as functions of \mathbf{p} , are shown in Figure 6.9 for the approximate
2 lognormal uncertainty distribution for ERR/Gy according to the NCI/CDC model as
3 represented in Figure 6.8.

4 This example shows that, when the probability of a dose threshold is uncertain,
5 the central estimate of the ERR/Gy for low doses decreases linearly with an increasing
6 probability that there is a threshold -- but the 95% upper limit remains quite high until the
7 probability of a threshold reaches 80-90%, after which it falls sharply. This indicates that
8 unless there is consensus agreement that a threshold is very likely, the potential for an
9 appreciable low-dose risk cannot be ruled out.

10
11 **Example 3**--Threshold certain but its dose level uncertain. A threshold is known
12 to exist somewhere between 5 and 25 mGy, but otherwise is completely uncertain:
13 $p(D;D_0) = 1$ for $D \leq D_0$, and $= 0$ for $D > D_0$, where D_0 is an uncertain (random) quantity
14 uniformly distributed between 5 and 25 mGy. Estimated ERR/Gy is zero below 5 mGy,
15 but the probability assigned to non-zero values by the uncertainty distribution for risk at
16 dose D increases linearly from zero at $D = 5$ mGy to one (or to the value assigned in the
17 absence of a threshold) at $D = 25$ mGy. The uncertainty distribution for ERR/Gy assigns
18 probability 1 to zero for D below 5 mGy, 100% to the non-threshold distribution for doses
19 above 25 mGy, and probability $(25-D)/20$ to zero and probability $(D-5)/20$ to the non-
20 threshold uncertainty distribution, for $5 < D < 25$. The mean and upper 95% uncertainty
21 limit at dose D are as given in Example 2, and shown in Figure 6.9, for $\mathbf{p} = (25-D)/20$.

22 The third example illustrates an important point: even when one is certain there is
23 a dose threshold but is still uncertain as to the dose level at which it occurs, the low-dose
24 ERR/Gy behaves very similarly to the result for Example 2 (which had a fixed threshold
25 dose but uncertainty as to whether there was a threshold). Specifically, there still is some
26 probability that the low-dose ERR/Gy is appreciable.

27
28 **Example 3a**—There is a threshold for each individual in a population, but the
29 dose level varies by individual. Thus, for a randomly chosen individual from the
30 population, there is a threshold, but its dose level is uncertain. Mathematically, this
31 example is essentially the same as example 3.

32
33 **Example 4**--Threshold probability very uncertain but its dose level, conditional
34 on existence of a threshold, is certain. A threshold may exist at 10 mGy, with uncertain

1 probability. Enough is known (or there is a consensus among experts, which may be a
2 compromise) to characterize the subjective uncertainty distribution of $p(D)$, for $D < 10$
3 mGy; for example, as

- 4 *i)* uniform between 0 and 1: $U(0, 1)$,
- 5 *ii)* triangular between 0 and 1 with peak at 0: $Tr(0, 0, 1)$,
- 6 *iii)* $Tr(0, 0.25, 1)$ (peak at $p = 0.25$),
- 7 *iv)* $Tr(0, 0.5, 1)$,
- 8 *v)* $Tr(0, 0.75, 1)$,
- 9 *vi)* $Tr(0, 1, 1)$.

10
11 In example 4, the proportion of the uncertainty distribution for ERR/Gy assigned
12 to zero is randomly distributed over the unit interval, and the mean and upper 95% limit
13 of the resulting distribution depends on the assumed distribution of p . Figure 6.10 shows
14 Monte Carlo estimates of the resulting uncertainty distributions for ERR/Gy for the six
15 cases, again using the NCI/CDC non-threshold distribution from Figure 6.8, and the
16 corresponding means and upper 95% uncertainty limits.

17 The probability distributions in Figure 6.10 show, not unexpectedly, that if the
18 consensus uncertainty distribution of p gives a high weight to the likelihood of a threshold
19 (e.g., subjective distribution *vi*), then the distribution of the low-dose ERR/Gy is weighted
20 toward small values, whereas the opposite is true when the probability of a threshold is
21 less likely (e.g., subjective distributions *ii* or *iii*). Nevertheless, even for distribution *vi*
22 the mean expected low-dose ERR/Gy of 5.7% is about a third as great as under the LNT
23 hypothesis (ERR/Gy = 17%), and it is between 40% and 70% as the LNT value for
24 distributions *i* – *v*.

25 **Example 5** – Dose-dependent uncertain probability of a threshold. Suppose that
26 the uncertainty distribution for a threshold at 10 mGy corresponds to Example 4,
27 distribution *ii*, i.e., $Tr(0,0,1)$, and that the uncertainty distributions for thresholds at 1
28 mGy and at 0.1 mGy correspond to Example 4 distributions *iv* ($Tr(0, 0.5, 1)$) and *vi* ($Tr(0,$
29 $1, 1)$), respectively. Then the subjective means and upper uncertainty limits for ERR/Gy
30 would be 11.5% and 27%, respectively, at 10 mGy, 8.6% and 21% at 1 mGy, and 5.7%
31 and 17% at 0.1 mGy. The corresponding values of ERR would be 0.115% and 0.27% at
32 10 mGy, 0.0086% and 0.021% at 1 mGy, and 0.00057% and 0.0017% at 0.1 mGy. The
33 mean and 95% upper limit for ERR at 0.1 mGy can be compared with the mean 0.0017%
34 and upper limit 0.0036% according to the LNT hypothesis.

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Of the five examples above, Example 5 probably best reflects our present state of knowledge about low dose risk, namely, that we are uncertain about the likelihood of a dose threshold, and that in addition, if there should be a dose threshold, we are uncertain about at what dose level it would be. However, as a counter to an agnostic viewpoint, it should be noted that the mechanistic and experimental data discussed in this monograph tend to give weight to a nonthreshold model, as do the solid tumor data in the Japanese atomic bomb study. (In addition to apparent linearity of dose response down to doses below 100 mGy, an analysis by Pierce and Preston (2000) found that a threshold above 60 mGy would be statistically inconsistent with LSS dose-response data for all solid cancers combined.)

6.4 Conclusions

Information on radiation-related cancer risk is needed (1) as guidance for radiation protection efforts, (2) as a basis for informed consent by persons who may be asked to accept a certain level of exposure in the interests of medical research, economic progress, or some other social good, (3) for adjudication of claims and disputes concerning cases of disease possibly related to past radiation exposure, and (4) for risk-benefit analyses of public policy initiatives related to radiation. As mentioned previously in this report, these issues are essentially political, in the sense that different people have different interests and points of view which must be taken into consideration when policies are developed. Moreover, implementation of such policies inevitably involves accommodation and consensus, and it is important that the policies are seen to be derived fairly and openly, on the basis of facts and assumptions that are wholly accessible to those affected by implementation.

Information useful for these purposes includes central estimates of dose-specific risk, but also, lower and (especially) upper probability bounds on risk. Probability bounds can reflect both statistical uncertainty, estimated by fitting a mathematical model to observational data, and subjective uncertainty that may take into account model assumptions that are necessary to calculate estimates but are themselves uncertain. Probability bounds provide a level of transparency substantially beyond that provided by a point estimate, such as the expected (mean) value of the uncertainty distribution for estimated excess risk. A lower probability bound (e.g., a 95% lower confidence limit or

1 uncertainty limit) greater than zero is evidence that there really is an excess risk;
2 however, the carcinogenicity of ionizing radiation exposure is already well established. A
3 lower bound corresponding to a risk that is intolerably high would, of course, be evidence
4 in support of diversion of financial resources for exposure reduction, even from the
5 viewpoint of those who would bear the expense.

6 From the viewpoint of those who would bear the risk, if any, associated with
7 exposure, and of those responsible for their protection, the questions of interest concern
8 (1) the extent to which risks associated with a given level of exposure are low enough to
9 be tolerated in view of competing risk and loss of benefits associated with avoidance of
10 that exposure, and (2) whether we can conclude that there is no risk at all associated with
11 a given exposure. An upper probability bound, if less than some “tolerable” level of risk,
12 can be used to help justify a favorable risk-benefit assessment for a particular exposure,
13 and can provide a margin of safety in decisions regarding risk protection or informed
14 consent related to possible hazards of radiation exposure. An upper probability bound of
15 zero or less would be evidence in favor of a threshold or, more likely, a beneficial effect
16 of low-dose radiation.

17 The implications of a possible, but uncertain, low-dose threshold for radiation
18 protection are summarized by the dependence of the mean value and the upper 95%
19 probability limit on the presumed threshold probability value (Figure 6.9) or on the
20 uncertainty distribution for that probability (Figure 6.10). The mean value of estimated
21 ERR/Gy is proportional to $1-p$ for known threshold probability p and proportional to $1-$
22 $E(p)$ for an uncertain threshold probability p with expected value $E(p)$. Thus, the effect
23 on the mean value is the same as that of an assumed constant DDREF equal to $1/p$ or
24 $1/E(p)$. The effect on the upper 95% probability limit is less drastic, unless the assumed
25 probability of a threshold is high. As shown in Figure 6.9, the upper limit decreases with
26 increasing p , but not nearly as steeply as for the mean until p approaches the
27 probability level of the upper limit, e.g., about 0.85 in the case of a 95% limit.

28 Obviously, the lower 95% limit (the 5th percentile of the distribution) is zero for $p \geq 0.05$.

29 As mentioned earlier in this chapter, an established, universal or near-universal,
30 low-dose threshold for radiation-related cancer risk would obviate concern about risks
31 from exposures at doses lower than the threshold value. Our present information,
32 summarized in NCRP Report 136 (2001) and the present report, offers little support for
33 the existence of a universal low-dose threshold, but it cannot be ruled out as an uncertain

1 possibility. However, the implications of the uncertain possibility of a threshold are
2 qualitatively not much different from those of an uncertain DDREF: central values and
3 upper uncertainty limits are reduced somewhat, but they do not become zero. Moreover,
4 the argument that radiation protection standards should be relaxed “because it is possible
5 that there may not be *any* risk at low doses” is unlikely to be persuasive to persons who
6 are concerned about the possibility that risk associated with very low doses may be
7 unacceptably high, and it may undermine the more realistic argument that the risk, which
8 is understood rather well compared to that associated with other common carcinogens, is
9 almost certainly less than some stated value which may be considered tolerable, for
10 various reasons such as economic benefits or consideration of risks associated with
11 alternative strategies involving less exposure.
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7. CONCLUSIONS

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3 1. Epidemiological studies of cancer risk following radiation exposure provide the
4 primary basis for estimation of radiation-related risk in human populations. These
5 studies demonstrate the existence of dose response and its modification by other
6 factors, and show some variation by cancer site and by histological subtypes
7 within sites. At low and very low radiation doses, statistical and other variation in
8 baseline risk tends to be the dominant source of error in both epidemiological and
9 experimental carcinogenesis studies, and estimates of radiation-related risk tend to
10 be highly uncertain both because of a weak signal-to-noise ratio and because it is
11 difficult to recognize or to control for subtle confounding factors. Thus,
12 extrapolation of risk estimates based on observations at moderate to high doses
13 continues to be the primary basis for estimation of radiation-related risk at low
14 doses and dose rates.
15
- 16 2. There is no direct evidence, from either epidemiological or experimental
17 carcinogenesis studies, that radiation exposure at doses on the order of 1 mGy or
18 less is carcinogenic, nor would any be expected because of the considerations
19 outlined in Conclusion 1. There is, however, epidemiological evidence, unlikely
20 on the whole to be an artifact of random variation, linking increased cancer risk
21 to exposures at doses on the order of 10 mGy. This evidence includes several
22 case-control studies of leukemia and solid cancers among different populations of
23 children exposed in utero to x-ray pelvimetry, cohort studies of breast cancer
24 among women given multiple fluoroscopy examinations during treatment for
25 tuberculosis or scoliosis, with average breast doses on the order of 10 mGy per
26 examination, and the observation that risk of mortality and morbidity among
27 atomic bomb survivors from all solid cancers combined is linear in radiation dose
28 down to about 100 mGy.
29
- 30 3. Overall, relevant animal tumor data from experimental carcinogenesis studies tend
31 to support a dose response that, at low doses, is linear with no threshold. This
32 inference does not conflict with experimental evidence for reductions in excess
33 risk per unit dose at low doses or with fractionation and/or protraction of dose.
34 Recent cytogenetic and molecular studies provide direct support for the view that

1 the critical radiation-associated events in the tumorigenic process are
2 predominantly early events involving DNA losses targeting specific genomic
3 regions harboring critical genes. The predominant importance of DNA DSB
4 induction and post-irradiation error-prone NHEJ repair for the induction of
5 aberrations, and the apparently critical role for radiation-induced aberrations in the
6 pathogenesis of cancer in these experimental models, argue against the
7 proposition of a low dose threshold in the dose response.

- 8
- 9 4. There is evidence from both epidemiological and experimental studies that
10 specific tissues and cancer sites may for various reasons vary from the general
11 rule articulated in Conclusions 1-3, in the sense that radiation carcinogenesis is
12 markedly and disproportionately less likely to occur at low doses than at high, and
13 may even suggest a threshold. Examples are the small intestine, bone, and skin.
14 However, these appear to be the exception rather than the general rule;
15 experimental studies of radiation-related life shortening, which represent the
16 integrated dose responses for a variety of tumor types, suggest linear dose
17 responses over a wide range of doses.
- 18
- 19 5. Ionizing radiation is able to produce a unique type of damage in which multiple
20 lesions are encountered within close spatial proximity. Even a single track through
21 a cell is likely to induce these unique clustered damages. This type of damage may
22 not be generated frequently endogenously or by other exogenous agents, and thus,
23 there may not have been a strong selective pressure driving efficient repair.
24 Although cells have a vast array of damage response mechanism that facilitate the
25 repair of DNA damage and the removal of damaged cells, these mechanisms are
26 not fool proof. Moreover, clustered radiation-induced lesions pose a particular
27 problem and current emerging evidence suggests that closely spaced lesions can
28 compromise the repair machinery. On this basis, there is not any strong evidence
29 for a radiation dose below which all radiation-induced damage can be repaired
30 with fidelity.
- 31
- 32 6. Although many of the cells containing such radiation-induced damage may be
33 eliminated by damage response pathways involving cell cycle checkpoint control
34 and apoptotic pathways, it is clear from analysis of cytogenetics and mutagenesis

1 that damaged or altered cells are capable of escaping these pathways and
2 propagating. This further argues against the likely possibility of a threshold for
3 radiation-induced cellular effects.

- 4
- 5 7. The processing and misrepair of radiation-induced DSBs, particularly complex
6 forms, are responsible for chromosome/gene alterations that manifest as
7 chromosome aberrations and mutations. Current understanding of mechanisms
8 and quantitative data on dose and time-dose relationships support a linear dose
9 response at low doses with no compelling evidence for the existence of a
10 threshold dose below which there would be no effect.
- 11
- 12 8. When considered as a whole, the emerging results with regard to radiation-related
13 adaptive response, genomic instability, and bystander effects suggest that the risk
14 of low level exposure to ionizing radiation is uncertain, and a simple extrapolation
15 from high dose effects may not be wholly justified in all instances. However, a
16 better understanding of the mechanisms for these phenomena, the extent to which
17 they are active *in vivo*, and how they are interrelated is needed before they can be
18 evaluated as factors to be included in the estimation of potential risk to the human
19 population of exposure to low levels of ionizing radiation.
- 20
- 21 9. Probability limits on risk provide additional information relevant to radiation
22 protection. In particular, a high lower limit attests to the reality of danger
23 associated with a given exposure, and a low upper limit provides assurance as to
24 the relative safety, and presumably the acceptability, of the exposure when seen in
25 the context of other hazards of daily life. The information reviewed in this report,
26 from epidemiology and from experimental studies of animal, cellular, and
27 molecular models, is consistent with proportionality between radiation-related
28 cancer risk at low doses and at low dose rates, including the dose delivered by a
29 single photon. It is also consistent, given uncertainties about the roles played by
30 repair and apoptosis at very low doses, with the existence of a dose threshold at a
31 dose level so low that radiation-related risk under the linear, no-threshold (LNT)
32 hypothesis would be statistically indistinguishable from random variation in
33 baseline risk. However, the uncertain possibility of a threshold does not drastically
34 reduce either central estimates or upper probability limits for low-dose risk

1 compared to those obtained using the LNT hypothesis, unless that possibility is
2 assumed to be very likely. To the contrary, the evidence reviewed in this report
3 suggests that a universal low-dose threshold above (say) 1 mGy of low-LET
4 radiation is an unlikely possibility. Thus, the LNT hypothesis remains the most
5 prudent risk model for guidance of radiation protection.

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TABLES

Table 2.1 Some sources and amounts of ionizing radiation exposure (unless noted, from Mettler and Upton (1995))

Exposure	Effective dose, in mSv	
	Normal background areas	High background areas
Natural background (world population)		
Cosmic rays	0.38 / year	2.0 / year
Terrestrial (rays	0.46 / year	4.3 / year
Radionuclides in tissue	0.25 / year	0.6 / year
Inhaled radionuclides	2.5 / year	
Medical diagnostic (U.S. population)	Per exam	
Skull	0.22	
Cervical spine	0.20	
Chest	0.08	
Cholioangiogram	1.89	
Lumbar Spine	1.27	
Upper gastrointestinal series	2.44	
Abdomen (KUB)	0.56	
Barium enema	4.06	
Intravenous pyelogram	1.58	
Pelvis	0.44	
Hip	0.83	
Extremities	0.01	
CT scan, head or body	1.11	
Pediatric CT scan, abdomen ¹	25 (stomach dose)	
Single screening mammogram ¹	3 (breast dose)	
Astronaut, 3-day space shuttle mission ²	2 – 3	
Astronaut, 60-day space station mission ²	50	
Average cumulative occupational dose in monitored radiation workers ³	Cumulative reported badge dose 20	
Average neutron-weighted colon dose for LSS population with doses between 0.005 and 4 Gy ⁴	Colon dose 200	

¹ Brenner et al, 2003; ² NCRP Report 138, 2001; ³ Gilbert, 2001; ⁴ Preston, 2003; computed using data set downloaded from RERF web site (Radiation Effects Research Foundation, 2003)

Table 2.2. Parameter estimates corresponding to the general dose-response model,

$$ERR(D) = \alpha D \times (1 + \beta D) \times \exp(-\gamma D),$$
where D is neutron-weighted (weight = 10), reconstructed radiation dose to the colon from the atomic bombings and ERR(D) is the dose-related excess relative risk of solid cancer morbidity, 1958-87, among members of the Radiation Effects Research Foundation's Life Span Study cohort of survivors of the bombings.

Parameter	Estimate	90% CI	p-value
α	0.52	0.16, 0.83	.02
β	0.94	0*, 6.8	.28
γ	0.84	0*, 0.68	.07
α	0.71	0.56, 0.87	<.001
β	0 [§]	----	----
γ	0.11	0*, 0.24	.07
α	0.57	0.48, 0.68	<.001
β	0*	----	----
γ	0 [§]	----	----
Analysis restricted to survivors with estimated doses 2 Sv and less			
α	0.40	0*, 0.85	.24
β	0.92	0*, 3.0	>.5
γ	0.53	0*, 1.3	>.5
α	0.61	0.35, 0.76	<.001
β	0.045	0*, 0.68	>.5
γ	0 [§]	----	----
α	0.64	0.54, 0.74	<.001
β	0 [§]	----	<.001
γ	0*, 0 [§]	----	<.001

* Estimate constrained to be \$0. [§]Estimate set = 0.

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Table 2.3 Modification of radiation-related risk by individual and lifestyle factors, and by other exposures.

Organ site / cancer	Population	Factor	Main factor effect on risk	Interaction with radiation exposure	References
Female breast	LSS cohort	Young age at 1 st full-term pregnancy	Decreased	Multiplicative ¹	Land et al, 1994
“	“	Multiple births	Decreased	Multiplicative ¹	“
“	“	Lengthy lactation history	Decreased	Multiplicative ¹	“
“	NY mastitis series	Assoc with 1 st delivery	Increased	Not tested	Shore, 1980
“	Massachusetts TB fluoroscopy series	Exposed yr of 1 st delivery	Increased (NS)	Not tested	Boice & Stone, 1978
Lung& bronchus	LSS cohort	Smoking history	Increased	Additive ²	Pierce, 2003
“	U.S. Uranium miners	“	“	NS closer to mult. than to additive	Lubin & Steindorf
Basal cell skin ca.	LSS cohort	Sun-exposed cf. covered areas of skin		Additive ²	Ron, 1998
“	NY tinea capitis series	White cf. Black patients	Higher in whites	Multiplicative ¹	Shore, 2002
Liver	LSS cohort	Hepatitis C infection	Increased	Strongly synergistic	Sharp, 2002
Female breast	LSS cf. Euro/American pops	Population rates	Japan rate 4-fold < US	Additive ²	Preston, 2002
Stomach	LSS cf. US peptic ulcer patients	Population rates	Japan rate 12-fold > US rate	NS, closer to multiplicative than to additive	Carr, 2002

6 ¹ Additive interaction model rejected (statistically inconsistent with data); ² Multiplicative interaction model
7 rejected

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Table 2.4. Statistical power calculations for a hypothetical study in which baseline cancer risk is known to be 10%, and the unknown radiation-related excess risk is 10% at 1 Gy and proportional to dose between 0 and 1 Gy.

Radiation dose	Excess risk	Total risk	Standard deviation of the estimated excess risk under the null and alternative hypotheses		Population size <i>N</i> needed for 80% power to detect the excess risk at the 5% significance level
			$0.316 / N^{1/2}$	$0.447 / N^{1/2}$	
1 Gy	10%	20%	$0.316 / N^{1/2}$	$0.447 / N^{1/2}$	124
100 mGy	1%	11%	$0.316 / N^{1/2}$	$0.332 / N^{1/2}$	6,800
10 mGy	0.1%	10.1%	$0.316 / N^{1/2}$	$0.318 / N^{1/2}$	624,000
1 mGy	0.01%	10.01%	$0.316 / N^{1/2}$	$0.316 / N^{1/2}$	61.9 million

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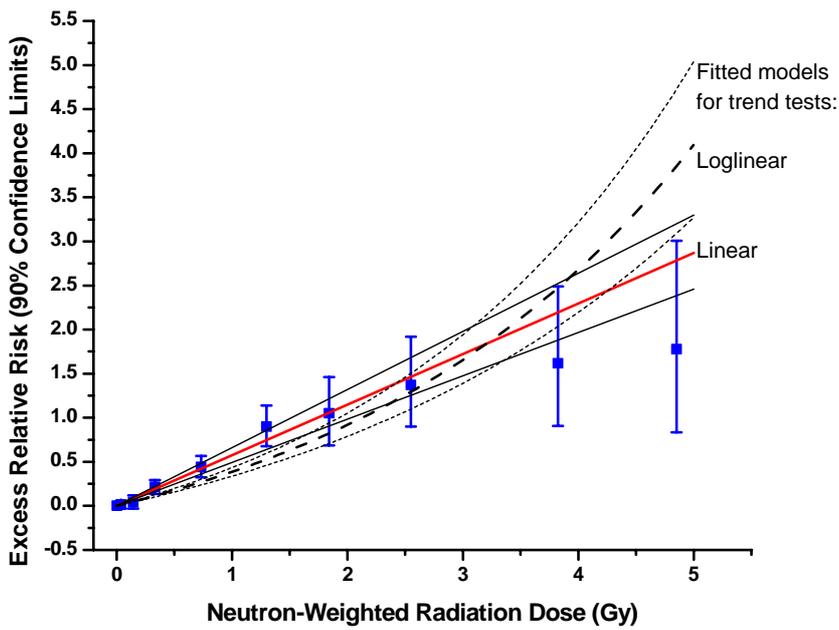
Table 2.5. Distribution of subjects, solid cancers, and estimated radiation-associated, excess solid cancers among 79,901 exposed members of the Life Span Study cohort of Hiroshima-Nagasaki atomic bomb survivors (Pearce and Preston, 2000).

Estimated colon dose	Number of subjects	Number of solid cancers	Estimated number of radiation-associated excess cancers*
Exposed beyond 3000 m	23,493	3,230	0
<5 mGy, exposed within 3000 m	10,159	1,301	1
5-100 mGy	30,524	4,119	77
100-200 mGy	4,775	739	60
200-500 mGy	5,862	982	164
0.5-1 Gy	3,048	582	177
1-2 Gy	1,570	376	165
>2 Gy	470	126	80

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* Fitted values, linear dose response

FIGURES



1
2 Figure 2.1. Dose-specific excess relative risk of solid cancer among atomic bomb
3 survivors, 1958-87, by interval of neutron-weighted, estimated radiation dose to the
4 colon. Fitted dose-response functions correspond to statistical tests of increasing trend
5 according to the linear ($RR = 1 + \alpha D$) and log-linear ($RR = \exp(\beta D)$) dose-response
6 models. The baseline risk is adjusted for city of exposure (Hiroshima or Nagasaki), sex,
7 and 5-year intervals of exposure age and age at observation for risk, using a saturated
8 model.

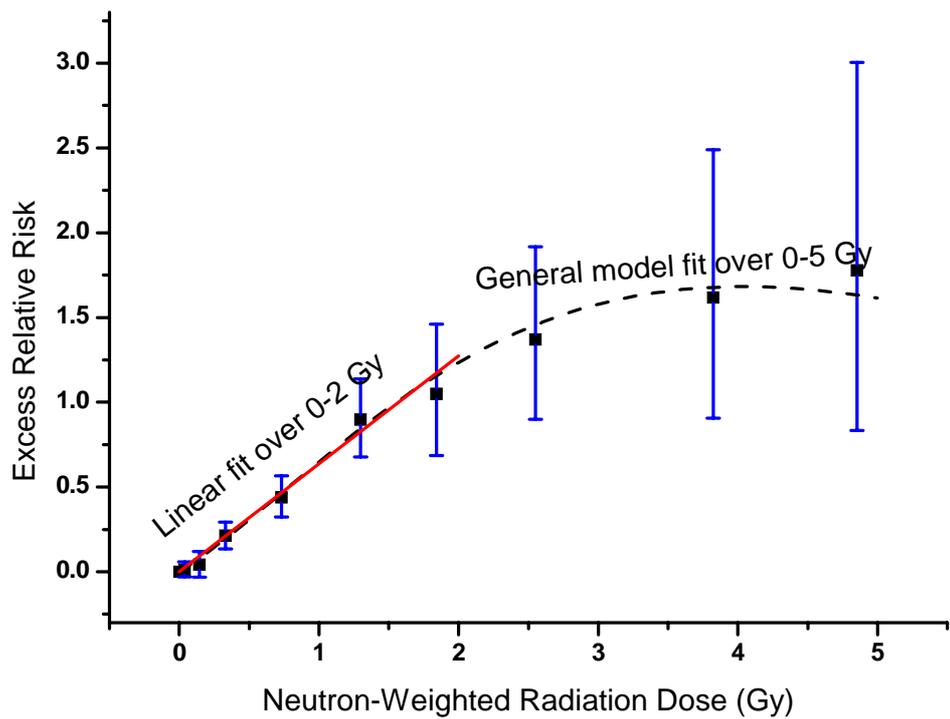
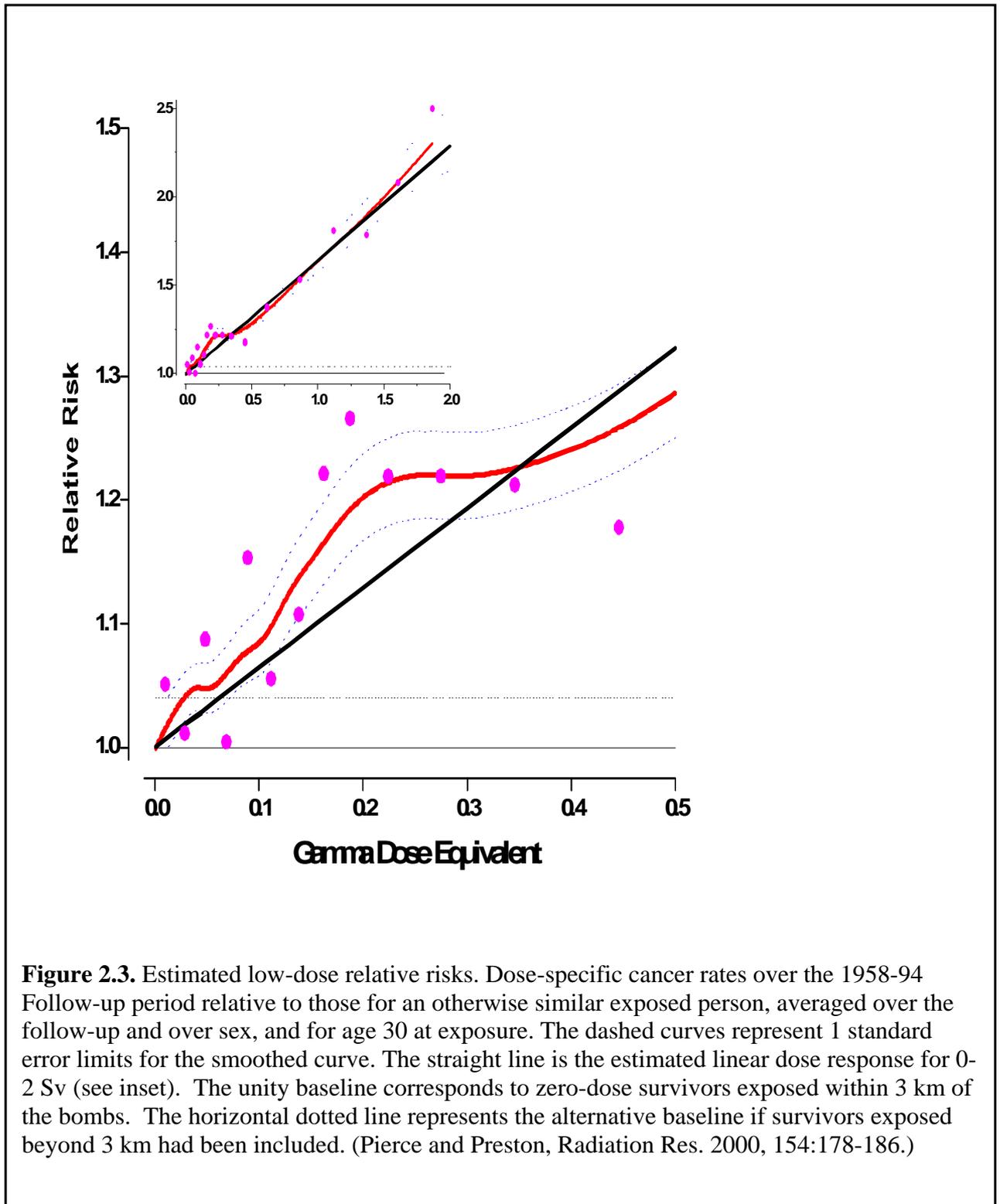
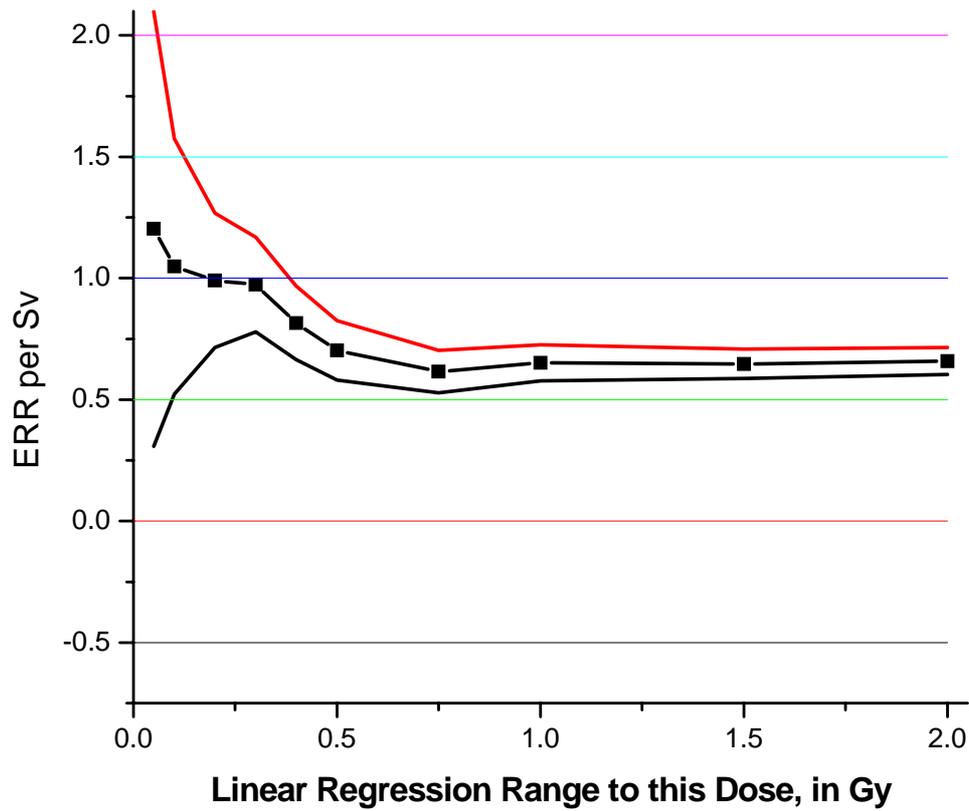


Figure 2.2. General dose-response model, $ERR(D) = \alpha D \times (1 + \beta D) \times \exp(-\gamma D - \delta D^2)$, fit to the dose-response data of Figure 1, and linear dose-response model, $ERR(D) = \alpha D$, fit to the data subset restricted to radiation doses between zero and 2 Sv. Details of the parameter estimates are given in Table 2.2.

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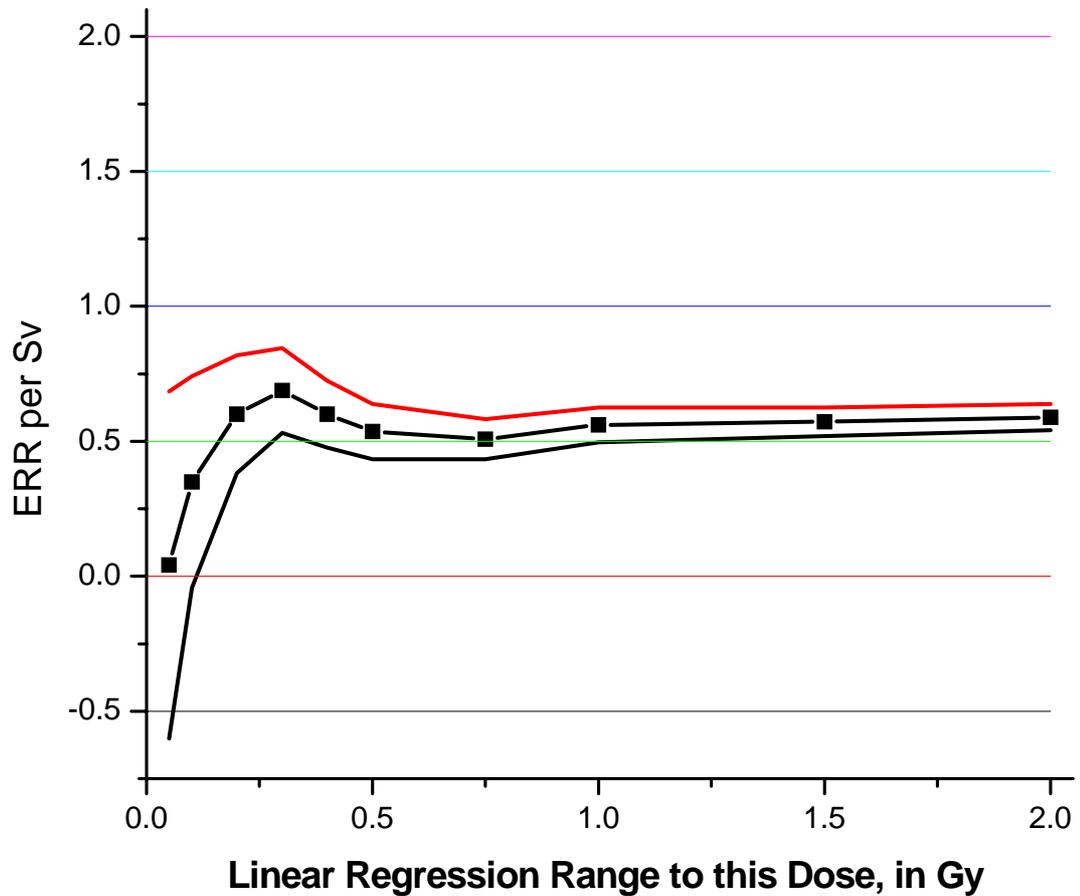


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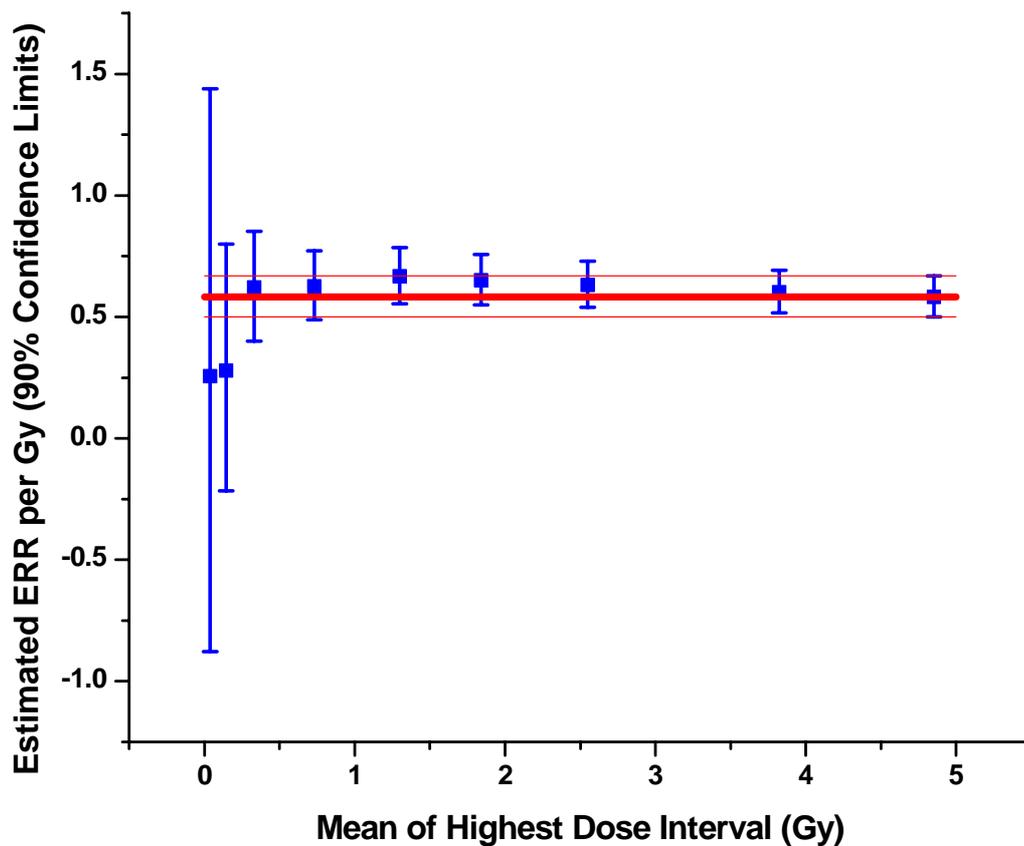
Figure 2.4. Linear regression estimates of the ERR per Gy (points and connecting line, with error bounds of \pm one SE) for solid cancer incidence, based on Poisson regression over dose intervals of differing ranges from zero to the horizontal coordinate of the plotted point. The analysis is limited to proximal survivors exposed at distances under 3 km.



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3 Figure 2.5. Linear regression estimates of the ERR per Gy (points and connecting line,
 4 with error bounds of \pm one SE) for solid cancer incidence, based on Poisson regression
 5 over dose intervals of differing ranges from zero to the horizontal coordinate of the
 6 plotted point. The analysis is based on all exposed survivors with estimated radiation
 7 doses less than 2 Gy.

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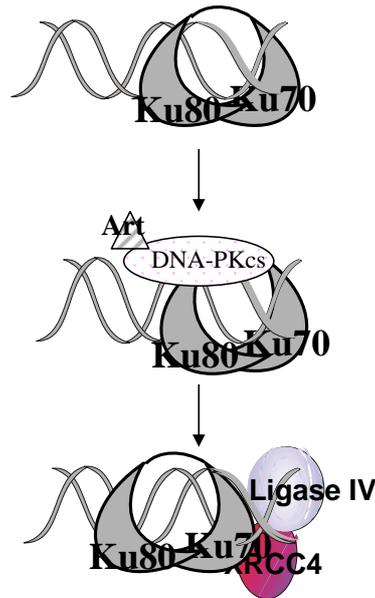


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 2 Figure 2.6. All-age linear regression estimates of ERR per Gy for female breast cancer
 3 assuming a 12-year minimum latent period, with dose-specific data trimmed from the
 4 right. Horizontal placement corresponds to the mean breast tissue dose for the highest
 5 neutron-weighted kerma interval included in the regression. Thus, the right-most point
 6 corresponds to the full dose range, the next point to the left to doses under 4 Gy, the next
 7 to doses under 3 Gy, and so on.

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FIGURE 3.1



1) binding of Ku to the double stranded DNA end. The crystal structure of Ku shows that the DNA passes through a cavity in the structure with Ku encircling the DNA

(a single DNA end is shown for simplicity)

2) DNA-PKcs is recruited and the kinase activity activated. Autophosphorylation and phosphorylation of artemis likely ensues, potentially leading to release of DNA-PKcs Artemis nuclease activity may enhance processing of ends.

3) Ku enhances the recruitment of DNA ligase IV/XRCC4 complex and Ku translocates inwards to allow ligase IV/XRCC4 access to the DNA ends.

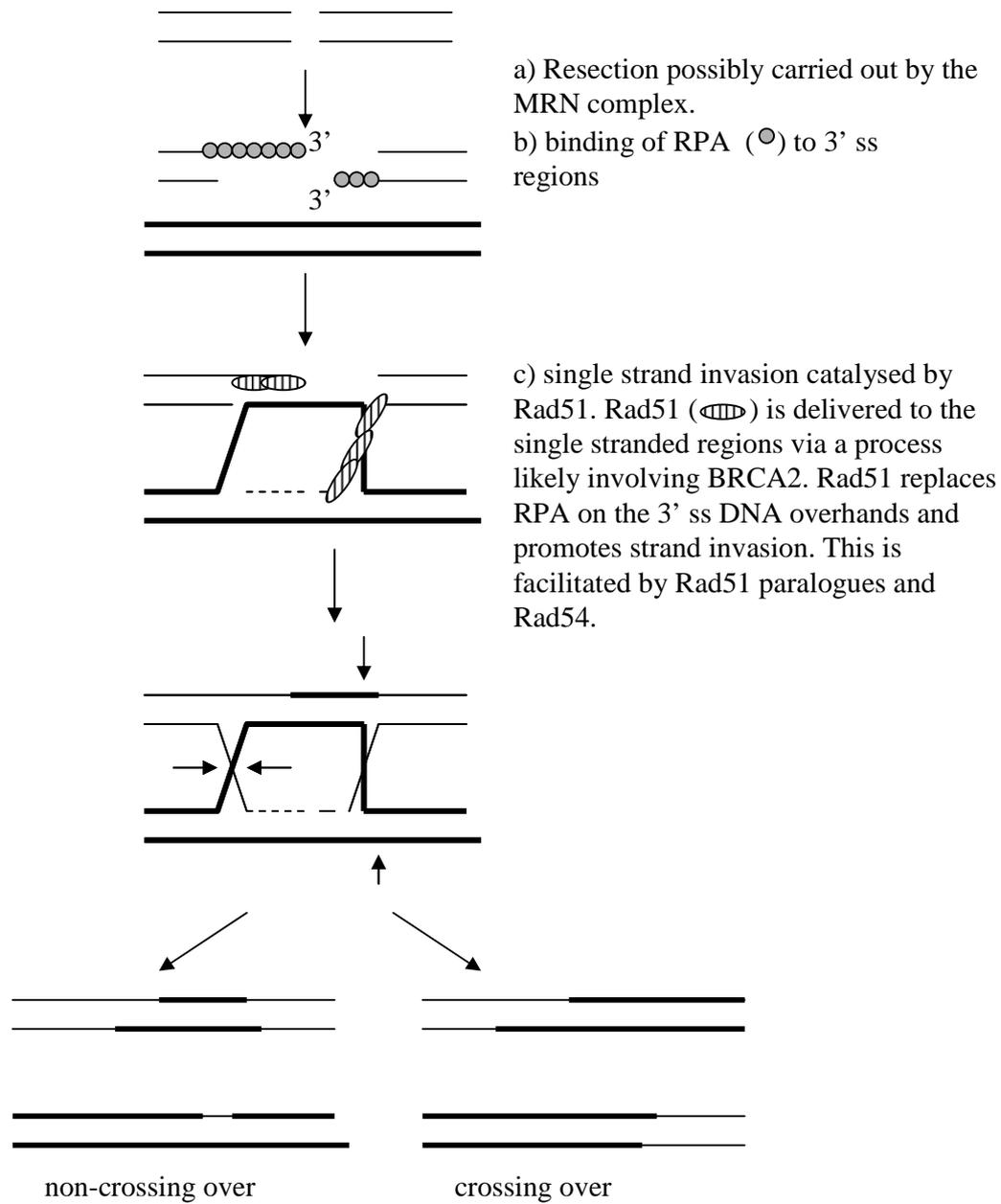
Fig. 3.1. Model for DNA NHEJ.

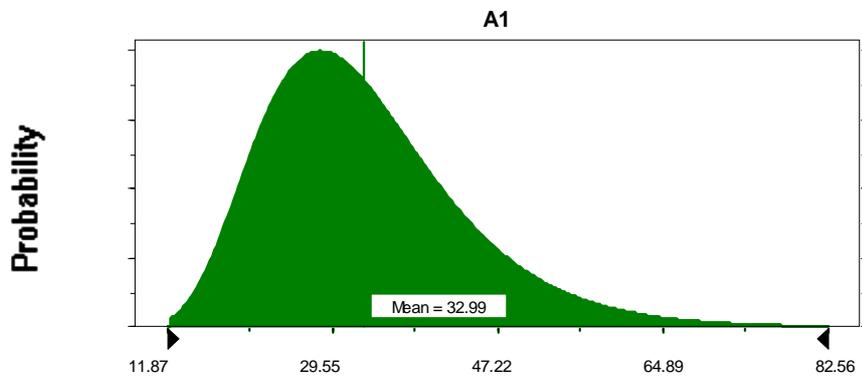
Proposed steps involved in the process

Note that only a single DNA end is shown for simplicity - one function of DNA-PKcs may be to enhance synapsis of the DNA ends.

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Figure 3.2. Depiction of homologous recombination.



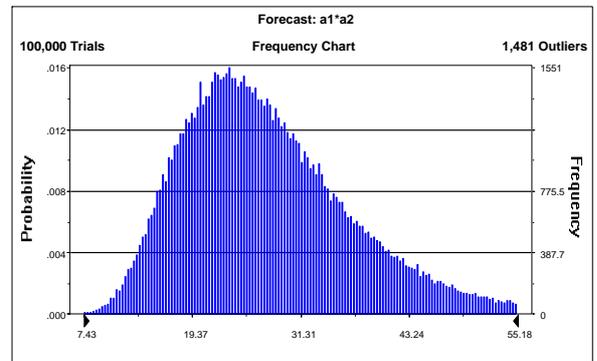
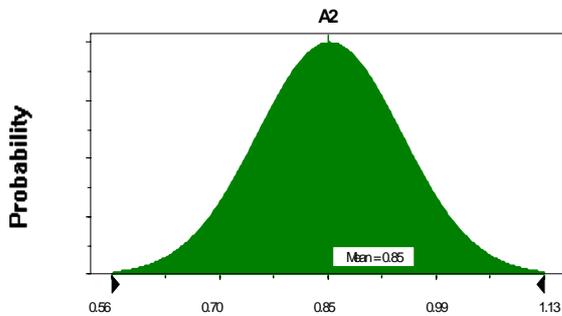


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10 Figure 6.1. Lognormal distribution with 90% confidence limits 18%-43%, representing
 11 statistical uncertainty about percent cancer excess relative risk per Gy.

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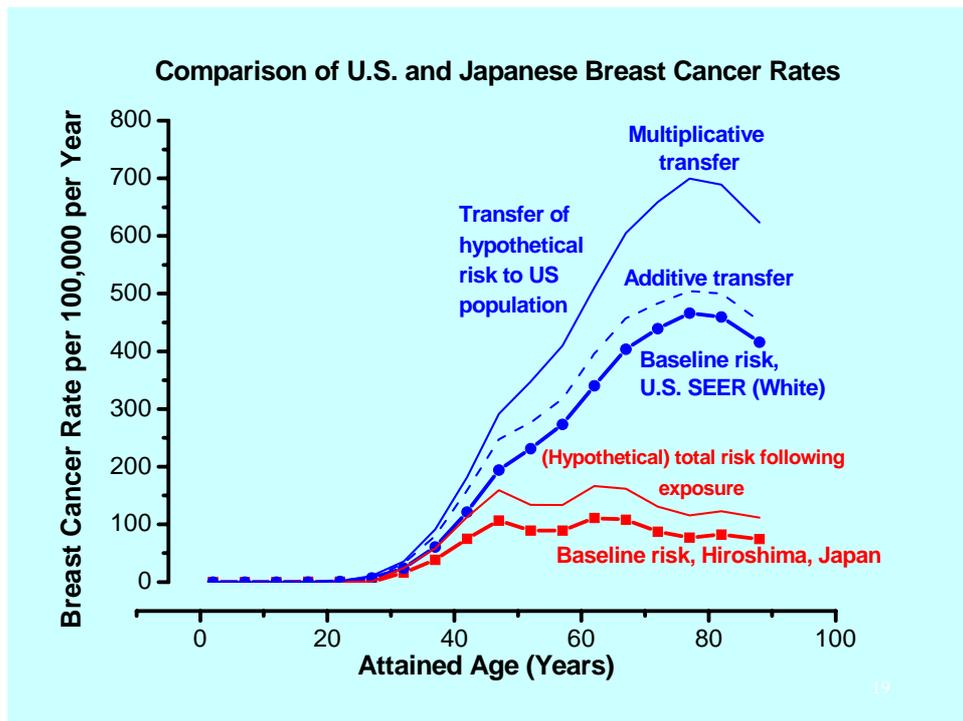
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Figure 6.2. Normal uncertainty distribution for dosimetry bias correction factor, with mean 0.84 and 90% uncertainty limits 0.69-1.00.

Figure 6.3. Approximately lognormal uncertainty distribution for ERR per Gy corrected for dosimetry bias, with mean 0.26 and uncertainty limits 0.15 – 0.46.

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Figure 6.4. Comparison of age-specific baseline rates for female breast cancer incidence in Japan and the United States (lower and upper polygonal lines with data points, respectively), estimated rate following a hypothetical radiation exposure of a Japanese population at age 15 (lower solid polygonal line without data points), and estimates for a U.S. population obtained by additive (dashed curve) and multiplicative transfer (upper solid curve) of estimated excess risk from the Japanese to the U.S. population.

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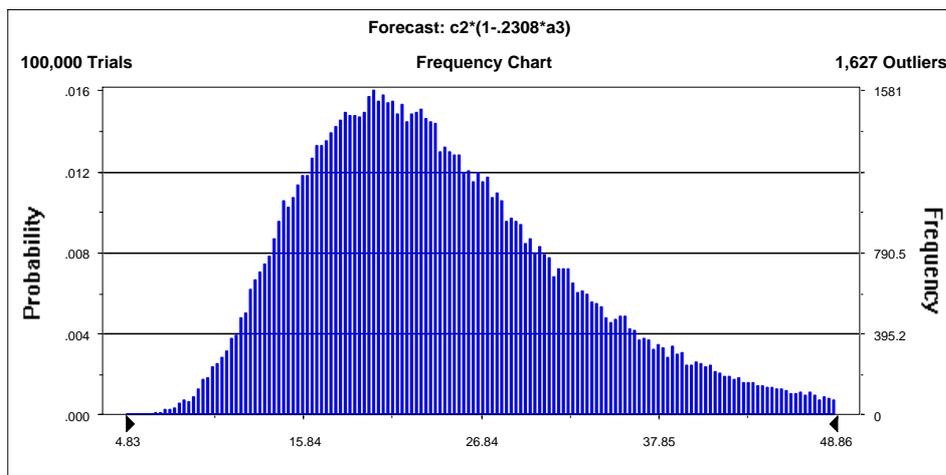


Figure 6.5. Monte Carlo simulation of the uncertainty distribution for cancer ERR (in percent) at 1 Gy, after transfer to a U.S. population: the simulated distribution is approximately lognormal with mean 0.25 and 90% probability limits 0.13-0.41.

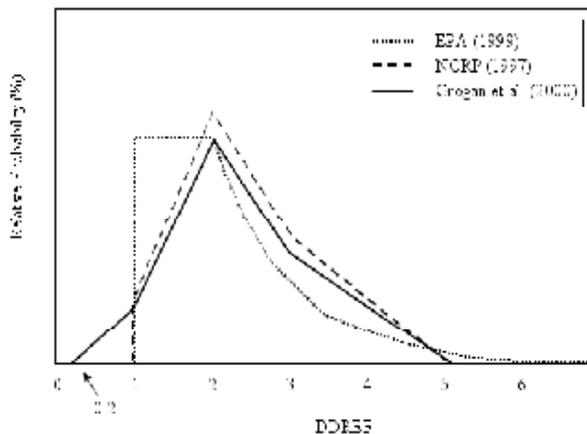


Figure 6.6. Continuous subjective uncertainty distributions for DDREF used in recent analyses.

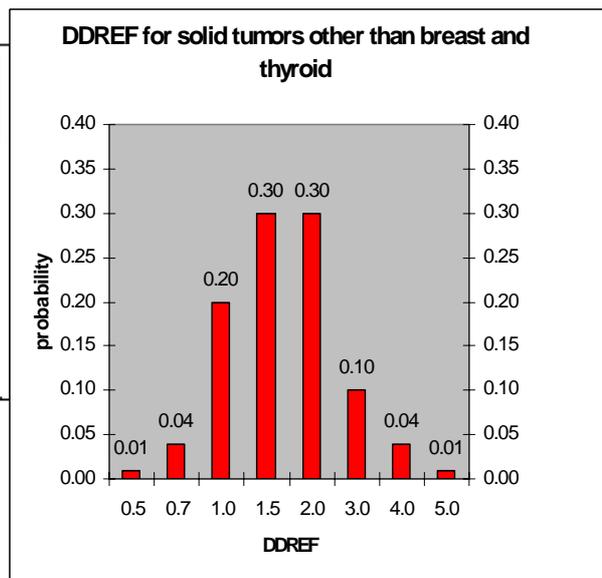


Figure 6.7. Discrete uncertainty Distribution for DDREF used in NCI/CDC (2003) analysis.

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Figure 6.8. Influence of DDREF assumptions on uncertainty for ERR/Gy (in percent).

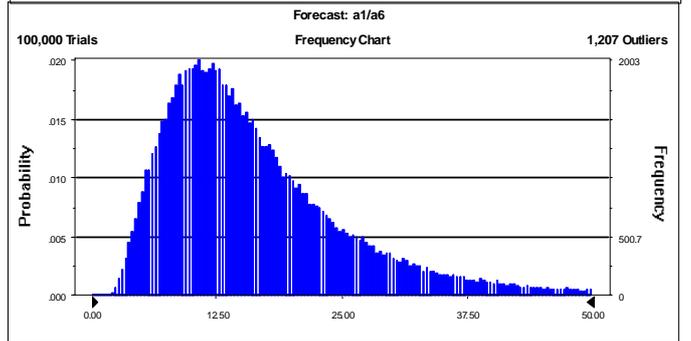
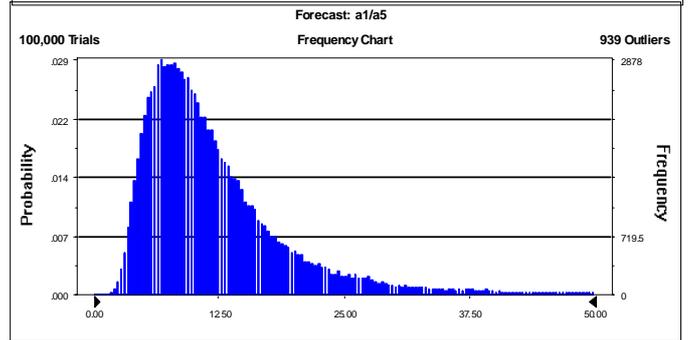
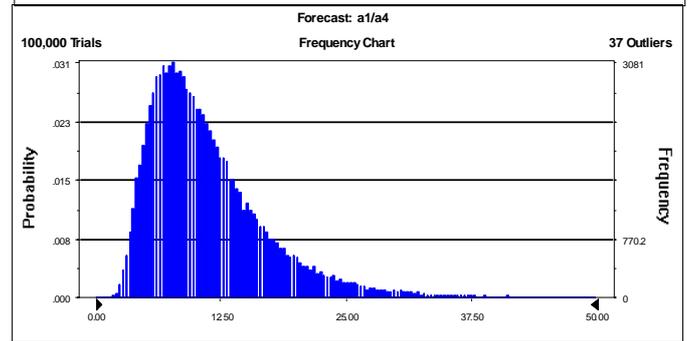
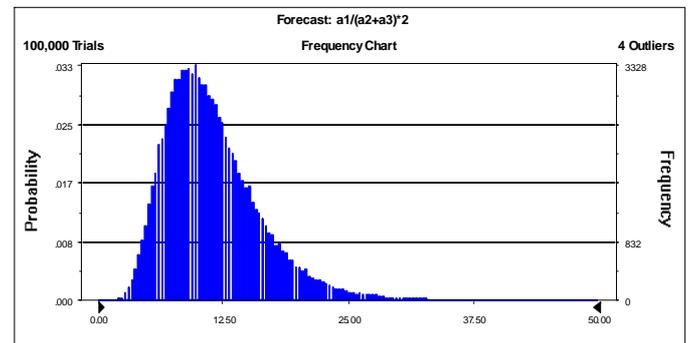
DDREF model Uncertainty. for
 (Source) ERR/Gy:
 Mean 95% limit

EPA (1999) 12% 20%

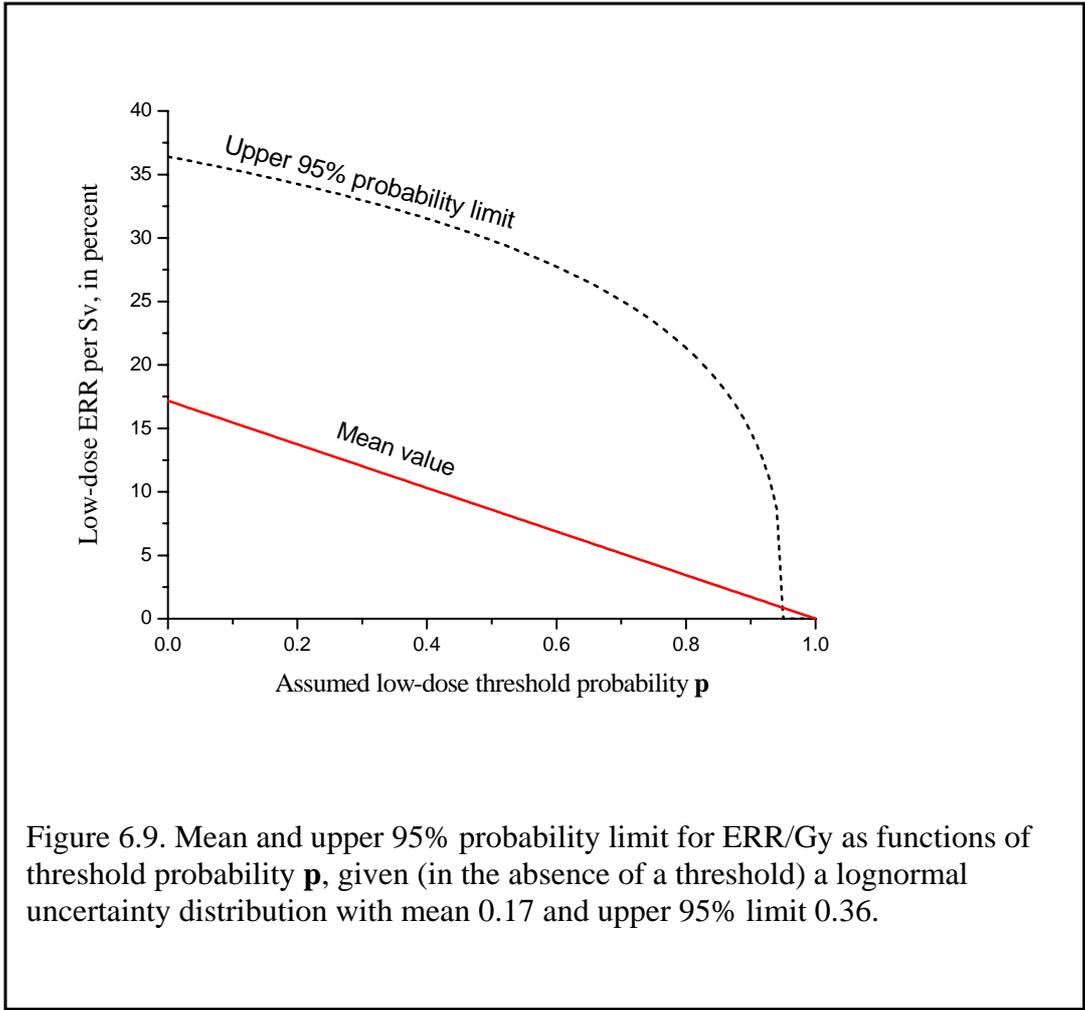
NCRP (1997) 11% 27%

Grogan et al (2000) 12% 28%

NCI/CDC (2003) 17% 36%



ERR/Gy at low doses

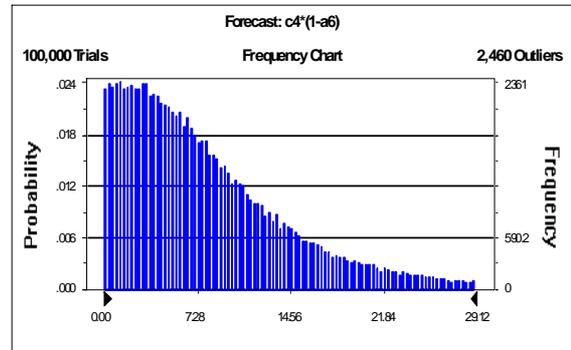
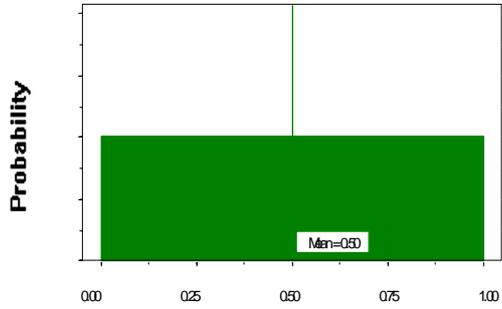


1 Figure 6.10. Effect of uncertain threshold probability on the uncertain distribution for
 2 low-dose ERR/Gy.

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Assumed uncertainty distribution for p

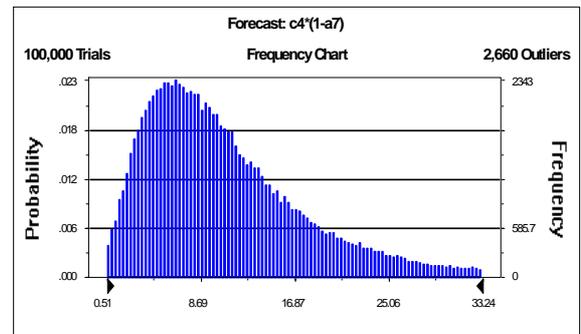
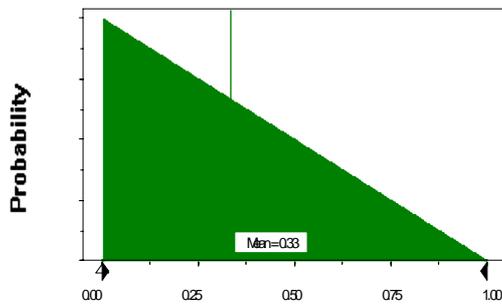
Resulting distribution for ERR/Gy



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7 i) For $p \sim U(0,1)$, the mean ERR/Gy is 8.6% and the 95% upper limit is 23%.

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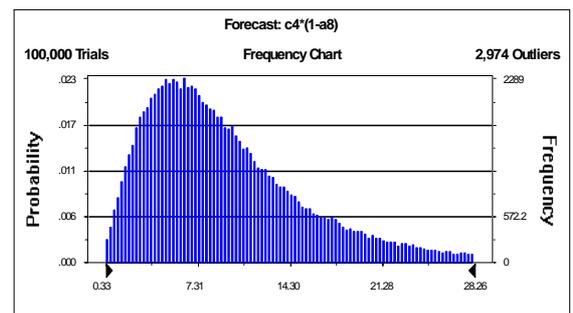
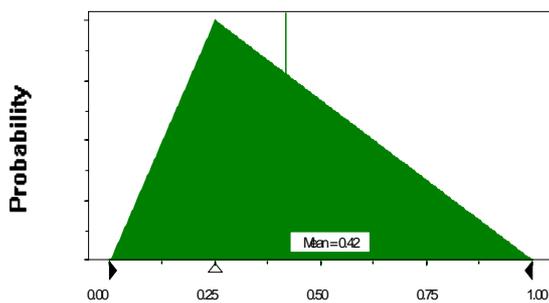
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10 ii) For $p \sim T(0,0,1)$, the mean ERR/Gy is 11.5% and the 95% upper limit is 27%.

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15 iii) For $p \sim T(0,0.25,1)$, the mean ERR/Gy is 10.4% and the 95% upper limit is 24%.

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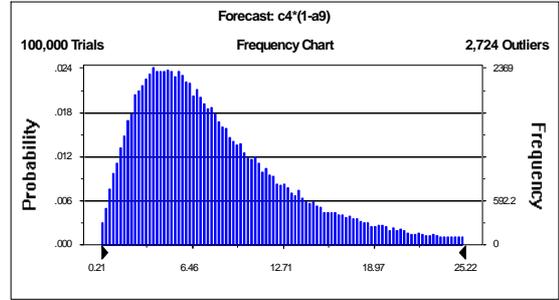
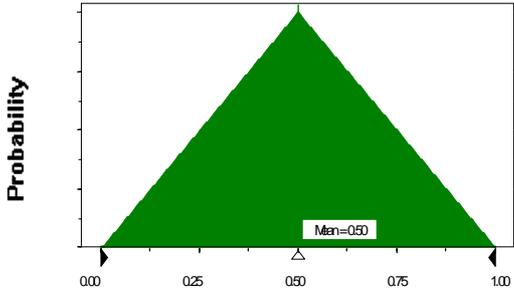
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Figure 6.10 (continued). Effect of uncertain threshold probability on the uncertain distribution for low-dose ERR/Gy.

Assumed uncertainty distribution for p

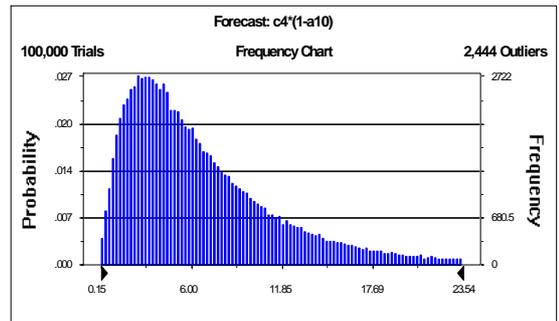
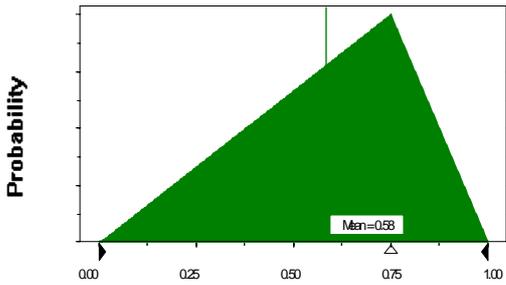
Resulting distribution for ERR/Gy



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9 iv) For $p \sim T(0,0.5,1)$, the mean ERR/Gy is 8.6% and the 95% upper limit is 21%.

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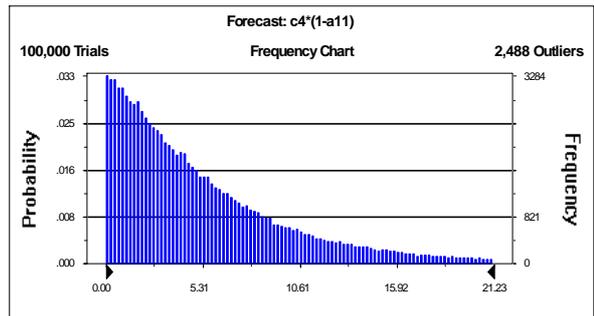
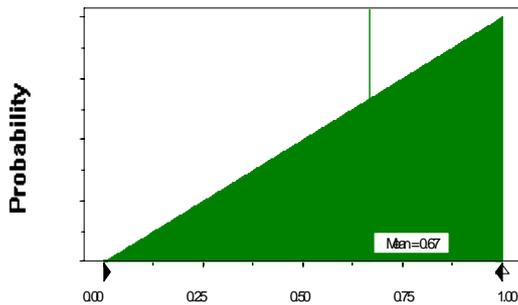


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12 v) For $p \sim T(0,0.75,1)$, the mean ERR/Gy is 7.2% and the 95% upper limit is 19%.

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16 vi) For $p \sim T(0,1,1)$, the mean ERR/Gy is 5.7% and the 95% upper limit is 17%.

